

Impact of climate change environmental conditions on the resilience of different formulations of the biocontrol agent *Candida sake* CPA-1 on grapes

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Running headline: Climate change effect on *C. sake*

Significance and impact of the study: Interacting climate change environmental factors will have a significant impact on food security and availability. Little information exists on how elevated temperature, drought stress and increased CO₂ will have on the efficacy of biocontrol agents. This study examined the impact that such factors may have on the viability of different formulations of the biocontrol yeast *Candida sake* on the surface of grapes for the first time. Such knowledge is critical in evaluating whether biocontrol will remain effective and identify formulations which have the necessary resilience under extreme climate change environmental conditions.

Abstract

Biocontrol agents have become components of integrated crop protection systems for controlling economically important fungal pathogens. *Candida sake* CPA-1 is a biocontrol agent of fungal pathogens of fruits, both pre- and post-harvest. While the efficacy of different formulations have been examined previously, few studies have considered the resilience of different formulations under changing climatic conditions of elevated temperature, drought stress and increased CO₂. This study examined the effect of (a) temperature × RH × elevated CO₂ (400 vs 1000 ppm) on the temporal establishment and viability of two dry and one liquid *C. sake* CPA-1 formulations on grape berry surfaces and (b) temperature stress (25 vs 35 °C) elevated CO₂ concentrations. This showed that temperature, RH and CO₂ concentration influenced the establishment and viability of the formulations but there was no significant difference between formulations. For the three-way climate change environmental factors, the increased temperature (35 °C) and lower RH (40%) reduced the viable populations on grapes. However, the interaction with elevated CO₂ improved the establishment of viable populations of the formulations tested. The viable populations recovered were higher than log 4 CFUs g⁻¹ from the grape surfaces suggesting that these had conserved resilience for control of *Botrytis* rot in grapes.

Keywords: yeast formulations, global warming, climate change, resilience, elevated CO₂, biocontrol, *Botrytis cinerea*.

1. Introduction

Climate change is expected to have profound impacts on agroecosystems and thus food security (Medina et al., 2017). Maximising food production under climate change (CC) conditions will require effective crop protection systems, including biocontrol of fungal pathogens and pests. The Intergovernmental Panel on Climate Change (2014) have suggested that there will be an increase in temperature by between 2-5 °C and that more extreme fluctuations in wet and drought periods will occur coupled with a doubling or tripling of the atmospheric CO₂ from 400 to 800-1200 ppm. Indeed, the EU Green paper has suggested that parts of southern Europe will be hotspots for CC impacts (European Commission 2013). These regions are important for many important agricultural and horticultural crops. It is thus important that biological control agents (BCAs) are able to have the necessary resilience under such environmental stresses. For example, Borisade and Magan (2015) showed that entomogenous fungi used for pest control were less effective under CC scenarios than under existing environmental conditions.

The yeast *Candida sake* CPA-1 is a well-known BCA and its efficacy has been demonstrated against blue mould, grey mould and *Rhizopus* rot on pome fruits (Viñas et al. 1998). CPA-1 has also been effective against *B. cinerea* (Cañamás et al. 2011; Calvo-Garrido et al. 2013a, 2014) and sour rot (Calvo-Garrido et al. 2013b) in grapes. Laboratory scale production of CPA-1 has been optimised (Arévalo 1998; Abadias et al. 2003a) and both liquid (Abadias et al. 2003b; Torres et al., 2003) and solid formulations have been developed (Abadias et al. 2001, 2005; Cañamás et al. 2008; Carbó et al. 2017a). Indeed, recently, two improved formulations of *C. sake* have been developed by the addition of biodegradable coatings using a fluidised-bed spray-drying system, which has resulted in film forming formulations on grapes which have better viability than liquid-based formulations (Carbó et al. 2017a,b).

There has been no attempt to examine the impact of CC environmental factors on the viability of formulations of BCAs for fungal pathogen control. Some studies have examined effects of CC factors on BCAs for pest control (Johns et al. 2003; Diaz et al. 2012; Wang et al. 2014; Reeves et al. 2015). However, these have predominantly examined individual component factors only such as elevated CO₂ or temperature. The interaction between the three factors have been suggested to be very important in examining the effects on fungal plant pathogens and indeed pests (Medina et al. 2014, 2015a,b; Medina et al. 2017; Borisade and Magan 2015).

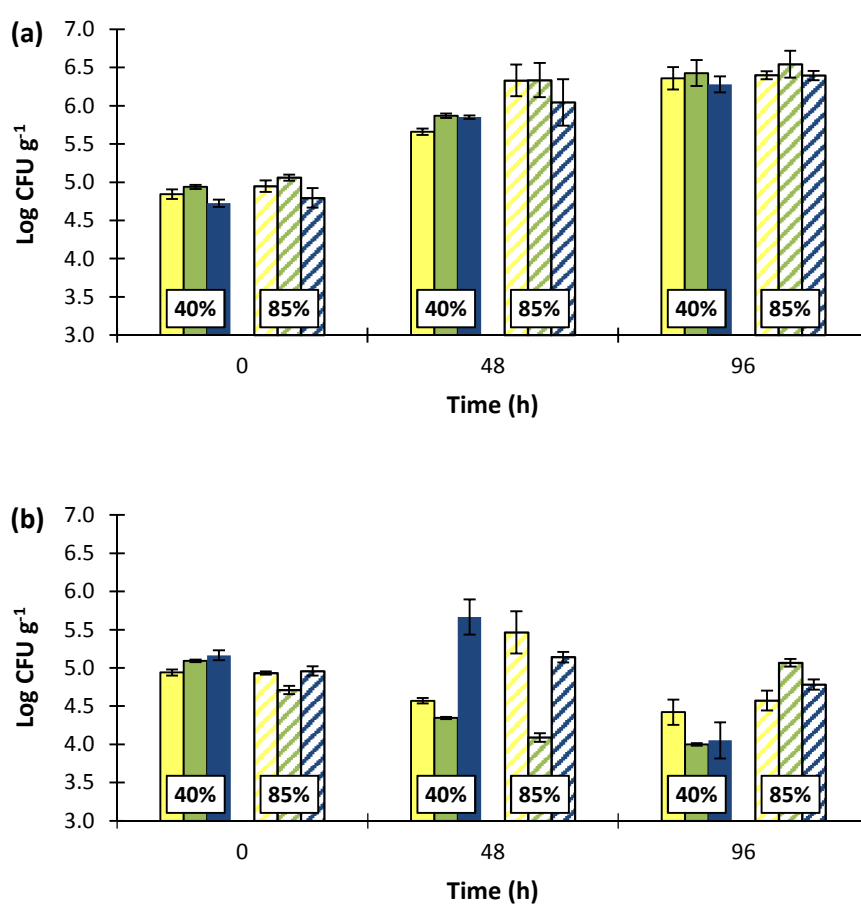
There is thus a dearth of studies on the resilience of formulations of BCAs in the context of tolerance of CC environmental factors. Recently, Carbó et al. (2017b) examined the population dynamics of two fluidised-bed spray-dried formulations in relation to viability on grapes. However, the potential resilience under extreme interacting environmental changes has not previously been examined. Thus, this study examined the effect of interacting CC environmental factors (25 vs 35 °C; 85 vs 40% RH; 400 vs 1000 ppm CO₂) on the resilience of one liquid and two dry formulations of *C. sake* CPA-1 cells in terms of population dynamics on the surface of grapes.

2. Results and discussion

2.1. Effect of interaction climate change environmental factors on population dynamics of *C. sake* formulations

Figure 1 shows the effect of interacting CC factors on the viability of *C. sake* CPA-1 formulations isolated from grape surfaces over a period of 96 h.

Under simulated present conditions (25 °C, 400 ppm; and 40 and 85% RH) all the formulations became established on the grape surfaces (Figure 1a). Indeed, the recovered cells of all three formulations after 48 and 96 h showed an increase at both 40 and 85% RH. From previous ecophysiological studies of *C. sake*, 25 °C was optimum for growth (Teixido et al., 1998). Overall, recovered formulations from grape surfaces showed that more than log 6 CFUs g⁻¹ was established in the control treatment.



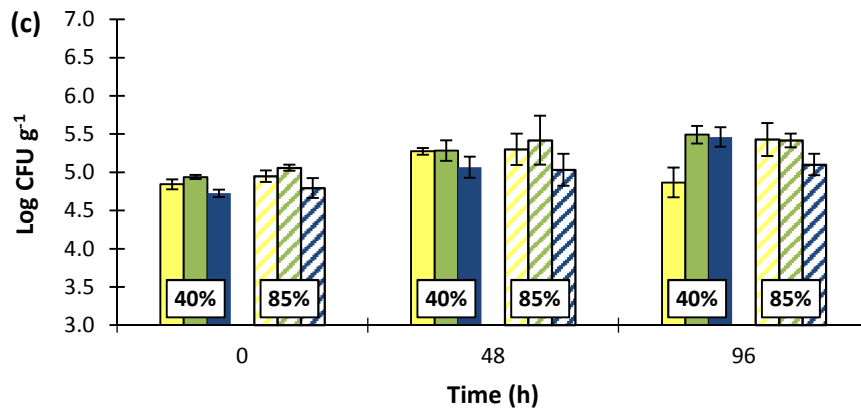


Fig. 1 Dynamics of populations of *C. sake* under the different treatment conditions: (a) current environmental conditions of 25°C and 400 ppm of CO₂; (b) elevated temperature scenario at 35°C and 400 ppm CO₂; and (c) interacting climate change environmental conditions of 35°C and 1000 ppm of CO₂. Key to treatments: Candifruit (■); potato starch formulation (■), and maltodextrin formulation (■) are represented as histograms for the 40% RH (solid colours) and 85% RH (striped bars) conditions. Mean values of three replicates are represented and vertical bars indicated standard error of the means.

Figure 1b shows the impact of increasing the temperature to 35 °C when exposed to existing CO₂ concentrations of 400 ppm at both RH conditions. In the high temperature stress, the recovered populations of each of the formulations generally decreased indicating a lower resilience at this temperature. There was a significant difference between survival of formulations after 48 h. The conditions under which all formulations had the least resilience was after 96 hrs at 35°C and 40% RH (log 0.52-Log1.12). After 96 h at 85% RH the formulations had recovered to between log 4.5-5 CFUs g⁻¹. At 40% RH this was between log 4-4.5 CFUs g⁻¹ in all the formulations. It has been previously shown that *C. sake* cells cannot survive at 37 °C, but were able to grow at 30 °C, although not as well as at 20-25 °C (Teixidó *et al.* 1998). Calvo-Garrido *et al.* (2014a) previously applied *C. sake* cells plus Fungicover® (a commercial coating) to grape berries and observed a decline of Log 5.6 units at extreme temperatures of 40 °C and 100% RH after 72 h and a reduction of Log 2.7 units at 40 °C and 30% RH after 48 h.

At 35 °C, the resilience of the three formulated cell treatments was better when exposed to elevated CO₂ at both RH levels with a similar trend (Figure 1c). The formulations of *C. sake* CPA-1 were recovered from the grape surfaces with viable numbers of at least around log 5 CFUs g⁻¹ and, with the exception of the liquid formulation at 40% RH, all the formulations had increased their populations after 96 h. This suggests that exposure to interacting conditions of elevated temperature × RH and increased CO₂ together resulted in better resilience of formulated cells on grape surfaces than under elevated temperature × RH stress alone. Thus, the CPA-1 formulated yeast cells colonisation at 35 °C was different depending on the CO₂ concentration with better resilience at 1000 ppm of CO₂ than at 400 ppm.

2.2. Effect of temperature on *C. sake* population dynamics (after 96h)

Overall, when examining the relative effect of 25 vs 35 °C there were no significant differences in the populations of *C. sake* cells recovered from the grapes for the three

different formulations tested (Table 1). This suggests that the initial drying temperatures used to make the dry formulations did not influence the resilience of the rehydrated cells on the grapes. Moreover, this also suggests that the fluidised-bed spray-dried formulations were more user-friendly than the liquid formulation, mainly because of the easier downstream handling of the biological product.

However, the interaction between temperature and RH (25 or 35; 40 or 85% RH) had a significant impact on the populations of cells recovered from the grape surfaces. The three-way interaction was not significant (see Table 1). The viable populations recovered from the grapes, regardless of treatment, was significantly better at 25 °C than 35 °C; and also higher in 85% RH than 40% RH. However, the formulations were resilient in the CC conditions examined. Previously, Calvo-Garrido *et al.* (2014) found that formulations of *C. sake* CPA-1 were able to become established and survived under relatively dry Mediterranean climatic conditions when the maximal daily temperature reached 31 °C, and the average minimum daily RH value was 39%.

Table 1 Analysis of variance of effect of formulations, temperature, RH (relative humidity) and two- and three-way interactions on growth of *C. sake* over grapes. Significant sources were itemised and different letters indicate significant differences ($P<0.05$) according to Student's test.

Source	DF	SS	F Ratio	Prob > F
Formulation	2	0.119262	1.1055 NS	0.3480
Temperature	1	31.464600	583.3040 **	<.0001
25 °C ^A				
35 °C ^B				
Formulation × Temperature	2	0.003171	0.0294 NS	0.9711
RH	1	1.236696	22.9264 **	<.0001
85% ^A				
40% ^B				
Formulation × RH	2	0.399328	3.7015 *	0.0404
Potato starch, 85% RH ^A				
Maltodextrin, 85% RH ^{AB}				
Candifruit, 85% RH ^{BC}				
Candifruit, 40% RH ^{BCD}				
Potato starch, 40% RH ^{CD}				
Maltodextrin, 40% RH ^D				
Temperature × RH	1	0.712771	13.2136 *	0.0014
25 °C, 85% RH ^A				
25 °C, 40% RH ^A				
35 °C, 85% RH ^B				
35 °C, 40% RH ^C				
Formulation × Temperature × RH	2	0.289837	2.6866 NS	0.0894

Note: SS, sum of square; * significant $P<0.05$; ** significant $P<0.001$; NS, not significant

In the present study, the *C. sake* populations were recovered from grapes in the highest numbers from the 25 °C treatment. Previously, Teixido *et al.* (1998) in ecological studies

showed that 20-25 °C was optimum for growth of unformulated cells of *C. sake*. With regard to the effect of RH on viability of cells from the formulations, the dry formulations gave better results than the liquid one at 85% RH. However, no differences were observed between formulations at 40% RH (see Table 1).

2.3. Effect of CO₂ concentration on *C. sake* population dynamics on grapes (after 96h)

There was little difference in the viability of the recovered *C. sake* cells from the formulations in the different CO₂ treatments (400 vs 1000 ppm; Table 2). However, because of the significant impact of the high temperature (35°C) on CPA-1 formulations, the effect of CO₂ at this temperature was also examined. This showed that the drying process did not influence the resilience of the rehydrated CPA-1 formulations on the grape surfaces.

Table 2 Analysis of variance of effect of formulations, CO₂, RH (relative humidity) and two- and three-way interactions on growth of *C. sake* over grapes. Significant sources and two-way interactions were itemised and different letters indicate significant differences ($P<0.05$) according to Student's test.

Source	DF	SS	F Ratio	Prob > F
Formulation	2	0.2024381	1.5088 NS	0.2432
RH	1	1.0437026	15.5572 *	0.0007
85% ^A				
40% ^B				
Formulation × RH	2	0.1516857	1.1305 NS	0.3409
CO ₂	1	5.3613231	79.9149 **	<.0001
1000 ppm of CO ₂ ^A				
400 ppm of CO ₂ ^B				
Formulation × CO ₂	2	0.0973929	0.7259 NS	0.4951
RH × CO ₂	1	0.8041026	11.9858 *	0.0022
85% RH, 1000 ppm of CO ₂ ^A				
40% RH, 1000 ppm of CO ₂ ^A				
85% RH, 400 ppm of CO ₂ ^B				
40% RH, 400 ppm of CO ₂ ^C				
Formulation × RH × CO ₂	2	1.1582690	8.6325 *	0.0017

Note: SS, sum of square; * significant $P<0.05$; ** significant $P<0.001$; NS, not significant

Despite this, regardless of the formulation, the viability of CPA-1 was significantly better in the elevated 1000 ppm CO₂ than 400 ppm of CO₂ treatment; and also higher at 85% RH than under water stress conditions. No significant differences were observed in the interaction between formulation and RH (Candifruit, Potato starch or Maltodextrin; 40 or 85% RH), or in the interaction between formulation and CO₂ (Candifruit, Potato starch or Maltodextrin; 400 or 1000 ppm CO₂). However, the interaction between RH and CO₂ (40 or 85%; 400 or 1000 ppm CO₂) had a significant impact on the populations of cells recovered from the grape surfaces. Regardless of the RH, the resilience of the formulations of *C. sake* was better under elevated CO₂ conditions. Also, the three-way interaction among formulation × RH × CO₂ was significant (see Table 2).

Previously, other fungal BCAs such as *Puccinia aprupta* var. *parthenicola* was shown to perform more effectively under increased CO₂ concentrations than existing levels (Shabbir *et al.*, 2014). However, this may vary with BCAs, as CC factors were shown to

have a negative impact on the efficacy of some entomopathogenic fungi for pest control (Borisade and Magan, 2015).

2.4. Estimated capacity of *C. sake* to control *Botrytis* rot under climate change scenarios

Previously, it was shown that populations of at least Log 3 to 5 CFUs g⁻¹ of *C. sake* cells had to be recovered from grape surfaces after the BCA application for effective control of *Botrytis* bunch rot on grapes (Calvo-Garrido *et al.*, 2013a). In the present study, under CC scenarios the three formulations tested would result in the establishment of Log 4.87 to 5.49 CFUs g⁻¹. This would be in the range over which *Botrytis* rot would be effectively controlled (Cañamás *et al.* 2011; Calvo-Garrido *et al.* 2014).

Indeed, even in the high temperature scenario (35 °C), which represented the most stressed condition, the recovered populations after 96 h were almost Log 4 CFUs g⁻¹. Therefore, the three formulations could be effective against *B. cinerea* on grapes under any tested climate scenario. Thus, provided conservation of the resilience of the yeast cells is maintained above the threshold necessary on the berry surface control of *Botrytis* rot would be achieved.

In summary, it has been demonstrated that the formulations of *C. sake* CPA-1 were resilient under the CC conditions examined and the viability of populations would be within the range necessary for *Botrytis* rot control on grapes. Indeed, under elevated CO₂ concentration, more viable cells were isolated from the formulations, even at the elevated temperature, regardless of RH. In addition, the fluidised bed drying process to produce dry formulations of CPA-1 did not significantly affect the resilience of *C. sake* cells on grape surfaces. It may be prudent to examine the relative resilience of different formulations of BCAs to ensure that control levels achieved under existing conditions will be as effective under future CC scenarios. In addition, the ecophysiology and pathogenicity of the pathogen may also change under CC scenarios and this may affect the relative efficacy of formulations of BCAs in the future (Vary *et al.* 2015).

3. Material and methods

3.1. Biocontrol agent and formulations

The yeast strain CPA-1 of *Candida sake* used in this study was obtained from University of Lleida-IRTA, Catalonia, Spain, and it was deposited at the Colección Española de Cultivos Tipo (CECT-10817) at the University of Valencia, Burjassot. *C. sake* stock cultures were stored at 4 °C on nutrient yeast dextrose agar plates (NYDA: nutrient broth, 8 g l⁻¹; yeast extract, 5 g l⁻¹; dextrose, 10 g l⁻¹; and agar, 15 g l⁻¹).

All assays were carried on with three different formulations of the BCA: (i) a liquid formulation registered in Spain under de name Candifruit™; (ii) a dry formulation based on potato starch; and (iii) a dry formulation based on maltodextrin. Both dry formulations were dried using a fluidised-bed spray-drying system by the addition of biodegradable coatings to enhance the survival under environmental stress conditions. The formulation process was done using the protocol described by Carbó *et al.* (2017b).

The number of CFUs ml⁻¹ was determined by plating 100 µl of serial dilutions on NYDA and incubating at 25 °C for 48 h. The viability of formulations was also checked by serial dilutions to calculate the required amount of product to achieve the final concentration of

2.5×10^7 CFUs ml⁻¹. The applied concentration of each treatment was also checked by serial dilutions on NYDA plates.

3.2. Inoculation and incubation conditions

The study was conducted using white seedless grapes washed with tap water to remove possible residues. Afterwards, grape bunches were left to dry in a flow bench and then cut into three-berry clusters leaving the pedicel attached. Three clusters formed one replicate and each treatment consisted of three replicates.

For each treatment, the required amount of formulation was dissolved in 200 ml of water to obtain a concentration of 2.5×10^7 CFUs ml⁻¹. To inhibit bacterial growth, 500 mg l⁻¹ of ampicillin was added to each treatment. Each formulation was placed into a glass beaker and clusters immersed three times into the treatment using sterile forceps, then the clusters were hung on glass rods and allowed to dry at room temperature. When the grape surfaces were dry, each replicate was placed into a glass container and they were all placed in a plastic box and incubated in each climate environmental condition.

3.3. Environmental chamber conditions

Treated grapes were exposed to three different climatic scenarios: (i) the current conditions of 25 °C and 400 ppm CO₂; (ii) elevated temperature of 35 °C and existing CO₂ conditions of 400 ppm and (iii) interacting future climate change scenario of 35 °C and 1000 ppm CO₂. In addition, two relative humidity (RH) conditions were tested for each scenario: (i) 40% and (ii) 85% RH.

When CO₂ concentrations of atmospheric air (400 ppm) were tested, the RH was controlled by introducing 2×500 ml beakers of glycerol/water solution with the same water activity (a_w) as the treatment condition to maintain the equilibrium relative humidity during incubation.

An incubator flushed with the required CO₂ concentration (1000 ppm) was used to simulate the possible climate change scenario. In this situation, the air moisture was controlled by inserting a container with 2 l glycerol/water solution with the same a_w as the treatment condition.

3.4. Evaluation of *C. sake* populations growth on grapes surface

Populations on grape berry surface were recovered after 0, 48 and 72 h. At the recovering time, the three berries of each cluster were separated cutting the pedicels with sterile scissors. The nine berries of each replicate were weighed and then placed into a sterile plastic bag containing 50 ml of sterile distilled water amended with Tween 80 (one drop per litre). Then, the bags were homogenised in a Stomacher 400 (Seward Ltd, Worthing, West Sussex, U.K.) for 10 min. Torres *et al.* (2012) recommended the use of the Stomacher as a consistent and rapid method for recovering the BCA populations from the fruit surface. Serial dilutions were then prepared as described previously to determine the CFUs ml⁻¹, with the results presented as CFUs g⁻¹. All tests were carried out with three replicates and repeated.

3.5. Statistical analyses

The results of CFUs g⁻¹ data were transformed to logarithmic values prior to analyses to improve the homogeneity of variances. Data were analysed by multiple-factor ANOVA using JMP8 software (SAS Institute Inc., NC, U.S.A.). When the analysis was statistically significant ($P < 0.05$), Student's test was used for means separation.

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Conflict of interest

There are no conflicts of interest.

References

- Abadias M., Teixidó N., Usall J., Benabarre A., Viñas I. (2001). Viability, efficacy, and storage stability of freeze-dried biocontrol agent *Candida sake* using different protective and rehydration media. *Journal of Food Protection* **64**, 856–861.
- Abadias M., Teixidó N., Usall J., Solsona C., Viñas I. (2005). Survival of the postharvest biocontrol yeast *Candida sake* CPA-1 after dehydration by spray-drying. *Biocontrol Science and Technology* **15**, 835–846.
- Abadias M., Teixidó N., Usall J., Viñas I. (2003a). Optimization of growth conditions of the postharvest biocontrol agent *Candida sake* CPA-1 in a lab-scale fermenter. *Journal of Applied Microbiology* **95**, 301–9.
- Abadias M., Usall J., Teixidó N., Viñas I. (2003b). Liquid formulation of the postharvest biocontrol agent *Candida sake* CPA-1 in isotonic solutions. *Phytopathology* **93**, 436–442.
- Arévalo S.M. (1998). Optimización de la producción del agente de biocontrol *Candida sake* CPA-1. Universitat de Lleida.
- Borisade O.A., Magan N. (2015). Resilience and relative virulence of strains entomopathogenic fungi under interactions of abiotic stress. *African Journal of Microbiology Research* **9**, 988–1000.
- Calvo-Garrido C., Elmer P., Viñas I., Usall J., Bartra E., Teixidó N. (2013a). Biological control of botrytis bunch rot in organic wine grapes with the yeast antagonist *Candida sake* CPA-1. *Plant Pathology* **62**, 510–519.
- Calvo-Garrido C., Teixidó N., Roudet J., Viñas I., Usall J., Fermaud M. (2014). Biological control of *Botrytis* bunch rot in Atlantic climate vineyards with *Candida sake* CPA-1 and its survival under limiting conditions of temperature and humidity. *Biological Control* **79**, 24–35.
- Calvo-Garrido C., Viñas I., Elmer P., Usall J., Teixidó N. (2013b). *Candida sake* CPA-1 and other biologically based products as potential control strategies to reduce sour rot of grapes. *Letters in Applied Microbiology* **57**, 356–361.
- Carbó A., Torres R., Usall J., Fons E., Teixidó N. (2017a). Dry formulations of the biocontrol agent *Candida sake* CPA-1 using fluidised bed drying to control the main postharvest diseases on fruits. *Journal of the Science of Food and Agriculture*, In press.
- Carbo et al. (2017b)
- Cañamás T.P., Viñas I., Torres R., Usall J., Solsona C., Teixidó N. (2011). Field applications of improved formulations of *Candida sake* CPA-1 for control of *Botrytis cinerea* in grapes. *Biological Control* **56**, 150–158.
- Cañamás T.P., Viñas I., Usall J., Magan N., Solsona C., Teixidó N. (2008). Impact of

- mild heat treatments on induction of thermotolerance in the biocontrol yeast *Candida sake* CPA-1 and viability after spray-drying. *Journal of Applied Microbiology* **104**, 767–775.
- Intergovernmental Panel on Climate Change (IPCC) (2014). Climate change 2014: synthesis report.
- Diaz R., Manrique V., He Z., Overholt W.A. (2012). Effect of elevated CO₂ on tropical soda apple and its biological control agent *Gratiana boliviana* (Coleoptera: Chrysomelidae). *Biocontrol Science and Technology* **22**, 763–776.
- European Commission (2013). Green Paper - A 2030 framework for climate and energy policies.
- Johns C. V., Beaumont L.J., Hughes L. (2003). Effects of elevated CO₂ and temperature on development and consumption rates of *Octotoma championi* and *O. scabripennis* feeding on *Lantana camara*. *Entomologia Experimentalis et Applicata* **108**, 169–178.
- Medina Á., Rodríguez A., Magan N. (2014). Effect of climate change on *Aspergillus flavus* and aflatoxin B1 production. *Frontiers in Microbiology* **5**, 1–7.
- Medina Á., Rodríguez A., Magan N. (2015a). Climate change and mycotoxigenic fungi: impacts on mycotoxin production. *Current Opinion in Food Science* **5**, 99–104.
- Medina Á., Rodríguez A., Sultan Y., Magan N. (2015b). Climate change factors and *Aspergillus flavus*: effects on gene expression, growth and aflatoxin production. *World Mycotoxin Journal* **8**, 171–179.
- Medina, A. Medina, A., Akbar, A., Baazeem, A., Rodriguez, A. & Magan, N. (2017). Climate change, food security and mycotoxins: do we know enough? *Fungal Biology Reviews* **31**, 143-154.
- Reeves J.L., Blumenthal D.M., Kray J.A., Derner J.D. (2015). Increased seed consumption by biological control weevil tempers positive CO₂ effect on invasive plant (*Centaurea diffusa*) fitness. *Biological Control* **84**, 36–43.
- Shabbir A., Dhileepan K., Khan N., Adkins S.W. (2014). Weed-pathogen interactions and elevated CO₂: growth changes in favour of the biological control agent. *Weed Research* **54**, 217–222.
- Teixidó N., Viñas I., Usall J., Sanchis V., Magan N. (1998). Ecophysiological responses of the biocontrol yeast *Candida sake* to water, temperature and pH stress. *Journal of Applied Microbiology* **84**, 192–200.
- Torres R., Usall J., Teixidó N., Abadias M., Viñas I. (2003). Liquid formulation of the biocontrol agent *Candida sake* by modifying water activity or adding protectants. *Journal of Applied Microbiology* **94**, 330–339.
- Torres R., Viñas I., Usall J., Remón D., Teixidó N. (2012). Influence of diluent and sample processing methods on the recovery of the biocontrol agent *Pantoea agglomerans* CPA-2 from different fruit surfaces. *International Journal of Food Microbiology* **158**, 85–88.
- Vary, Z., Mullins, E., Mcelwain, J.C. & Doohan, F. (2015). The severity of wheat diseases increases when plants and pathogens are acclimated to elevated carbon dioxide. *Global Change Biology* **21**, 2661-2669.
- Viñas I., Usall J., Teixidó N., Sanchis V. (1998). Biological control of major postharvest pathogens on apple with *Candida sake*. *International Journal of Food Microbiology* **40**, 9–16.
- Wang G. H., Wang X. X., Sun Y. C., Ge F. (2014). Impacts of elevated CO₂ on *Bemisia tabaci* infesting Bt cotton and its parasitoid *Encarsia formosa*. *Entomologia Experimentalis et Applicata* **152**, 228–237.



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