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Effect of competition between petroleum-degrading bacteria and indigenous compost microorganisms on the efficiency of petroleum sludge bioremediation: field application of mineral-based culture in the composting process

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Abbreviations list[1]

[1]BH, Bushnel-Haas; C_{org}, organic carbon; FC, finished compost ; ICM, indigenous compost microorganisms; MATH, microbial adhesion to hydrocarbon; MBC, mineral-based culture; OD, optical density; PDB, petroleum degrading bacteria; PHCs, petroleum hydrocarbons; PWS, petroleum waste sludge; TPHs, total petroleum hydrocarbons

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Abstract: The effect of competition between isolated petroleum-degrading bacteria (PDB) and indigenous compost microorganisms (ICM) on the efficiency of composting process in bioremediation of petroleum waste sludge (PWS) was investigated. After isolating two native PDB (*Acinetobacter radioresistens* strain KA5 and *Enterobacter hormaechei* strain KA6) from PWS, their ability for growth and crude oil degradation was examined in the mineral-based culture (MBC). Then, the PDB isolate were inoculated into the composting experiments and operated for 12 weeks. The results showed that the PDB degraded 21.65-68.73% of crude oil (1-5%) in the MBC after 7 days. The PDB removed 84.30% of total petroleum hydrocarbon (TPHs) in the composting bioreactor containing the initial TPH level of 20 g kg^{-1}. Removal of petroleum hydrocarbons (PHCs) in the composting experiments proceeded according to the first-order kinetics. The computed values of degradation rate constants and half-lives showed a better performance of the PDB than ICM for TPHs removal. This finding suggests that simultaneous application of the PDB and ICM in the composting reactors resulted in a decline in the effectiveness of the PDB which is due to competition between them. The study also verified that the capability of PDB in degrading PHCs can be successfully scaled-up from MBC to composting process.

Keywords: Petroleum waste sludge; Petroleum-degrading bacteria; Indigenous compost microorganisms; Bioremediation

1. Introduction

Petroleum hydrocarbons (PHCs) contamination is still posing significant environmental and human health concerns around the world due to the release of petroleum waste sludge (PWS) generated from oil refinery plants (Robichaud et al., 2019; Siles and Margesin, 2018). The complex constituents of PWS, including PHCs, ammonia, sulfides, etc., make conventional treatment methods inefficient to meet national and international environmental regulations (Srikanth et al., 2018). Over the years, different physical and chemical strategies have been used, with varying efficacy, to solve the environmental issues of PHCs (Chen et al., 2019). The existing treatment methods suffer from a number of problems such as such as high energy requirement, processes complexity, and incomplete contaminant removal which has led to the search for alternative sustainable treatment technologies (Srikanth et al., 2018; Varjani, 2017). Bioremediation which relies on the use of biological processes has been recognized as an economical, efficient, and environmentally friendly approach to treat petroleum-contaminated environments (Awasthi et al., 2018; Huang et al., 2019). From this point of view, composting process has been frequently deemed for treating PHCs (Ren et al., 2018).

The application of hydrocarbon-degrading bacteria as a consortium could improve the degradation efficacy and promote the bioremediation process (Awasthi et al., 2018; Varjani, 2017). Over the years, several petroleum-degrading bacteria (PDB) from PWS have been isolated, which are able to grow rapidly and degrade various concentrations of PHCs (Awasthi et al., 2018; Demichelis et al., 2017). Although bioremediation of PWS has been greatly advanced in recent years, there are still some challenges when the developed bioremediation technology under laboratory conditions is implemented under real environmental conditions. Thus, one of the most important challenges of bioremediation is the low potential of field application of

culture-based experiments. This may be attributed to unfavorable environmental conditions in the field and competition from other indigenous microorganisms in natural environments such as compost (Awasthi et al., 2018; Li et al., 2019b; Tao et al., 2019). Numerous studies have reported that PDB isolated from contaminated environments, exhibit decreased enzymatic activities as the time progresses because of unfavorable conditions (Awasthi et al., 2018; Demichelis et al., 2017). Sayara et al. (2011) reported that the inoculation of the fungus *T. versicolor* did not increase remediation efficiency since indigenous microorganisms were probably better adapted to the environment. Since the effectiveness of bioremediation is dependent on the survival and activity of the inoculated consortia mainly affected by the competition of PDB with indigenous microorganisms, debate around the benefits of bacterial inoculation and its capacity to increase the microbial degradation of PHCs continues (Jiang et al., 2016; Tao et al., 2019).

Therefore, it is necessary to pay attention to the survival and competition of PDB with the indigenous microorganisms present in petroleum-contaminated environments. Considering this background, the novelty of this work lies in the investigation of the competition between exogenous PDB and indigenous compost microorganisms (ICM) during the field application of a mineral-based culture (MBC) in a composting process for bioremediation of PWS. Hence, this study was performed to: (i) isolate PDB with high degradation potential for PHCs and bioaugmentation; (ii) investigate the possible competition between the inoculated consortia and ICM during PWS composting; and (iii) assess the field application of MBC in PWS bioremediation through bioaugmentated composting process.

2. Materials and methods

2.1. Collection and preparation of the PWS and finished compost (FC) Samples

The PWS, collected from an oil refinery plant in Iran, had the following basic characteristics (dry weight basis): TPHs value of 255.05 g kg^{-1}, organic carbon (C_{org}) level of 528.96 g kg^{-1}, moisture content of 27.63%, pH of 6.10, total nitrogen content of 1.75 g kg^{-1}, and total phosphorus content of 1.03 g kg^{-1}. The FC used as a bulking agent was purchased from a local company and passed through a 2-mm sieve to remove sands and other impurities. The initial properties of the FC were as follows (dry weight basis): TPHs background level of 0.79 g kg^{-1}, C_{org} value of 344.85 g kg^{-1}, moisture content of 35.32%, pH of 7.35, total nitrogen content of 4.25 g kg^{-1}, and total phosphorus content of 2.78 g kg^{-1}. The PWS and FC samples were stored in a dark at room temperature before the start of the composting experiments. Some samples of the PWS and FC were steam sterilized at 121 °C for 2 h to eliminate any existing indigenous microorganisms.

2.2. Isolation and identification of the PDB

The PDB used in this research were isolated from the PWS. For PDB isolation, 5 g of the PWS was mixed with 100 ml of mineral Bushnell-Haas (BH) medium. Crude oil (1%) was also added to the medium to be used as a source of energy and carbon. After shaking for 1 week at 30 °C, fresh medium was inoculated with 5 ml of the incubated culture. Following repeating this enrichment procedure for several times, 100 µl of the medium was spread over the nutrient agar surface and incubated. Finally, a total of 24 strains were isolated from the enrichment tests. In order to examine the growth rate and degradation abilities of all the isolated strains, they were added to the mixture of BH and 1% crude oil. Cell growth was determined using optical density (OD) at 600 nm after 2, 4, 7, 10, and 12 days.

A number of tests including citrate, urease, catalase, oxidase, triple sugar iron, H_2S production, nitrate reduction, indole production, and gram staining were conducted for the identification of the strains (Table 1). Molecular identification was performed by the 16S rRNA gene amplification and sequencing according to the standard procedure available in the previous papers (Koolivand et al., 2019b; Poorsoleiman et al., 2019).

2.3. Optimization of culture-based experiments

2.3.1. Impact of pH

To study the effect of pH on crude oil degradation by the PDB, experiments were conducted at different pH values including 4, 5, 6, 7, 8, and 9. HCl and NaOH were used to adjust pH to the appropriate level. The bacterial mixture (0.5 McFarland) was added to BH medium with crude oil (1%) and incubated for 7 days at 30 °C. After the incubation period, both the growth rate and crude oil biodegradation were measured. The biodegradation of crude oil was measured on the basis of the removal rate of TPHs in the culture-based experiments. The removal rates of TPHs were determined as the TPHs removal against the control experiments containing no bacterial addition. The TPHs removal rate was calculated as follows:

$$\text{TPHs removal rate} = [(\text{initial TPHs} - \text{final TPHs})/\text{initial TPHs}] \times 100$$

where the initial TPHs and final TPHs are the amounts of TPHs before and after treatment, respectively.

2.3.2. Impact of initial TPHs concentration

The strains mixture was exposed to the incremental concentrations of crude oil ranging from 1 to 5% so as to determine the highest levels of biodegradation by the PDB. The above-mentioned

amounts of crude oil were mixed with BH solution of 500 ml in the Erlenmeyer flasks. The PDB (1.5×10^8 CFU/ml) was added to them (at initial pH of 7) and incubated at 30 °C. After an incubation period of 7 days at 120 rpm, the TPHs degradation was measured. The crude oil concentration exhibiting the maximum TPHs removal was selected as the optimized level to be used in the composting bioreactors. It is worth pointing out that the control tests with no bacterial addition were also conducted, under the same conditions, in all the above mentioned culture-based experiments.

2.3.3. Measurement of microbial adhesion to hydrocarbon (MATH)

Determination of MATH was performed through cell surface hydrophobicity as described by Chen et al. (2018) with slight modifications. Briefly, after adding the PDB to nutrient agar and incubating for 24 h at 30 °C, one colony was mixed with a buffer solution and the initial concentration of microbial cell was measured in the prepared suspensions. Then, in a clean glass tube, 5 mL of the bacterial suspension was added to 200 µl of hexadecane. The tube was stirred by vortex (2 min) and set aside (30 min) for phase separation. By using a Pasteur pipette, a sample of the suspension was retrieved when the hydrocarbon droplet size was stable. Next, the cell concentration of the retrieved bacterial sample was determined. The MATH was calculated based on the following equation:

$$\text{MATH}\ (\%) = [1 - (OD_1/OD_2)] \times 100$$

where OD_1 and OD_2 are the concentrations of the bacterial suspension before and after mixing, respectively.

2.3.4. Measurement of emulsification activity (E_{24})

Emulsification activity was conducted using the bacterial culture medium and cell-free supernatant according to the procedure published previously (Bayat et al., 2016; Patowary et al., 2017). Briefly, the PDB were blended with nutrient broth, incubated and then the cell-free supernatant was obtained from the cultures centrifugation. After centrifugation, the cell-free supernatant and culture medium were mixed with crude oil of 1% concentration and vortexed vigorously for 2 min. The solution was allowed to stand (24 h) at room temperature. The E_{24} was measured using the following equation:

$$E_{24}\ (\%) = (H_e/H_t) \times 100$$

where H_e (cm) is the emulsified height, and H_t (cm) is the total mixture height. E_{24} was expressed as the percentage of the overall height occupied by the emulsion.

2.4. Composting experiments

2.4.1. Experimental design of the bioreactors

Composting experiments were carried out in the chemical resistant polypropylene containers to evaluate the removal efficiency of the process. Each composting container (5-L capacity) contained 3 Kg of a mixture of PWS and FC. Four conditions were tested as follows:

D_1: autoclaved PWS + autoclaved FC with no PDB inoculation as a control experiment to determine abiotic loss of TPHs

D_2: autoclaved PWS + autoclaved FC + PDB inoculation to evaluate the PDB potential for TPHs degradation

D_3: autoclaved PWS + non-autoclaved FC to investigate the ICM potential for TPHs degradation

D_4: autoclaved PWS + non-autoclaved FC + PDB inoculation for measurement of the combined potential of the PDB and ICM for TPHs degradation to determine the PDB and ICM competition

The initial TPH concentrations in the composting reactors were adjusted at 20 g kg^{-1} based on the findings of the bioremediation tests performed in the mineral-based experiments (section 2.3.2). All the experiments were conducted in triplicate.

2.4.2. Operation of the bioreactors

The mixture of the PWS and FC was thoroughly blended in an attempt to minimize the heterogeneous distribution of TPHs. Each reactor was turned over several times to allow good aeration over the 12-week duration of the process. To enhance aeration and guarantee sufficient oxygen, aeration (at 1 l min^{-1} kg^{-1}) was provided by oil-free pumps (Koolivand et al., 2019c). The nitrogen and phosphorus were balanced to an optimum C/N/P ratio of 100/5/1 (Koolivand et al., 2018). During the composting phase, watering was conducted twice a week to maintain the moisture content in the range of 45-55%. The water was taken from the city of Arak's municipal supply. The reactors D_2 and D_4 were designed for the PDB inoculation with an initial concentration of approximately 1.5×10^8 CFU g^{-1} dry mixture.

2.4.3. Sampling procedure

Seven samples were collected at predetermined intervals (0, 2, 4, 6, 8, 10, and 12 weeks). For each sampling event, three sub-samples were taken from various depths, blended and homogenized into one composite sample for each bioreactor. The homogenized samples were placed in a glass jar and kept at dark and room temperature until analysis.

2.5. Quantification methods

TPHs in the samples was determined following the TNRCC (2001) method. First, 5 gram of composting sample was weighed and dried at 105 °C. Then, 5 ml of n-pentane was added, mixed well, and preserved until the time of analysis. After extraction of PHCs content in the samples, a gas chromatography equipped with a flame ionization detector (GC-FID) (Shimadzu, Japan) was used for quantification of TPHs. The detailed description of the GC-FID method is available in a paper by Koolivand et al. (2013a). After shaking the mixture of composting samples (1 g) with distilled water (10 mL) for 30 min, pH was determined on the aqueous extract (Khadra et al., 2019; Tmecc, 2002). C_{org} was determined by loss on incineration according to the method of TMECC (2002). Moisture readings were taken gravimetrically by oven drying at 105 °C. All the chemicals used in this study were of analytical grade and bought from Sigma-Aldrich Company.

2.6. Data analysis

In the present study, the statistical analyses were performed using Microsoft Excel 2013 and SPSS package (version 11.0). All the tests were performed in triplicate to get reliable data. The obtained results were indicated as means ± standard deviations on the basis of dry weight. Linear regression analysis was applied to show the relation between the variables. Chromas software was used to analyze the sequencing results. The sequences were aligned with the CLUSTAL X 2.0. Similarity searches of the sequences compared to NCBI database were done using BLAST. The phylogenetic trees were drawn using MEGA 7 software.

3. Results and discussion

3.1. Taxonomic and biochemical identification of the PDB

213 The results of NCBI search indicated that the isolated PDB are *Acinetobacter radioresistens*
214 strain KA5 and *Enterobacter hormaechei* strain KA6. The phylogenetic trees for the two strains
215 are shown in Fig. 1 and Fig. 2. The sequences of the strains KA5 and KA6 have been deposited in GenBank (NCBI) under the accession numbers MK127547 and MK127548, respectively. Table 1 also presents the results of the biochemical tests conducted for them.

Fig. 1

Fig. 2

Table 1

3.2. Culture-based experiments

3.2.1. Metabolic properties of the PDB

The capability of the PDB for growth and crude oil consumption in the MBC was examined. The OD_{600} of 0.42, 1.12, 1.67, 1.75, and 1.49 were obtained after the incubation time of 2, 4, 7, 10, and 12 days, respectively. It can be deduced from these values that the growth logarithmic phase of the PDB lasted 7-10 days. For this reason, the 7-day incubation duration was selected for all the experiments performed in the MBC. The values of MATH and E_{24} were calculated to be 23.78 and 31.18%, respectively, showing the possibility of the PDB affinity to PHCs and biosurfactant production.

3.2.2. Impact of pH on crude oil removal

It is necessary to pay attention to pH as a basic parameter for bacterial growth and activity. Any fluctuations in pH can change the solubility and degradation of hydrocarbons (Li et al., 2019a; Muangchinda et al., 2018). Hence, in the current work, the dependency of bacterial growth and crude oil degradation to pH in the MBC was evaluated. This helps to find the best pH

and perform the composting experiment at the optimized pH. The obtained results (Table 2) showed that the PDB had the highest rate (67.46%) of crude oil degradation at pH 7. The growth rate of the PDB and bioremediation rate slightly declined at pHs of 6 and 8, while the removal efficiency dropped dramatically to the values of 43.21 and 45.90% respectively in the pH values 5 and 9. These observations are in line with the results reported by Muangchinda et al. (2018). It has also been reported that the maximum bacterial growth and metabolic activity occurred in the appropriate pH range of 6-8 (Li et al., 2019a; Ma et al., 2016). By using these MBC results for scaling-up bioremediation process, we operated the composting experiments under the neutral pH.

Table 2

3.2.3. Impact of crude oil concentration

The optimization of TPHs level is one of the most determining factors affecting pollutants degradation during bioremediation process. Thereupon, the effect of crude oil concentration was also investigated to scale-up the PDB ability for decomposing PHCs from MBC to composting process. As can be inferred from Table 3, the PDB grew well in BH medium in the presence of 1-5% crude oil. The degradation rates were computed to be 67.46, 68.73, 62.88, 37.54, and 21.65% respectively for the oil amounts of 1, 2, 3, 4, and 5% after the incubation period of 7 days. It comes to no surprise that the removal efficiency went down with the higher initial concentration as the huge amounts of petroleum compounds are toxic to the bacteria. However, in the case of 2% crude oil, the bacterial growth as well as biodegradation effectiveness was higher than that of 1%. The possible reason for this behavior is that a very low concentration of PHCs is not detectable and thereby bacterial activity and bioremediation efficiency are limited (Vaidya et al., 2017; Varjani and Upasani, 2017). Accordingly, it can be stated that the initial

content of crude oil is of utmost importance in performing bioremediation strategies. The findings resulted from the MBC experiments demonstrated that 2% concentration of oil was the optimal level in which the PDB exhibited the highest degradation rate.

Table 3

Two reasons may account for the potential of the PDB for growth and mineralization of the wide range of oil concentrations (1-5%). First, it is because of the inherent metabolic capabilities of the strains KA5 and KA6 isolated from the PWS. The second reason can be attributed to the fact that the synergistic effects between these two different strains makes the overall process more efficient. Since PWS has been made from various hydrocarbons, and each strain metabolizes specific compounds, application of strains mixture yields better results (Kamyabi et al., 2017; Mnif et al., 2015).

Tables 2 and 3 also indicated that the higher biomass production ($OD_{600\ nm}$) of the PDB leads to more biodegradation of crude oil. The regression analysis provided in Fig. 3 showed a strong correlation between biomass production and oil degradation exhibited by the PDB.

Fig. 3

3.3. Field application of the MBC in the composting process

3.3. 1. Degradation of the PWS using biological composting experiments

As numerous factors can impact the TPHs-removal achieved in a bioremediation system, field studies may yield different results than laboratory ones (Robichaud et al., 2019). In order to examine the possibility of field PHCs bioremediation using bioaugmentation, the potential of the PDB in degrading the PWS was determined in the composting treatments. The experimental study of the PWS degradation in the composting reactors was done based on the MBC results. Accordingly, the primary TPHs content in the bioreactors was 20 g kg^{-1} through adjusting the

281 mixing ratios of the PWS to the FC. Adjustment of this mixing ratio as an effective variable of
282 composting process is of importance since it impacts the degradation rate of the target pollutants
283 and also the process cost (Farzadkia et al., 2019; Koolivand et al., 2013b).
284 The variations of TPHs concentration with time in each composting reactor have been shown
285 in Fig. 4. After 12 weeks, TPHs content reduced from the initial level of 20 g kg^{-1} to 18.86, 3.14,
286 12.74, and 5.12 g kg^{-1}, respectively in the experiments D_1, D_2, D_3, and D_4. These amounts
287 correspond to the total TPHs removal of 5.70, 84.30, 36.30, and 74.4%, respectively. The very
288 low level of removal in the control treatment (D_1) demonstrated that TPHs dissipation is mainly
289 due to the metabolic activities of the microbial community present in the composting
290 experiments. Accordingly, the abiotic mechanisms such as volatilization were negligible and
291 were not responsible for the remarkable reduction of TPHs.
292 In the present study, the variation of TPHs content in the composting experiments D_2, and D_4
293 containing the PDB showed an almost similar decreasing trend with time. It can be observed that
294 the most rapid TPHs biodegradation was obtained during the initial 8 weeks of composting,
295 followed by a gradual reduction of biodecomposition rate over the remaining composting time. It
296 has been also reported from the previous works (Koolivand et al., 2019a; Koolivand et al., 2014)
297 that degradation of petroleum compounds during composting proceeds rapidly in the beginning
298 weeks of the process and slows down in the later. A possible reason for this trend is that easily
299 accessible and biodegradable hydrocarbons which can be dissolved in water phase or can be
300 absorbed by the microbial community are degraded first, while the remaining fractions are
301 difficult to degradation (Ren et al., 2018; Robichaud et al., 2019). Furthermore, the generation of
302 petroleum metabolites such as naphthenic acids and fatty acids during the process can limit

303 further biodegradation due to the suppressed degradative activities of the microbial population
304 (Chen et al., 2019; Pacwa-Płociniczak et al., 2019).
305 Fig. 4

306 *3.3.2. Competition between the PDB and ICM*

307 The percentage of TPHs degradation observed in the treatments D_3 (36.30%) indicated limited
308 capabilities of the ICM for biodegradation. In such cases when indigenous degrading-
309 microorganisms are not efficient enough, it is useful to introduce active PDB to enhance the
310 remediation efficacy. As the treatment D_2 was found to have the highest TPHs removal, the high
311 ability of the PDB for biodecomposition of PHCs was verified. Hence, the PDB inoculation was
312 introduced into the treatment D_4 containing the ICM in order to enhance degradation capacity.
313 Although the overall biodegradation efficacy of the reactor D_4 increased due to the PDB
314 addition, it was still lower than that of the D_2. The possible reason may be attributed to the
315 competition between the PDB and ICM. This means that not all inoculated microorganisms are
316 able to be active in contaminated environments due to the competition of the native with
317 nonnative microorganisms for growth and consuming the available sources of carbon and
318 energy. The native species of compost may also act as a barrier to the activities of inoculated
319 nonnative species due to the fact that the inoculated microorganisms may be preyed by some
320 intrinsic species such as protozoa. Moreover, the metabolite of the ICM can inhibit the activity of
321 the PDB since the major populations observed in the mixed culture may be involved in the
322 antagonistic interactions between microbes (Liang et al., 2018; Ren et al., 2018). There are some
323 studies reporting no significant difference in TPHs removal for the experiments with addition of
324 petroleum-degrading microorganisms. Thion et al. (2012) observed antagonistic interactions

between the mixed culture of microorganisms in bioremediation of contaminated soils. Tao et al. (2019) also reported competition between exogenous inoculated and indigenous microorganisms.

Table 4 provides the percentages of TPHs removal in each sampling week. A very important point to make about this Table is that the removal efficiency of the experiment D_4 is not improved greatly until the week 4 which is due to the PDB adaptation and their competition with the ICM. During the week 4-8, the degradation and removal of TPHs enhanced considerably. During this period, the percentage of TPHs removal occurred in the reactor D_4 was by far higher than that of the reactor D_3. This indicated that the PDB isolated from the PSW can effectively survive and adapt to this environment containing the ICM only at this period. Again, the decrease in the ability of the PDB to degrade TPHs over the last 4 weeks of the study could be attributed to the recalcitrant nature of the remaining PHCs. Besides, possible nutrient deficiency and lower levels of the residual TPHs can lead to increased competition between the PDB and ICM.

Table 4

The performed regression analysis of data plotted in Fig. 4 showed that the PHCs biodegradation can be regarded as the first-order kinetics. The biodegradation rate constants calculated for the experiments D_2, D_3, and D_4 were 0.160, 0.042, and 0.132 d^{-1}, respectively. The corresponding half-lives were found to be 4.33, 16.50, and 5.25 days, respectively. The first-order rate constants reported by He et al. (2019) and Gomez and Sartaj (2013) were respectively in the range of 0.0003-0.0049 and 0.004-0.043 d^{-1}. The reason for these differences is due to the fact that the rate constant is dependent on multiple factors such as pollutants nature and concentration, bioremediation method, and characteristics of the microbial population (He et al., 2014; Kulikowska, 2016). The higher biodegradation constant and lower half-life observed in the

reactor D_2 compared to those in the experiment D_3 indicated that the PDB are more efficient than the ICM when being used alone. However, the comparisons of these values with those observed in the reactor D_4 verified the competition between them. In the future, more studies should be directed at competition mechanisms of the inoculated strains with the intrinsic populations.

3.3.3. Competitive impact of FC addition

In the composting of PWS, co-substrates such as FC are added to provide sufficient readily degradable C_{org}. These materials also provide microbial communities and slow-release nutrients that can support the degradation of target pollutants (Chen et al., 2019). Additionally, it acts as a physical support for composting environment, reduces the toxicity of PHCs, and adjust the moisture and aeration of the process so as to promote microbial activity and thereby contributes to effective TPHs degradation (Kästner and Miltner, 2016; Robichaud et al., 2019). However, since microorganisms have a more tendency to readily degradable C_{org} than the resistant petroleum compounds, the excess addition of these materials may compete with the microbial metabolism of PHCs. Therefore, a proper content of FC should be taken into account to balance the competing effect with PWS and motivating effect on microorganisms (Ren et al., 2018). In order to clarify the competing or motivating effect of the FC addition, the evolution of C_{org}/TPHs were presented in Fig. 5a. The increment pattern of C_{org}/TPHs observed in the reactors D_2 and D_4 was an indication of the higher metabolization of PHCs than that of C_{org}. This demonstrated that the FC contents of the composting reactors did not exhibit competing effect with the PWS. It was reported from the two field experiments conducted by Gomez and Sartaj (2013) and Robichaud et al. (2019) that the addition of compost accelerated the removal of target contaminants over the same time span (3 months) of the present study.

370 Fig. 5b showed that the trend of C_{org} change is similar to that of TPHs. Since the base
371 structure of TPHs is C_{org}, there must have been a correlation between TPHs and C_{org}. So as to
372 determine this correlation and related equations, the regression analysis was conducted. It was
373 clear from Fig. 5c that there was a strong linear correlation between C_{org} and TPHs
374 biodegradation. Experiments D_2 and D_4 show very similar correlation results while experiment
375 D_3 is not so efficient in TPHs removal compared to C_{org} removal. The provided equations can be
376 used in the field-scale composting plants for estimating the TPHs degradation rates according to
377 the data of C_{org} removal.

378 Fig. 5

379 **4. Conclusions**

380 The field application of the MBC in the composting process as well as the competition of the
381 PDB with the ICM during PWS bioremediation was investigated. The isolated PDB were
382 capable of degrading crude oil and PWS in the MBC and composting process. The decline in the
383 PDB efficiency, when inoculated into the ICM containing reactor, indicated a competition
384 between the PDB and ICM. However, in the presence of the ICM, the PDB could effectively
385 metabolize TPHs at the weeks 4-8 of the composting time.

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389

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Table titles

Table 1. Biochemical identifications of the two bacterial strains isolated from the petroleum waste sludge

Table 2. Effect of pH (at crude oil concentration of 1%) on the biodegradation rate of crude oil in mineral-based culture after a period of 7 days

Table 3. Effect of initial crude oil concentration (at the pH value of 7) on the biodegradation rate of crude oil in mineral-based culture after a period of 7 days

Table 4. Percentages of TPHs removal in the composting reactors over the process duration

Figure captions

Fig. 1. Phylogenetic tree based on 16S rRNA gene sequences of *Acinetobacter radioresistens* strain KA5 isolated from the petroleum waste sludge

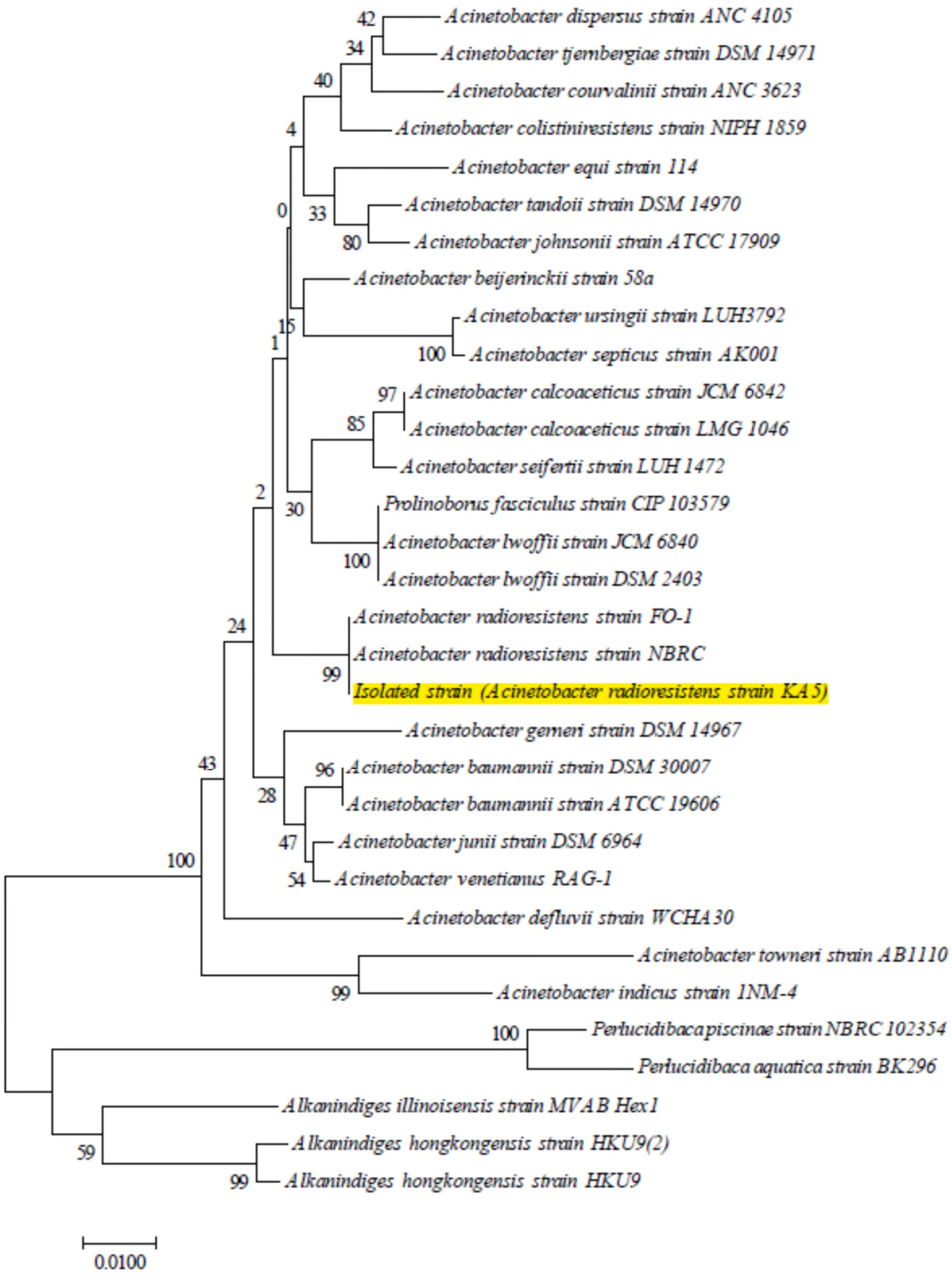
Fig. 2. Phylogenetic tree based on 16S rRNA gene sequences of *Enterobacter hormaechei* strain KA6 isolated from the petroleum waste sludge

Fig. 3. Correlation between bacterial growth and crude oil degradation in the mineral-based culture

Fig. 4. Trend of TPHs degradation in the composting reactors over the process duration (D_1: autoclaved PWS + autoclaved FC with no PDB inoculation as a control experiment; D_2: autoclaved PWS + autoclaved FC + PDB inoculation; D_3: autoclaved PWS + non-autoclaved FC; D_4: autoclaved PWS + non-autoclaved FC + PDB inoculation)

Fig. 5. Trend of (a) C_{org}/TPHs and (b) C_{org} changes in the composting reactors and (c) correlation between TPHs and C_{org} over the process duration (D_1: autoclaved PWS + autoclaved FC with no PDB inoculation as a control experiment; D_2: autoclaved PWS + autoclaved FC +

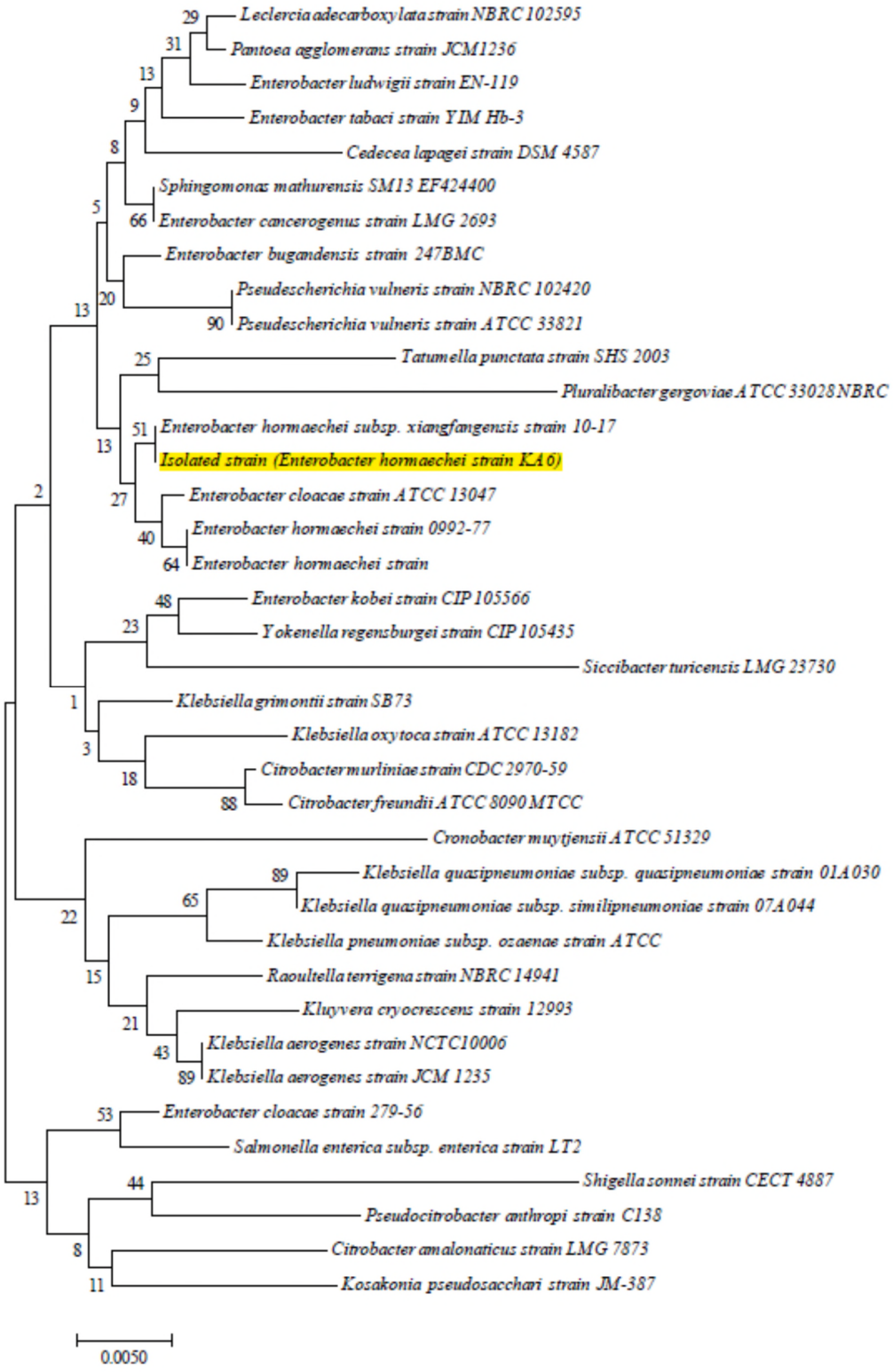
546 PDB inoculation; D_3: autoclaved PWS + non-autoclaved FC; D_4: autoclaved PWS + non-
547 autoclaved FC + PDB inoculation)
548


Acinetobacter dispersus strain ANC 4105
Acinetobacter tjernbergiae strain DSM 14971
Acinetobacter courvalinii strain ANC 3623
Acinetobacter colistiniresistens strain NIPH 1859
Acinetobacter equi strain 114
Acinetobacter tandoii strain DSM 14970
Acinetobacter johnsonii strain ATCC 17909
Acinetobacter beijerinckii strain 58a
Acinetobacter ursingii strain LUH3792
Acinetobacter septicus strain AK001
Acinetobacter calcoaceticus strain JCM 6842
Acinetobacter calcoaceticus strain LMG 1046
Acinetobacter seifertii strain LUH 1472
Prolinoborus fasciculus strain CIP 103579
Acinetobacter lwoffii strain JCM 6840
Acinetobacter lwoffii strain DSM 2403
Acinetobacter radioresistens strain FO-1
Acinetobacter radioresistens strain NBRC
Isolated strain (Acinetobacter radioresistens strain KA5)
Acinetobacter gerneri strain DSM 14967
Acinetobacter baumannii strain DSM 30007
Acinetobacter baumannii strain ATCC 19606
Acinetobacter junii strain DSM 6964
Acinetobacter venetianus RAG-1
Acinetobacter defluvii strain WCHA30
Acinetobacter towneri strain AB1110
Acinetobacter indicus strain 1NM-4
Perlucidibaca piscinae strain NBRC 102354
Perlucidibaca aquatica strain BK296
Alkanindiges illinoisensis strain MVAB Hex1
Alkanindiges hongkongensis strain HKU9(2)
Alkanindiges hongkongensis strain HKU9
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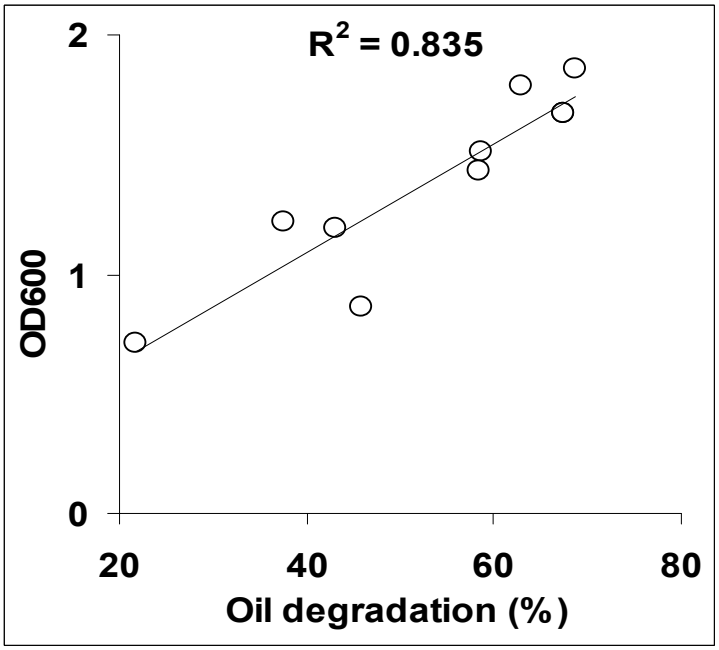
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553 Figure 2

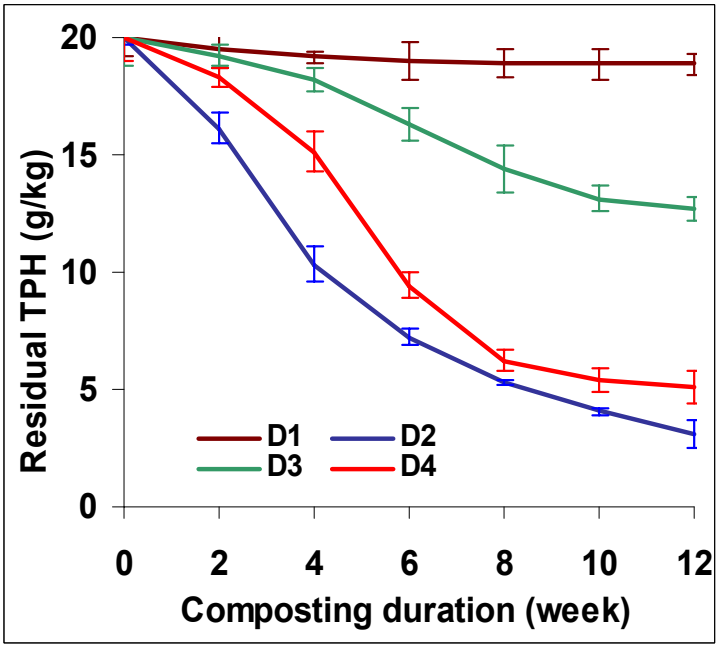
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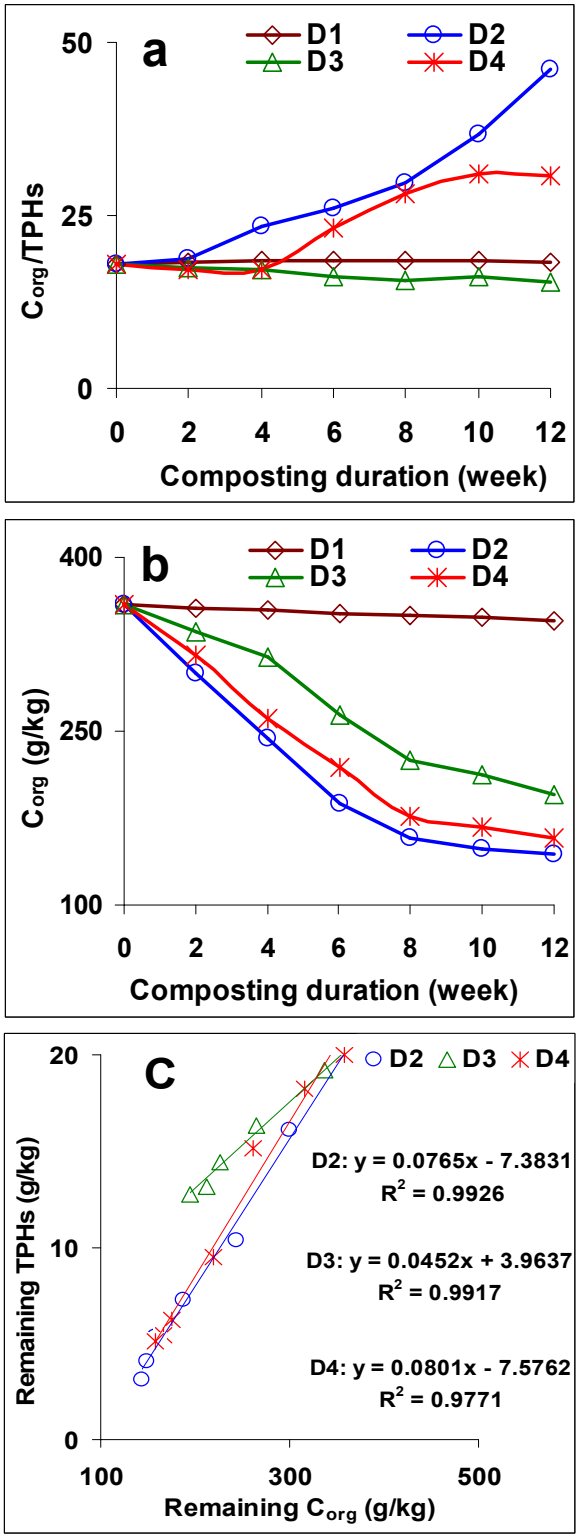
556 Figure 3

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559 Figure 4



560 Figure 5

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562 Table 1

Tests	***Acinetobacter radioresistens* Strain KA5**	***Enterobacter hormaechei* Strain KA6**
Gram stain	Gram negative	Gram negative
Oxidase	–	–
Catalase	+	+
Nitrate reduction	–	+
Citrate	–	+
Urease	–	–
H_2S production	–	–
Indole production	–	–
Triple sugar iron	Alkaline/Alkaline	Acid/Acid

563

564 Table 2

pH	**OD_{600}**	**Crude oil removal (%)**
5	1.19	43.21
6	1.51	58.67
7	1.67	67.46
8	1.43	58.48
9	0.86	45.90

565

566 Table 3

Crude oil concentrations (%)	**OD_{600}**	**Crude oil removal (%)**
1	1.67	67.46
2	1.86	68.73
3	1.79	62.88
4	1.22	37.54
5	0.71	21.65

567

568 Table 4

Process time (week)	TPH removal rate (%)			
	D_1*	D_2**	D_3***	D_4****
0	0.00	0.00	0.00	0.00
2	2.25	19.40	3.95	8.60
4	1.90	28.85	5.05	15.70
6	0.75	15.55	9.40	28.45
8	0.40	9.65	9.60	16.10
10	0.35	6.25	6.30	4.15
12	0.05	4.60	2.00	1.40
Total	5.70	84.30	36.30	74.40

569 * autoclaved PWS + autoclaved FC with no PDB inoculation as a control experiment

570 ** autoclaved PWS + autoclaved FC + PDB inoculation

571 *** autoclaved PWS + non-autoclaved FC

572 **** autoclaved PWS + non-autoclaved FC + PDB inoculation

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Effect of competition between petroleum-degrading bacteria and indigenous compost microorganisms on the efficiency of petroleum sludge bioremediation: field application of mineral-based culture in the composting process

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