

Accepted Manuscript

Evaluation of anaerobic digestibility of energy crops and agricultural by-products

A. Spence, E. Blanco Madrigal, R. Patil, Y. Bajón Fernández



PII: S2589-014X(18)30114-2

DOI: <https://doi.org/10.1016/j.biteb.2018.11.004>

Reference: BITEB 113

To appear in: *Bioresource Technology Reports*

Received date: 12 September 2018

Revised date: 10 November 2018

Accepted date: 11 November 2018

Please cite this article as: A. Spence, E. Blanco Madrigal, R. Patil, Y. Bajón Fernández , Evaluation of anaerobic digestibility of energy crops and agricultural by-products. BITEB (2018), <https://doi.org/10.1016/j.biteb.2018.11.004>

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

Evaluation of anaerobic digestibility of energy crops and agricultural by-products

Spence, A.^a, Blanco Madrigal, E.^b, Patil, R.^b, and Bajón Fernández, Y.^{a*}

^aCranfield Water Science Institute, School of Water, Energy and Environment,
Cranfield University, Cranfield, Bedfordshire, MK43 0AL, UK

^bAnaero Technology, Unit 5, Ronald Rolph Court, Cambridge, CB5 8PX

*Corresponding author; y.bajonfernandez@cranfield.ac.uk

Abstract

Abundant wastes from the food and drink supply chain are valuable and infrequently used as anaerobic digestion (AD) substrates. This study quantifies their biomethane potential to contribute to solid waste reduction and energy production. 29 organic materials were evaluated: energy crops (6), pre-treated agricultural by-products (5), livestock slurries (3), agro-industrial wastes (7), fruit and vegetable wastes (4) and co-digestion mixtures of chicken litter (CL) and fruit wastes (4). Results showed highest biogas yields for rendered fat washings (1379 ± 125 mL/g VS_{feedstock}), fish waste (898 ± 107 mL/g VS_{feedstock}) and potato waste (768 ± 27 mL/g VS_{feedstock}). Synergistic benefits of co-digestion were evidenced. CL (20%) with avocado pulp (80%) led to 84% higher biogas than expected from contribution of single substrates.

Keywords

Agricultural by-products; energy crops; co-digestion; livestock manure; anaerobic digestion

1. Introduction

In 2015 the world energy supply was 13,647 million tonnes of oil equivalent and it is predicted to continue increasing due to population growth and advancement of developing countries (IEA, 2017a). However, concern over reserves and over their environmental impact is driving developments on renewable sources of energy. Renewable energy currently accounts for 13.4% of the total world production while petroleum and other liquids, natural gas and coal comprise the majority (IEA, 2017b).

The United Kingdom (UK) used 17,296 thousand tonnes of oil equivalent of renewable fuels in 2016 which accounted for 12% of the country's total energy consumption (UK Department for Business, 2017). The UK continues to progress in terms of using renewable energy sources and have already exceeded their Renewable Energy Directive target (UK Department for Business, 2017). Of the alternative options, anaerobic digestion (AD) plays a key role: it recovers valuable products, such as nutrients, reduces carbon dioxide (CO₂) emissions, and yields renewable energy.

In the past, AD plants typically used sewage sludge to yield biogas but in more recent years the AD industry has enhanced gas production by employing energy crops and food wastes (FW) as a feedstock. Currently, there are over 300 digesters in the UK operating with feedstocks other than sewage sludge (USDA, 2018) and biogas generated in ADs treating substrates other than sewage sludge is predominant in several European countries (EurObserv'er, 2017).

Despite the increase in biogas production from alternative substrates, there is a drive to identify alternative feedstocks to enhance renewable energy production, either by mono digestion of single waste streams or by co-digesting with feedstocks such as FW and sewage sludge (Chiu and Lo, 2016). A promising alternative source is the

agricultural sector, which is responsible for one of the largest waste streams; generating natural wastes such as manure and arable by-products that could be used for renewable energy generation by AD. To illustrate, nearly 64 million tonnes of fish waste is generated annually worldwide, most of which is underutilised as low-value animal feed (Nges et al., 2012). Avocado processing generates ca. 18,144 tonnes of avocado waste annually for Mexico alone (Clarke et al., 2008; Siddiq et al., 2012). Both in the UK and the United States (US) chickens are the most farmed livestock followed by beef cattle and dairy cows (USDA, 2018). All of which constitute examples of waste sources that, if treated by AD, could significantly reduce disposal costs and the amount of uncontrolled methane (CH_4) being released to the atmosphere, with concomitant increases in renewable energy production. There are previous studies available assessing the biodegradability of energy crops and agricultural by-products (Table 1). However, a single study that allows comparison of methane yields and formation kinetics for a wide range of substrates, performed with the same laboratory procedure, is still lacking.

This study investigates the benefits of anaerobically digesting a wide variety of energy crops and agricultural by-products, as single substrates or as co-digestion. 29 different organic materials were selected and its biogas formation potential (yield and kinetics) evaluated: energy crops (6), pre-treated agricultural by-products (5), livestock slurries (3), agro-industrial wastes (7), fruit and vegetable wastes (4) and co-digestion mixtures of chicken litter (CL) and fruit wastes (4).

2. Materials and methods

2.1. Feedstock and inoculum

The inoculum used in all tests was the effluent from a FW AD plant located in Southwest England and treating 80,000 tonnes of FW per year. All substrates were collected from May to July 2017. All energy crops obtained were harvested in their milky stage, manually cut to approximately 2 cm and stored at 4°C until the moment of use. Grass was collected in its young leaf stage. Straw was collected before silage. Untreated wheat straw was manually cut to approximately 2 cm while pre-treated wheat straw was steam treated at 4, 6, 8 and 10 bar with temperatures ranging from 140°C (4 bar) to 180°C (10 bar) using a continuous pressurised refiner (Andritz Sprout-Bauer, Denmark) and then kept frozen until used. CL contained small amounts of bedding (consisting of sand, wood chips and straw bales) and cracked egg shells. The young chick litter, cow manure, onion waste and potato waste were all obtained from the same farm (Table 2). The young chick litter was collected from the tray beneath the cages and contained a small amount of feed. Cow manure was stored outside on the farm and contained wood chips and residual onion from their diet and a small amount of straw from the bedding. Onion waste consisted of peels, onion juice and damaged onions which are used to feed cattle on the farm along with potato waste. Potato waste, originating from potato processing, was stored outdoors while on site and was contaminated with small amounts of onion waste. Onion waste and potato waste were not further processed before being anaerobically digested. Cheese whey originated from blue cheese production. The spent grain was collected from open containers located outside the brewery and waste yeast was collected from a refrigerated, air tight container. Sugar beet tops were processed with a meat mincer to a particle size below 3 mm. All substrates were stored at 4°C since collection until use, with the exception of wheat straw that was frozen. The banana peels (BP) used in

the batch tests were fully ripe and some were flecked with brown. BP were manually cut into approximately 1.5 cm². Fresh potatoes were peeled manually and the peels used for anaerobic digestion without further processing. Avocado seeds were tested for single substrate digestion experiments after being processed in a meat mincer to a particle size below 3 mm.

Waste over-ripe bananas were manually separated from the flesh. BPs, whole bananas (WB) and avocados pulp were processed (< 3mm particle size) through a meat mincer to homogenize the substrates before preparing the co-digestion blends on wet weight basis of 50:50 BP with CL (50:50-BP:CL), 50:50 WB with CL (50:50-WB:CL), 80:20 avocado pulp with CL (80:20-A:CL) and 50:50 avocado pulp with CL (50:50-A:CL) which were then stored at 4°C until further use.

2.2. Batch digestion experiments

Biomethane potential tests were conducted in 1 litre High-density polyethylene reactor bottles (Anaero Technology, Cambridge, UK) with a 700 mL working volume. The reactors were submerged in a water bath and maintained at 38°C in order to operate at mesophilic conditions. An inoculum to substrate VS ratio of 4:1 was used for all tests except soya (6:1), rendered fat washings (6:1) and CL (8:1). Inoculum was diluted for the rendered fat washings and cheese whey experiments (Table 2). BMP equipment (Anaero Technology, Cambridge, UK) normalised gas flow to standard temperature and pressure (STP) conditions (273°K and 1013.25 kPa). All reactors were continuously stirred at 45 rpm by a paddle mixer. A triplicate of inoculum without any feedstock was used in each set of tests as a control to determine the baseline gas production per gram of inoculum VS. All feedstocks were tested in triplicate except for: rye (whole-

crop), soya (whole-crop), and untreated and pre-treated wheat straw which were performed in duplicate due to insufficient sample. Sampling on site was performed once for each substrate, with sub-samples of the collected feedstock used for test replication.

2.3. Analytical methods

All substrates were analysed for total solids (TS) and volatile solids (VS) according to the American Public Health Association (2005). A 1-L Tedlar gas bag was connected to the discharge point of the gas flow meter. Biogas produced in the batch experiments was collected for the first 15 days and composition (CH₄) was analysed once using a Geotech BIOGAS 5000 gas monitor.

Biogas yield expressed in mL/g VS added was calculated as the cumulative volume of biogas produced per g of VS added to the reactor then subtracting the average biogas yield obtained from the triplicate control reactors based on inoculum VS. The termination criterion for ultimate biogas yield was when daily biogas production was less than 5%. The longest time it took for a substrate to reach less than 5% daily biogas production was 20 days. All substrates yields are reported to 20 days.

2.3.1. Kinetic study

Lag phase (λ) was estimated using the modified Gompertz equation (Ghatak et al., 2014) fitted to experimental values and applying the Microsoft Excel Solver tool by performing a non-linear least-square regression analysis (3-1).

$$P = P_{max} \times \exp \left\{ -\exp \left[\frac{R_{max} \times e}{P_{max}} (\lambda - t) + 1 \right] \right\} \quad (\text{Eq. 1})$$

Where, P is the cumulative biogas yield for a given time ($\text{mL/g VS}_{\text{feedstock}}$); P_{max} is the biogas production potential ($\text{mL/g VS}_{\text{feedstock}}$); R_{max} is the ultimate biogas production rate ($\text{mL/g VS}_{\text{feedstock}}/\text{d}$); λ is the lag phase (d); t is time (d) and e is a mathematical constant, $\exp(1)$, ($e=2.178282$). The Gompertz equation was applied as it is commonly reported to provide the highest correlation value for simulating cumulative biogas production in batch digestion as compared to the first order kinetic model.

Analysis of variance (ANOVA) was used to determine statistical significance with a threshold p-value of 0.05.

3. Results

3.1. Biomethane potential tests

3.1.1. Characterisation of inoculum and substrates

The TS and VS of all substrates and inoculum tested in this study are shown in Table 2. Due to the wide range of substrates tested, the VS content of all substrates ranged from 0.2 to 69%, with rendered fat washings being the lowest and pre-treated straw at 6 bar the highest. The inoculum used in all tests had an average TS and VS content of $4.75 \pm 0.9\%$ and $3.09 \pm 1.0\%$, respectively.

3.1.2. Digestion performance: Biogas potential and kinetics

Biogas yield, CH_4 yield and CH_4 content of all substrates are shown in Table 3. Lag phase for each experiment calculated with Gompertz equation are also reported.

a. Energy crops

Digestion of grass obtained the highest biogas yield ($622 \pm 127 \text{ mL/g VS}_{\text{feedstock}}$) within the energy crops tested, achieving its ultimate biogas yield in around 8 days (Figure 1). For ryegrass, triticale, rye, wheat and soya, the cumulative biogas yields were 481 ± 34 ,

510±18, 432±19, 404±38 and 594±31 mL/g VS_{feedstock}, respectively. However, the CH₄ yield of soya was the greatest compared to all other energy crops tested (376±19 mL/g VS_{feedstock}). The CH₄ yields of the remaining energy crops varied between 283 to 359 mL/g VS_{feedstock} (Table 3). It was observed that grass was the only energy crop to have a calculated lag phase of 0 days while triticale had the longest compared to all substrates tested, 6.8 days (Table 3).

b. Pre-treated straw

All pre-treated straw samples showed a similar biogas production rate on days 1 to 3 (Figure 1). However, the untreated straw had a longer lag period (4.9 days) in comparison to all pre-treated straw (4 bar: 3.1, 6 bar: 2.0, 8 bar: 3.0 and 10 bar: 2.9 days) (Table 3). The biogas yields for untreated straw, pre-treated straw at 4, 6, 8 and 10 bar were 326±22, 441±42, 313±17, 426±7 and 356±11 mL/g VS_{feedstock}, respectively. Before day 13 all substrates had produced approximately 80% of their ultimate cumulative biogas yield. It was observed that straw pre-treated at 4 bar had the longest lag phase (3.1 days) compared to other pre-treated samples but resulted in the highest CH₄ yield (271±26 mL/g VS_{feedstock}). In comparison, straw pre-treated at 6 bar had the shortest lag phase (2.0 days) but generated the lowest CH₄ yield (198±11 mL/g VS_{feedstock}). The CH₄ content of all straw digested ranged from 60.4 to 63.2% (Table 3).

c. Livestock slurries

The biogas yields of CL, chick litter and cow manure were 307±24, 371±23 and 123±32 mL/g VS_{feedstock}, respectively. By day 9, chick litter and CL had produced more than twice the amount of biogas that cow manure produced during the 20-day test duration (Figure 1). CH₄ content for all livestock substrates was similar (59.9, 61.8 and 61.6% for CL, chick litter and cow manure, respectively) (Table 3).

d. Agro-industrial waste

Rendered fat washings experienced a lag phase of 5.3 days but had a significantly higher cumulative biogas yield ($1,379 \pm 125$ mL/g VS_{feedstock}) in comparison to all other substrates tested. Its yield was *ca.* 54% more than the next highest yielding substrate, fish waste (898 ± 107 mL/g VS_{feedstock}). Similarly, cheese whey generated a biogas yield of 818 ± 10 mL/g VS_{feedstock}. Comparing the two brewery wastes, spent grain generated a higher biogas yield (644 ± 42 mL/g VS_{feedstock}) than waste yeast (412 ± 33 mL/g VS_{feedstock}) which reached its full biogas potential by day 6. Sugar beet tops produced 95% of its total biogas potential by day 5, earlier than all other agro-industrial substrates tested (Figure 1). A lag phase was observed for potato waste until day 6 and a biogas yield of 768 ± 27 mL/g VS_{feedstock} was recorded. CH₄ content of fish waste, potato waste, sugar beet tops, whey, spent grain and waste yeast was 60.7, 51.7, 65.5, 43.9, 60.5 and 53.3%, respectively. The CH₄ content of whey was the lowest out of all substrates tested although its CH₄ yield was 359 ± 4 mL/g VS_{feedstock}.

e. Fruit and vegetable waste

The avocado seed reached its ultimate biogas potential much earlier than potato peels and banana peels, producing 90% of its total biogas potential by day 7 (Figure 1). Biogas yields (mL/g VS_{feedstock}) were 568 ± 17 , 633 ± 7 , 474 ± 23 and 576 ± 15 for BPs, potato peels, avocado seed and onion waste, respectively. Potato peels exhibited a shorter lag phase in comparison to potato waste (3.7 and 6.2 days, respectively) but generated a lower CH₄ yield (331 ± 3 mL/g VS_{feedstock}). Onion waste performed the highest in terms of CH₄ content, yielding 62.5% and after a lag phase of 6.5 days (Table 3).

f. Co-digestion

Biogas yields of 80:20-A:CL, 50:50-A:CL, 50:50-BP:CL and 50:50-WB:CL were 811 ± 19 , 602 ± 94 , 425 ± 16 and 625 ± 3 mL/g $VS_{\text{feedstock}}$, respectively (Table 3). 80:20-A:CL and 50:50-WB:CL produced the highest CH_4 yields of all co-digested substrates (517 ± 13 and 422 ± 46 mL/g $VS_{\text{feedstock}}$, respectively). 50:50-WB:CL and 50:50-BP:CL exhibited similar initial (2.2 and 1.9 days, respectively) before producing 95% of their ultimate biogas potential before day 5. Both avocado mixtures showed a steady production of cumulative biogas during the 20-day test. CH_4 content of all co-digested mixtures ranged from 63.8 to 67.5%.

4. Discussion

a. Energy crops

Energy crops are recognised for providing substantial CH_4 yields in contrast to sewage sludge or livestock slurries. Many AD plants in the UK operate with one energy crop as a feedstock; commonly maize, whole-crop cereals and grass silage. In general, energy crops have a recalcitrant structure, with high contents of lignin, cellulose and hemicellulose (Croce et al., 2016; Theuretzbacher et al., 2015). Lignin content covers the hemicellulose and cellulose layers preventing contact from enzymes, consequently preventing hydrolysis (Ferreira et al., 2014) and resulting in slow biogas production. This gradual digestion is observed in the batch assays for triticale, wheat and rye which had the longest calculated lag phases (6.8, 5.6 and 3.5 days, respectively). It is probable they contain greater amounts of lignocellulose as compared to ryegrass, grass and soya which all exhibited faster digestion times (2.6, 0.0 and 1.8 days calculated lag phase, respectively). In this study, soya was harvested early before beginning bloom which could have decreased the proportion of lignocellulosic structures allowing for it to be degraded more easily. Cellulose fibres are difficult for bacteria to degrade when they

are covered by lignin which prevents access to the cellulases. Biogas production of lignocellulosic biomass is influenced in several ways such as, biochemical composition, harvesting season and particle size (Amon et al., 2007). No studies have been reported on the digestion of whole-crop soya. However, all other substrates tested were in accordance to yields reported in literature (Table 1).

b. Steam treated straw

Straw is an abundant agricultural by-product consisting of the dry stalks of cereal crops and accounts for approximately half of the crop yield; making it a particularly suitable feedstock for AD (Croce et al., 2016). Due to its lignocellulosic nature pre-treatment methods such as steam are often applied to weaken the structures and increase degradability. Wheat straw is generally composed of 44.3% lignin, 24.5% cellulose and 22.3% hemicellulose (Croce et al., 2016). Utilising steam pre-treatment has been reported to increase biogas and CH₄ yields by altering the chemical composition (Bauer et al., 2014; Theuretzbacher et al., 2015). In this study, the only samples to produce biogas yields significantly higher than untreated straw were pre-treated at 4 and 8 bar and produced approximately 35 and 31%, respectively, more biogas than untreated straw which is similar to other studies that found an increase from using steam pre-treatment (Amon et al., 2007). These yields obtained were in accordance to the range of values reported in literature (Table 1). The slight differences in yields could be attributed to a temperature difference during pre-treatment or possibly the pressure the steam was injected at. Ferreira et al., (2014) found a 19 to 24% increase in CH₄ yield when using steam treated straw in the temperature range of 170 to 220°C which is higher compared to this study (Amon et al., 2007). The higher biogas yields suggest steam pre-treatment increases the biodegradability of lignocellulosic feedstocks. In

contrast, straw treated at 6 bar generated almost 4% less biogas than untreated straw and straw treated at 10 bar only produced approximately 9% more than untreated straw. Theuretzbacher et al., (2015) conducted a study using similar temperatures to this study (140 to 178°C) and concluded that steam treated straw produced similar or lower biogas yields compared to untreated straw (Theuretzbacher et al., 2015).

Although pressure was not reported in Theuretzbacher et al., (2015) study, it was stated the “Economizer SE” was used for pre-treatment which operates between 5 and 8 bar; allowing the assumption that the same pressure was applied to the samples tested. Similarly, their research found the highest CH₄ yield to be from the sample treated at 140°C which is in accordance to results from this study. Other studies have reported that pre-treatment temperature has the greatest impact on biomass composition and hence CH₄ yield (Theuretzbacher et al., 2015). Untreated straw experienced a lag in biogas production until day 5 where it reached a cumulative gas production rate like the pre-treated substrates. The initial lag period indicates steam pre-treatment increases the digestibility of lignocellulosic biomass by breaking down its complex structures increasing its accessibility to bacteria (Theuretzbacher et al., 2015).

The yields observed in this study indicate that higher yields are obtained at lower pressure and temperature (4 bar and 140°C) and when increasing pre-treatment pressure and temperature there is an optimal range to remain within to avoid producing yields lower or similar to untreated straw. These variations in findings challenge the economic feasibility of steam pre-treatment of wheat straw when optimum conditions are not used. As a result, depending on the pre-treatment intensity (temperature and pressure), the pre-treated straw can result in lower yields (Theuretzbacher et al., 2015).

c. Livestock slurries

CL and chick litter both produced higher biogas yields and at a faster rate than cow manure which could be due to their contrasting composition. Niu et al., (2014) reported that CL has a high protein content of 25% TS which is one of the largest contributions to anaerobic fermentation (Niu et al., 2014). In comparison, cow manure has a low protein and lipid content and approximately one-third of its total organic matter is comprised of fibre (Niu et al., 2014). It is probable that the cow manure evaluated in this study also consisted of a high fibre content which prompted its low yield and slow degradation. It has been noted that CL has a higher percentage of readily biodegradable material than cow manure which could explain the difference in initial cumulative biogas production rates (Kelleher et al., 2002). In relation to literature, the CH₄ potential of cow manure (76±20 mL/g VS_{feedstock}) was lower than most values (Table 1). It is plausible that the cow's diet (potato starch, wood chips and onion waste) is responsible for the lower yield. Studies have noted that manure characteristics and biogas potentials are reliant on a series of factors: animal species, diet, protein and fibre content, age, housing and more (Kelleher et al., 2002). Costa et al., (2013) found that cows fed with diets richer in concentrate (protein mixed with maize silage) as opposed to forage had lower levels of fibre and resulted in higher biogas yields in AD. Besides, livestock manure as a feedstock for AD can be precarious as it contains a mixture of feathers or hair, spilled feed, bedding and mortality, which leads to a great variability in reported biogas and CH₄ yields from previous research (Table 1).

d. Agro-industrial waste

The biogas production of fish waste exhibited a calculated lag phase of 3.7 days which can be attributed to the high lipid and protein content in fish waste which takes longer for bacteria to degrade but in turn produces high yields (Costa et al., 2013). Nges et al., (2012) estimated that nearly 64 million tonnes of fish waste is generated annually worldwide, mostly being underutilised as low-value animal feed (Sosa-Hernandez et al., 2016), which evidences the potential for renewable energy formation if treated by AD. Rendered fat washings similarly consists mostly of protein (37%) and lipids (53%) (Hejnfelt and Angelidaki, 2009; Pitk et al., 2012) The measured biogas yield of rendered fat washings was high (1379 ± 125 mL/g $VS_{\text{feedstock}}$) but corresponds to values presented in literature (Table 1). The biogas yield from potato waste (768 ± 27 mL/g $VS_{\text{feedstock}}$) was slightly higher than values in previous studies but the CH_4 content (51.7%) corresponds with reported values (50.8%) (Kryvoruchko et al., 2009). Potato waste generated the fourth highest biogas yield of all substrates tested; a possible explanation for this is its C/N ratio. The C/N ratio of potato waste is approximately 26 which falls within the optimal ratio for biogas production (Kryvoruchko et al., 2009; Mu et al., 2017).

e. Fruit and vegetable waste

Potato peels comprised a relatively low CH_4 content (52.3%) producing an ultimate CH_4 yield of 331 ± 2.5 mL/g $VS_{\text{feedstock}}$. This could be explained by its high starch composition which is easily converted to VFAs and CO_2 but not further to CH_4 (Mu et al., 2017). Previous literature reports CH_4 yields of 239 to 390 mL/g $VS_{\text{feedstock}}$ (Table 41). This study has observed a similar yield in a decreased digestion time of 20 days (Liang and McDonald, 2015; Parawira et al., 2005). Onion waste, BPs and avocado seed exhibited a similar biogas production pattern where gas rapidly increased in the initial days

before slowing down around day 8. A possible explanation is the more easily degraded compounds were consumed in the first 8 days and slow degradation of more complex compounds followed after (Parawira et al., 2004). The biogas yields produced from BP (568 ± 17 mL/g VS_{feedstock}) and onion waste (576 ± 15 mL/g VS_{feedstock}) correspond with values reported in literature (Table 1). To the best of our knowledge, no studies have been reported for avocado seed as a feedstock for AD, which could constitute a viable source of renewable energy as avocado processing generates a significant amount of waste; with approximately 18,144 tonnes of avocado residues generated in 2008 for Mexico alone (Siddiq et al., 2012).

f. Co-digestion

Co-digestion is often recommended when dealing with animal manure to enhance biogas yields, decrease ammonia content and improve the C/N ratio (Akyol et al., 2015). Chicken litter is a high ammonia substrate, with C/N ratios typically around 9 – 13 (Zahan et al., 2018), hence why many studies have been performed co-digesting CL with substrates such as, lignocellulosic biomass, various animal manures, FW and fruit and vegetable wastes. Conversely, to the best of our knowledge no studies have been reported co-digesting CL with banana waste or avocado waste. Both mixtures tested containing ACL reached their ultimate biogas potential on day 19 as compared to day 5 for BP/WBCL due to the high lipid content in avocado pulp which takes longer for bacteria to degrade. In comparison, overripe WBs and BPs are comprised of 16 and 33% monosaccharides, respectively; making them easily degradable at a faster rate (lag phase: 2.2 and 1.9 days, respectively) (Table 3) (Emaga et al., 2011). It is probable the banana mixtures generated higher CH₄ content compared to avocado pulp mixtures (Table 3) due to their high sugar content which a portion is transformed directly to

acetic acid and made available to acetate-utilising methanogens. It is likely to be the reason why 50:50-WBCL (422 ± 46 mL/g VS_{feedstock}) generated a higher CH₄ yield than 50:50-ACL (393 ± 62 mL/g VS_{feedstock}). However, it was observed that the co-digested substrate with the least amount of CL (80:20-ACL) produced the highest CH₄ yield (517 ± 13 mL/g VS_{feedstock}). It is to be remarked that the biogas production of 80:20-ACL and 50:50-ACL was 84% and 54% higher than the expected from their calculated individual contribution as single substrates, suggesting a synergistic effect this co-digestion mixture.

5. Conclusions

All energy crops and agricultural by-products tested can be successfully anaerobically digested to produce biogas. The highest biogas yield (1379 ± 125 mL/g VS_{feedstock}) was obtained from digestion of rendered fat washings. Steam pre-treatment of wheat straw does not guarantee higher biogas yields in comparison to untreated wheat straw, indicating that it may not be economically attractive if pre-treatment conditions are not optimised. Synergistic benefits of co-digesting CL with fruit wastes were evidenced. Co-digesting CL with 50% and 80% avocado pulp led to a 54% and 84% higher biogas production, respectively, than expected from contribution of the separate substrates.

6. Bibliography

1. Akyol, C., Ozbayram, E., Ince, O., Kleinsteuher, S., Ince, B., 2015. Anaerobic co-digestion of cow manure and barley: effect of cow manure to barley ratio on methane production and digestion stability. *Environ. Prog. Sustain. Energy* 35, 589–595.
2. American Public Health Association, 2005. Standard methods for the examination of water and wastewater, 21st ed. American Public Health Association, Washington.
3. Amon, T., Amon, B., Kryvoruchko, V., Machmüller, A., Hopfner-Sixt, K., Bodiroza, V., Hrbek, R., Friedel, J., Pötsch, E., Wagentristsl, H., Schreiner, M., Zollitsch, W., 2007. Methane production through anaerobic digestion of various energy crops grown in sustainable crop rotations. *Bioresour. Technol.* 98, 3204–3212.
4. Antonelli, J., Lindino, A., Rodrigues De Azevedo, J.C., Nelson, S., De Souza, M., Cremonez, P.A., Rossi, E., 2016. Biogas production by the anaerobic digestion of whey. *Produção de biogás por digestão anaeróbia do soro de leite. Rev. Ciências Agrárias* 39, 463–467.
5. Bardiya, N., Somayaji, D., Khanna, S., 1996. Biomethanation of banana peel and pineapple waste. *Bioresour. Technol.* 58, 73–76.
6. Bauer, A., Lizasoain, J., Theuretzbacher, F., Agger, J.W., Rincón, M., Menardo, S., Saylor, M.K., Enguídanos, R., Nielsen, P.J., Potthast, A., Zweckmair, T., Gronauer, A., Horn, S.J., 2014. Steam explosion pretreatment for enhancing biogas production of late harvested hay. *Bioresour. Technol.* 166, 403–410.

7. Bochmann, G., Drosch, B., Fuchs, W., 2015. Anaerobic digestion of thermal pretreated brewers' spent grains. *Environ. Prog. Sustain. Energy* 34, 1092–1096.
8. Chiu, S.L.H., Lo, I.M.C., 2016. Reviewing the anaerobic digestion and co-digestion process of food waste from the perspectives on biogas production performance and environmental impacts. *Environ. Sci. Pollut. Res.* 23, 24435–24450.
9. Clarke, W.P., Radnidge, P., Lai, T.E., Jensen, P.D., Hardin, M.T., 2008. Digestion of waste bananas to generate energy in Australia. *Waste Manag.* 28, 527–533.
10. Costa, M.S.S.M., Costa, L.A.M., De Lucas Junior, J., Pivetta, L.A., 2013. Potentials of biogas production from young bulls manure fed with different diets. *Eng. Agric.* 33, 1090–1098.
11. Croce, S., Wei, Q., D'Imporzano, G., Dong, R., Adani, F., 2016. Anaerobic digestion of straw and corn stover: The effect of biological process optimization and pre-treatment on total bio-methane yield and energy performance. *Biotechnol. Adv.* 34, 1289–1304.
12. Dandikas, V., Heuwinkel, H., Lichti, F., Drewes, J.E., Koch, K., 2014. Correlation between biogas yield and chemical composition of energy crops. *Bioresour. Technol.* 174, 316–320.
13. Dido, M.S., Rieker, C., Bursche, J., Lopez Velarde Santos, M., Hamhaber, J., 2014. Anaerobic Digestion of spent grains: Potential use in small-scale Biogas Digesters in Jos, Nigeria, Anaerobic Digestion of spent grains: Potential use in small-scale Biogas Digesters in Jos, Nigeria.

14. Emaga, T.H., Bindelle, J., Agneesens, R., Buldgen, A., Wathelet, B., Paquot, M., 2011. Ripening influences banana and plantain peels composition and energy content. *Trop. Anim. Health Prod.* 43, 171–177.
15. EurObserv'er, 2017. Biogas barometer [WWW Document]. Biogas Barom. 2017. URL <https://www.eurobserv-er.org/category/all-biogas-barometers/> (accessed 11.6.18).
16. Fang, C., Boe, K., Angelidaki, I., 2011. Anaerobic co-digestion of by-products from sugar production with cow manure. *Water Res.* 45, 3473–3480.
17. Ferreira, L.C., Nilsen, P.J., Fdz-Polanco, F., Pérez-Elvira, S.I., 2014. Biomethane potential of wheat straw: Influence of particle size, water impregnation and thermal hydrolysis. *Chem. Eng. J.* 242, 254–259.
18. Ghatak M., Mahanta P. Comparison of kinetic models for biogas production rate from saw dust. *International Journal of Research in Engineering and Technology.* 2014; 3(7): 248–254.
19. Heiermann, M., Plöchl, M., Linke, B., Schelle, H., Herrmann, C., 2009. Biogas Crops - Part I : Specifications and Suitability of Field Crops for Anaerobic Digestion. *Agric. Eng. Int. CIGR Ejournal* XI.
20. Hejnfelt, A., Angelidaki, I., 2009. Anaerobic digestion of slaughterhouse by-products. *Biomass and Bioenergy* 33, 1046–1054.
21. Hübner, M., Oechsner, H., Koch, S., Seggl, A., Hrenn, H., Schmiedchen, B., Wilde, P., Miedaner, T., 2011. Impact of genotype, harvest time and chemical composition on the methane yield of winter rye for biogas production. *Biomass and Bioenergy* 35, 4316–4323.

22. IEA (International Energy Agency), 2017a. World energy balances: overview. Paris.
23. IEA (International Energy Agency), 2017b. Renewables information: Overview, IEA Statistics. Paris.
24. Kelleher, B.P., Leahy, J.J., Henihan, a M., O'Dwyer, T.F., Sutton, D., Leahy, M.J., 2002. Advances in poultry litter disposal technology--a review. *Bioresour. Technol.* 83, 27–36.
25. Kryvoruchko, V., Machmüller, A., Bodiroza, V., Amon, B., Amon, T., 2009. Anaerobic digestion of by-products of sugar beet and starch potato processing. *Biomass and Bioenergy* 33, 620–627.
26. Lehtomäki, A., Huttunen, S., Rintala, J.A., 2007. Laboratory investigations on co-digestion of energy crops and crop residues with cow manure for methane production: Effect of crop to manure ratio. *Resour. Conserv. Recycl.* 51, 591–609.
27. Li, Y., Zhang, R., Chen, C., Liu, G., He, Y., Liu, X., 2013. Biogas production from co-digestion of corn stover and chicken manure under anaerobic wet, hemi-solid, and solid state conditions. *Bioresour. Technol.* 149, 406–412.
28. Liang, S., McDonald, A.G., 2015. Anaerobic digestion of pre-fermented potato peel wastes for methane production. *Waste Manag.* 46, 197–200.
29. Mshandete, A., Kivaisi, A., Rubindamayugi, M., Mattiasson, B., 2004. Anaerobic batch co-digestion of sisal pulp and fish wastes. *Bioresour. Technol.* 95, 19–24.
30. Mu, H., Zhao, C., Zhao, Y., Li, Y., Hua, D., Zhang, X., Xu, H., 2017. Enhanced methane production by semi-continuous mesophilic co-digestion of potato waste and cabbage

- waste: Performance and microbial characteristics analysis. *Bioresour. Technol.* 236, 68–76.
31. Nathoa, C., Sirisukpoca, U., Pisutpaisal, N., 2014. Production of hydrogen and methane from banana peel by two phase anaerobic fermentation, in: *Energy Procedia*. pp. 702–710.
32. Nges, I.A., Mbatia, B., Björnsson, L., 2012. Improved utilization of fish waste by anaerobic digestion following omega-3 fatty acids extraction. *J. Environ. Manage.* 110, 159–165.
33. Niu, Q., Hojo, T., Qiao, W., Qiang, H., Li, Y.Y., 2014. Characterization of methanogenesis, acidogenesis and hydrolysis in thermophilic methane fermentation of chicken manure. *Chem. Eng. J.* 244, 587–596.
34. Panjičko, M., Zupančič, G.D., Zelić, B., 2015. Anaerobic biodegradation of raw and pre-treated brewery spent grain utilizing solid state anaerobic digestion. *Acta Chim. Slov.* 62, 818–827.
35. Parawira, W., Murto, M., Read, J.S., Mattiasson, B., 2005. Profile of hydrolases and biogas production during two-stage mesophilic anaerobic digestion of solid potato waste. *Process Biochem.* 40, 2945–2952.
36. Parawira, W., Murto, M., Zvauya, R., Mattiasson, B., 2004. Anaerobic batch digestion of solid potato waste alone and in combination with sugar beet leaves. *Renew. Energy* 29, 1811–1823.
37. Pitk, P., Kaparaju, P., Vilu, R., 2012. Methane potential of sterilized solid slaughterhouse wastes. *Bioresour. Technol.* 116, 42–46.

38. Poulsen, T.G., Adelard, L., 2016. Improving biogas quality and methane yield via co-digestion of agricultural and urban biomass wastes. *Waste Manag.* 54, 118–125.
39. Rincón, B., Heaven, S., Banks, C.J., Zhang, Y., 2012. Anaerobic digestion of whole-crop winter wheat silage for renewable energy production. *Energy and Fuels* 26, 2357–2364.
40. Romano, R.T., Zhang, R., 2011. Anaerobic digestion of onion residuals using a mesophilic Anaerobic Phased Solids Digester. *Biomass and Bioenergy* 35, 4174–4179.
41. Siddiq, M., Ahmed, J., Gloria Lobo, M., Ozadali, F., 2012. Tropical and subtropical fruits: postharvest physiology, processing and packaging. Wiley-Blackwell, Ames, USA.
42. Skripsts, E., Dubrovskis, V., Jasko, J., Zabarovskis, E., Kotelenecs, V., 2011. Investigation of biogas production of cheese whey in processing with ozone before anaerobic digestion, in: 10th International Scientific Conference Engineering for Rural Development.
43. Sosa-Hernandez, O., Parameswaran, P., Aleman-Nava, G.S., Torres, C.I., Parra-Saldi, Var, R., 2016. Evaluating biochemical methane production from brewer's spent yeast. *J. Ind. Microbiol. Biotechnol.* 43, 1195–1204.
44. Theuretzbacher, F., Lizasoain, J., Lefever, C., Saylor, M.K., Enguidanos, R., Weran, N., Gronauer, A., Bauer, A., 2015. Steam explosion pretreatment of wheat straw to improve methane yields: Investigation of the degradation kinetics of structural compounds during anaerobic digestion. *Bioresour. Technol.* 179, 299–305.

45. UK Department for Business, E.& I.S., 2017. Digest of UK Energy Statistics (DUKES): renewable sources of energy.
46. USDA, 2018. United States Department of Agriculture. Economics, Statistics and Market Information System [WWW Document]. URL <http://usda.mannlib.cornell.edu/MannUsda/homepage.do> (accessed 6.5.18).
47. Vítěz, T., Koutný, T., Geršl, M., Kudělka, J., Nitayapat, N., Ryant, P., Hejduk, S., Lošák, T., Vítězová, M., Mareček, J., 2015. Biogas and methane yield from rye grass. *Acta Univ. Agric. Silvic. Mendelianae Brun.* 63, 143–146.
48. Wang, X., Yang, G., Feng, Y., Ren, G., Han, X., 2012. Optimizing feeding composition and carbon-nitrogen ratios for improved methane yield during anaerobic co-digestion of dairy, chicken manure and wheat straw. *Bioresour. Technol.* 120, 78–83.
49. Zahan, Z., Georgiou, S., Muster, T.H., Othman, M.Z., 2018. Semi-continuous anaerobic co-digestion of chicken litter with agricultural and food wastes: A case study on the effect of carbon/nitrogen ratio, substrates mixing ratio and organic loading. *Bioresour. Technol.* 270, 245–254.
50. Zhang, T., Yang, Y., Liu, L., Han, Y., Ren, G., Yang, G., 2014. Improved biogas production from chicken manure anaerobic digestion using cereal residues as co-substrates. *Energy and Fuels* 28, 2490–2495.

7. Figure captions

- Fig. 1 - Cumulative biogas yield for a) energy crops, b) untreated and steam pre-treated wheat straw, c) livestock slurries, d) agro-industrial wastes, e) fruit and vegetable wastes and f) co-digested substrates.
- Table 1 – Comparison of biogas and methane yields from literature for substrates tested in BMP tests.
- Table 2 – Characteristics of substrates tested prior to anaerobic digestion tests.
- Table 3 – Biogas and methane yields with calculated lag phases using the modified Gompertz model for all substrates

Table 1 – Comparison of biogas and methane yields from literature for substrates tested in BMP tests.

Feedstock	Biogas yield mL/g VS _{added}	Methane yield mL/g VS _{added}	Digestion time days	Temperature °C	TS % w/w	VS % w/w	Reference
Energy crops							
Ryegrass	431	249	25	38	15.2	13.2	Vitez et al. 2015
	654	399	25	38	25.1	21.9	
	621	318	35	37	35.4	33.3	Hubner et al. 2011
Grass	520	267	30	39	91.2	85.2	Dandikas et al. 2014
	370	228	70	55	18.9	15.3	Poulsen and Adelard 2016
	650	329	30	39	91.2	85.3	Dandikas et al. 2014
Triticale (whole crop)	661	388.0	28	35	33.7	32.0	Heiermann et al. 2009
	614	356	28	35	29.3	27.4	Heiermann et al. 2009
Rye (whole crop)	668	336	30	39	91.6	86.4	Dandikas et al. 2014
Wheat (whole crop)	679	360	40	35	36.3	34.7	Rinco et al. 2012
Steam pre-treated straw							
Untreated straw	-	245	50	35	89.5	82.1	
Pre-treated 170°C	-	291	50	35	89.5	82.1	Ferreira et al. 2014
Pre-treated 200°C	-	304	50	35	89.5	82.1	
Untreated hay	420	234	30	37.5	87.1	82.0	
Pre-treated 170°C	469	281	30	37.5	35.1	32.6	Bauer et al. 2014
Untreated wheat straw	613	276	40	37.5	89.6	85.4	
Pre-treated 140°C	550	275	40	37.5	24.0	23.0	
Pre-treated 160°C	555	261	40	37.5	22.1	21.3	Theuretzbacher et al. 2015
Pre-treated 178°C	563	270	40	37.5	14.6	14.0	
Livestock slurries							
	496	291	28	37	24.9	19.4	Li et al. 2013
Chicken litter	311	127	30	35	26.8	16.7	Wang et al. 2012
	529	282	60	35	26.8	18.8	Zhang et al. 2014
	-	204	20	35	6.5	5.3	Lehtomaki et al. 2007
Cow manure	276	149	40	37	12.4	11.2	Akrol et al. 2015
Agro-industrial wastes							
Rendered fat washings	-	976	36	37.5	99.0	100	Pilk et al. 2012
	-	562	35	55	99.4	99.2	Hejnifelt and Angelidaki 2009
Fish waste	1104	826	33	37	41.2	35.5	Ivo et al. 2012
	672	390	29	27	32.2	17.8	Mshandete et al. 2004
Cheese whey	270	174	9	32	3.0	2.7	Antonelli et al. 2016
	732	450	8	37.5	4.5	4.0	Skripets et al. 2011
Waste yeast	604	284	20	37	8.2	8.1	Sosa-Hernandez et al. 2016
	770	445	24	38.5	93.3	88.7	Dido et al. 2014
Spent brewer's grain	660	389	67	37	21.1	20.1	Bochmann et al. 2015
	386	229	8	37	25.0	23.4	Panicko et al. 2015
Sugar beet tops	-	520	60	37	11.0	9.2	Parawira et al. 2004
	-	280	25	55	11.0	8.5	Fang et al. 2011
Potato waste	654	332	38	37.5	16.4	15.0	Kryvoruchko et al. 2009
Fruit and vegetable waste							
	535	251	12	37	16.5	14.8	Nathoa et al. 2014
Banana peels	327	180	25	37	10.7	9.3	Bardiva et al. 1996
	-	210	23	38	14.6	14.1	Clarke et al. 2008
Potato peels	373	239	40	35	9.2	7.5	Liang and McDonald 2015
	557	390	50	37	19	16.1	Parawira et al. 2005
Onion waste	560	342	14	35	92.6	89.0	Romano and Zhang 2011

ACCEPTED MANUSCRIPT

Table 2 – Characteristics of substrates tested prior to anaerobic digestion tests.

Substrate	TS % w/w	VS % w/w	VS/TS % w/w	Origin
Energy crops				
Ryegrass	35.18 ± 0.01	33.98 ± 0.01	96.59 ± 0.00	Silage field in SE England
Grass	27.08 ± 0.00	25.01 ± 0.00	92.36 ± 0.00	
Triticale (whole crop)	26.54 ± 0.00	24.89 ± 0.00	93.78 ± 0.00	
Soya (whole crop)	14.13 ± 0.00	11.73 ± 0.00	83.01 ± 0.00	Crop trial plots in East England
Rye (whole crop)	39.59 ± 0.01	37.91 ± 0.01	95.76 ± 0.00	
Wheat (whole crop)	38.20 ± 0.00	36.83 ± 0.00	96.41 ± 0.00	
Pre-treated straw				
Untreated straw	49.56 ± 0.03	47.11 ± 0.03	95.06 ± 0.00	Pilot plant in North Wales
Pre-treated 4 bar	37.82 ± 0.02	35.39 ± 0.02	93.57 ± 0.00	
Pre-treated 6 bar	74.74 ± 0.00	69.06 ± 0.00	92.40 ± 0.00	
Pre-treated 8 bar	49.04 ± 0.01	45.85 ± 0.01	93.50 ± 0.00	
Pre-treated 10 bar	42.03 ± 0.01	39.49 ± 0.01	93.96 ± 0.00	
Livestock slurries				
Chicken litter	38.10 ± 0.02	13.48 ± 0.00	35.38 ± 0.03	Free-range egg farm in SE England
Chick litter	33.97 ± 0.00	29.42 ± 0.00	86.61 ± 0.00	Farm in SE England
Cow manure	30.63 ± 0.01	24.21 ± 0.01	79.04 ± 0.01	
Agro-industrial wastes				
Rendered fat washings	0.28 ± 0.00	0.20 ± 0.00	71.43 ± 0.05	Rendering facility in SE England
Fish waste	15.22 ± 0.00	13.73 ± 0.00	90.21 ± 0.00	Fish processing plant in SE England
Cheese whey	6.32 ± 0.00	5.79 ± 0.00	91.61 ± 0.00	Cheese farm in Norfolk, England
Waste yeast	8.11 ± 0.00	7.24 ± 0.00	89.27 ± 0.00	Brewery in Waterbeach, England
Spent grain	47.13 ± 0.01	44.95 ± 0.01	95.37 ± 0.00	
Sugar beet tops	13.35 ± 0.00	11.32 ± 0.00	84.79 ± 0.01	
Potato waste	33.92 ± 0.00	32.16 ± 0.00	94.81 ± 0.00	Farm in SE England
Fruit and vegetable waste				
Banana peels	9.12 ± 0.00	7.77 ± 0.00	85.20 ± 0.00	Local supermarket in Cambridge, England
Potato peels	21.45 ± 0.01	20.26 ± 0.01	94.45 ± 0.00	
Avocado seed	43.99 ± 0.00	42.40 ± 0.00	96.39 ± 0.00	
Onion waste	13.71 ± 0.00	12.86 ± 0.00	93.80 ± 0.00	Farm in SE England
Co-digestion				
80% Avocado 20% Chicken litter	27.03 ± 0.00	20.28 ± 0.00	75.03 ± 0.01	Market in London and a free-range egg farm in Southeast England
50% Avocado 50% Chicken litter	29.50 ± 0.00	16.99 ± 0.00	57.59 ± 0.00	
50% Banana peels 50% Chicken litter	24.08 ± 0.00	11.67 ± 0.00	48.46 ± 0.00	
50% Whole banana 50% Chicken litter	26.11 ± 0.00	13.00 ± 0.00	49.79 ± 0.01	
Inoculum for batch digestion				
Inoculum	4.75 ± 0.91	3.09 ± 0.96	65.05 ± 6.15	Full-scale AD plant in Southwest England
Diluted inoculum (Rendered fat)	0.37 ± 0.00	0.29 ± 0.00	78.38 ± 0.02	
Diluted inoculum (Cheese whey)	1.47 ± 0.00	0.96 ± 0.00	65.31 ± 0.01	

Note: The inoculum was diluted when testing rendered fat and cheese whey in order to maintain the required inoculum to substrate VS ratio while requiring a lower amount of feedstock sample, which was limited in these cases.

Table 3 – Biogas and methane yields with calculated lag phases using the modified Gompertz model for all substrates

Substrate	Biogas yield mL/g VS _{added}	Methane yield mL/g VS _{added}	Methane content %	λ days	R ²
Energy crops					
Ryegrass	481 ± 34.0	294 ± 20.8	61.1	2.57	1.000
Grass	622 ± 127.4	359 ± 73.5	57.7	0.00	0.996
Triticale (whole crop)	510 ± 18.0	338 ± 11.9	66.3	6.80	1.000
Soya (whole crop)	594 ± 30.7	376 ± 19.4	63.3	1.79	1.000
Rye (whole crop)	432 ± 18.7	-	-	3.50	1.000
Wheat (whole crop)	404 ± 38.2	283 ± 26.8	70.1	5.61	1.000
Pre-treated straw					
Untreated straw	326 ± 22.3	197 ± 13.5	60.4	4.86	1.000
Pre-treated 4 bar	441 ± 41.5	271 ± 25.5	61.5	3.11	1.000
Pre-treated 6 bar	313 ± 17.0	198 ± 10.8	63.2	2.02	1.000
Pre-treated 8 bar	426 ± 6.6	262 ± 4.1	61.5	2.97	1.000
Pre-treated 10 bar	356 ± 10.6	217 ± 6.5	61.0	2.87	1.000
Livestock slurries					
Chicken litter	307 ± 24.3	184 ± 14.0	59.9	2.18	1.000
Chick litter	371 ± 23.3	229 ± 11.5	61.8	1.62	1.000
Cow manure	123 ± 32.3	76 ± 19.9	61.6	2.89	1.000
Agro-industrial wastes					
Rendered fat washings	1379 ± 125.3	-	-	5.27	1.000
Fish waste	898 ± 107.0	545 ± 65.0	60.7	3.69	1.000
Cheese whey	818 ± 9.9	359 ± 4.4	43.9	3.51	1.000
Waste yeast	412 ± 3.3	220 ± 1.8	53.3	0.00	1.000
Spent grain	544 ± 42.1	390 ± 25.5	60.5	2.50	1.000
Sugar beet tops	686 ± 26.3	449 ± 45.8	65.5	1.16	1.000
Potato waste	768 ± 27.3	397 ± 14.1	51.7	6.21	1.000
Fruit and vegetable waste					
Banana peels	568 ± 17.4	-	-	4.27	1.000
Potato peels	633 ± 7.0	331 ± 2.5	52.3	3.66	1.000
Avocado seed	474 ± 23.1	-	-	2.21	1.000
Onion waste	576 ± 15.2	360 ± 9.5	62.5	6.53	1.000
Co-digestion					
80% Avocado 20% Chicken litter	811 ± 19.1	517 ± 12.5	63.8	3.85	1.000
50% Avocado 50% Chicken litter	602 ± 93.7	393 ± 62.0	65.3	4.62	1.000
50% Banana peels 50% Chicken litter	425 ± 15.9	283 ± 10.6	66.6	1.87	1.000
50% Whole banana 50% Chicken litter	625 ± 2.8	422 ± 46.0	67.5	2.16	1.000

The termination criteria for ultimate biogas and CH₄ yield was when daily biogas production was less than 5%. 20 days was the longest time for a substrate to reach the criteria.

Highlights

1. The biomethane potential of 29 different organic substrates was evaluated.
2. Highest biogas recorded for rendered fat washings (1379 ± 125 mL/g VS_{feedstock}).
3. Synergistic benefits of co-digesting CL with fruit wastes were evidenced.
4. Co-digestion of chicken litter with avocado pulp led to synergistic biogas yields.

Conflicts of interest

I declare that there are no conflicts of interest to declare for manuscript BITEB-D-18-00198

ACCEPTED MANUSCRIPT

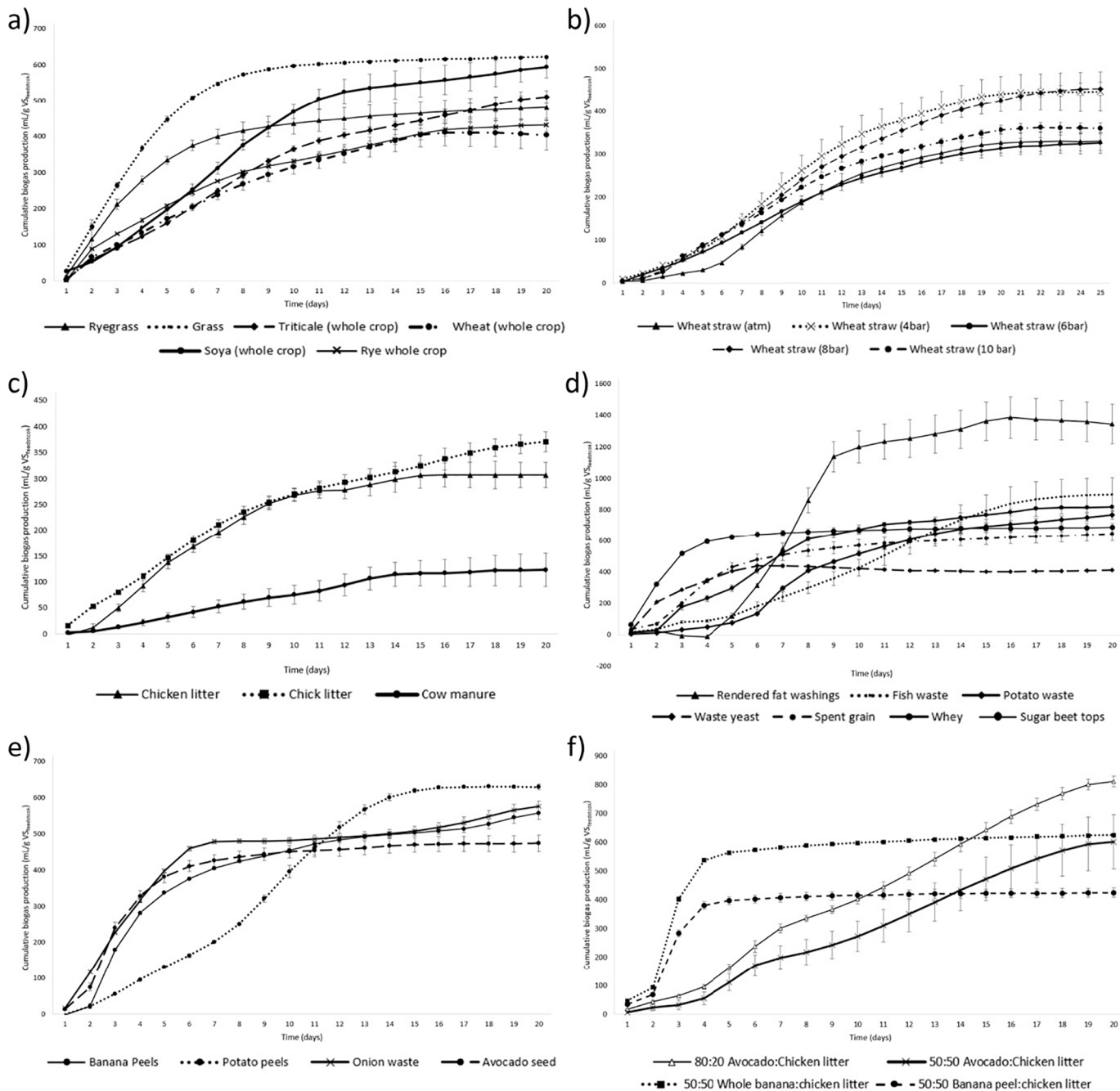


Figure 1



DSPACE

<https://dspace.org/>

Evaluation of anaerobic digestibility of energy crops and agricultural by-products

Spence, A.; Blanco Madrigal, E.; Patil, R.; Bajón Fernández, Yadira

2018-11-16

Elsevier

<https://doi.org/10.1016/j.biteb.2018.11.004>

Spence A, Madrigal EB, Patil R, Bajón Fernández Y. (2019) Evaluation of anaerobic digestibility of energy crops and agricultural by-products. Bioresource Technology Reports, Volume 5, February 2019, pp. 243-250