



Do temperature and moisture conditions impact soil microbiology and aggregate stability?

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Abstract

Purpose Studies predicting the impacts of climate change on erosion have considered numerous variables, such as rainfall erosivity and vegetation cover, but have not considered potential changes in soil erodibility. Erodibility is an intrinsic property of the soil, strongly correlated with the stability of soil aggregates. It is influenced by soil physico-chemical attributes, including the microbiological community. The study aim was to determine how shifts in temperature and moisture conditions, which other studies have shown affect microbiological communities, might affect aggregate stability.

Methods Using an experimental approach with laboratory microcosms, aggregates from a sandy loam soil and a clay soil were incubated at three temperatures and three moisture conditions in a factorial experimental design. Aggregate stability was quantified using rainfall simulation. Microbiological indicator metrics were measured to evaluate treatment microbiological impacts, including community composition (PLFA), biomass carbon, and respiration.

Results Temperature and moisture content affected aggregate stability significantly, but differently for the two soil types tested. For the sandy loam soil, aggregate stability decreased significantly with increasing moisture content. For the clay soil, aggregate stability increased significantly with increasing temperature. For both soil textures, temperature and moisture content affected microbiological community composition and respiration. Regression analysis indicated that microbiological properties were significant predictors of aggregate stability.

Conclusion Our results emphasise the dynamic nature of soil aggregate stability. Changes in microbiological metrics suggest possible biological mechanisms for aggregate stability changes, which should be investigated further to better understand the potential impacts of climate change on soil erodibility and erosion.

Keywords Soil erosion · Erodibility · Climate change · Microbial community · Biostabilisation

1 Introduction

Climate change is predicted to impact the frequency, severity, and extent of soil erosion, threatening soil sustainability (Nearing et al. 2004; Mullan 2013). Whilst the potential impacts of climate change on rainfall erosivity have been well studied (Nearing 2001), few studies have considered the impacts on soil erodibility, i.e. the susceptibility of soil to erosive forces (Favis-Mortlock and Boardman 1995). Soil erodibility is an intrinsic property of the soil, determined

by the soil properties (e.g. soil texture, organic matter, and biological activity) that influence the forces internal to the soil (e.g. gravity, cohesion, and adhesion) that resist surface erosion. It is represented methodologically as an erosion threshold (i.e. rainfall intensity or overland flow that initiates erosion) or, once the threshold has been surpassed, an erosion rate. Soil erodibility has been found to be strongly correlated inversely with aggregate stability; thus, aggregate stability has been widely used as an indicator of soil erodibility (Bryan 1968; Salvador Sanchis et al. 2008). Aggregate formation and stability are controlled by numerous abiotic and biotic factors that affect the binding of soil particles and aggregates (Six et al. 2004; Bronick and Lal 2005; Regelink et al. 2015). For example, soil microbes can aid aggregation through the physical enmeshment of particles by fungal hyphae (Degens et al. 1996; Lehmann and Rillig 2015) and by the production of exudates (e.g. extracellular polymeric

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substances; EPS) that promote particle adhesion (Tisdall and Oades 1982; Ritz and Young 2004; Blankinship et al. 2016). By working at the aggregate scale, researchers can focus on how soil properties affect erodibility, before scaling up to consider the interactions and feedbacks caused by other environmental or anthropogenic factors important at field scale.

Despite several proposed relationships between seasonal conditions and aggregate stability (e.g. Bullock et al. 1988), there is limited empirical evidence to support the prediction of climate change impacts on aggregate stability. Seasonal variations in aggregate stability have been observed (Lavee et al. 1996; Amézketa 1999; Cosentino et al. 2006; Salvador Sanchis et al. 2008), which suggest that it is responsive to changes in environmental conditions. However, there is a lack of consensus in the literature. Aggregate stability has been observed to both increase and decrease with changes in temperature and moisture content (Deneff et al. 2001; Cosentino et al. 2006). Whilst some of the differences between studies could be explained by different methodologies and soil properties (Amézketa 1999) or climate-associated influences on aggregation creation and breakdown due to freeze-thaw or wetting-drying, it remains unclear how shifts in temperature and soil moisture, individually and in combination, might affect aggregate stability, and, in turn, erodibility.

Previous research has demonstrated that microbiological communities and the exudates they create are highly responsive to physical and environmental conditions (e.g. particle size distribution, soil moisture, light, and temperature) (Cosentino et al. 2006; Knapen et al. 2007; Nunan 2017; Baveye et al. 2018; Walker et al. 2018; Li et al. 2019; König et al. 2020). For example, changes in soil temperature and moisture have been shown to affect the composition of the microbiological community (Evans and Wallenstein 2014; Supramaniam et al. 2016), through a variety of mechanisms (e.g. changes in metabolic activity, available oxygen, resource availability and accessibility, microbial motility, and soil microstructure connectivity) (Franzluebbers 1999; Or et al. 2007b; Moyano et al. 2013). These changes would affect microbiological groups, including fungi, gram-positive bacteria, and gram-negative bacteria, differently due to structural and physiological differences, such as hyphal connectivity to resources and cell wall structure (Nazih et al. 2001; Uhlířová et al. 2005; Schimel et al. 2007). The variation in the effects of climatic conditions on microbiological groups affects ecological strategy and resource allocation and, in turn, modifies hyphal growth and EPS production and characteristics (Roberson and Firestone 1992; Evans and Wallenstein 2014). Thus, shifts in the abundance of fungi, gram-positive, and gram-negative bacteria associated with climatic conditions may have a varying effect upon aggregate stability. Therefore, whilst we know that a changing climate will affect the microbiological community, and microbiological communities and their exudates affect erodibility and aggregate stability, we have not yet explored the response of aggregate stability (Cosentino et al. 2006).

The aim of this study is to determine how shifts in temperature and moisture conditions, which have been shown to affect microbiological communities, affect soil aggregate stability. A laboratory experiment was designed to test the following hypotheses: (i) aggregate stability will be greater under higher temperatures; (ii) aggregate stability will be lowest under the driest soil conditions and greatest at intermediate moisture content; (iii) interaction between temperature and moisture treatments will yield the highest aggregate stability under the warmest and intermediate soil moisture conditions; and (iv) the effects of temperature and moisture on aggregate stability will be influenced by soil texture and duration of climate incubation. This research was inspired by the long history of research on biological aggregation and its effects on erodibility in aquatic sediment, with was a core area of research for Dr. Ian Droppo, for whom this special issue is dedicated.

2 Methods

2.1 Soil collection and preparation

Two surface soils (depth 0–150 mm) were used in this experiment: a sandy loam and a clay from the Silsoe Experimental Farm (Bedfordshire, England, National Grid Reference TL075356/TL075351). The sandy loam soil is from the Bearsted series (6% organic matter, 69% sand, 20% silt, 11% clay), and the clay soil is from the Evesham series (6% organic matter, 42% sand, 15% silt, 44% clay). These soils were selected because they had the same organic content, were managed by the same farmer, and were in close proximity (< 500 m apart), thus would have a similar microbiological species pool, even if community composition varied. The two soils were used because particle size distribution and clay content have been shown to alter numerous physical and chemical soil properties (e.g. porosity, soil pH, hydration characteristics and organic matter and cation interactions), which may affect the responses of the microbial community to temperature and moisture and influence soil aggregation processes (Bronick and Lal 2005; Regelink et al. 2015). On collection, the field moist soil was gently broken apart by hand following planes of least resistance, taking care to avoid compaction and smearing. Soils were then air-dried for 72 h away from direct sunlight and sieved to obtain aggregates with diameters between 2 and 5.6 mm. Any plant material retained was gently removed by hand.

The sieved aggregates (25 g, 2–5.6 mm) were assigned to microcosms, which consisted of a 2-mm aperture mesh (Wondermesh) fitted across a hoop (95 mm in diameter) and enclosed in an air-tight container (185 × 130 × 45 mm, $l \times w \times d$) with a cotton wool wetting bed to facilitate soil moisture content through capillary rise. To minimise the

level of initial disruption and the risk of slaking caused by the methodological addition of moisture, the wetting bed within the microcosms was designed to generate a slow rate of initial wetting. The aggregates were spread across the microcosm mesh in a single layer with minimal contact between aggregates to enable the investigation of single aggregate mechanisms. Nine treatments were prepared for each soil texture in a 3×3 multifactorial design, with three levels of temperature (5 °C, 15 °C, and 30 °C) and three levels of moisture content representing wet, intermediate, and dry conditions. Temperature was controlled by placing the microcosms in environmental chambers, whilst moisture content was controlled by adjusting the wetness of the wetting bed. For the wet treatment, 80 ml of deionised water was added to the wetting bed, and aggregates reached a moisture content of 32% (sandy loam) and 44% (clay) moisture content. In the intermediate treatment, 60 ml of water was added, and aggregates reached a moisture content of 14% (sandy loam) and 20% (clay). No water was added for the dry treatment resulting in a moisture content of < 2% (sandy loam) and < 7% (clay). Microcosm containers were opened regularly to allow air circulation at the same time as moisture content maintenance. To evaluate the effects of incubation duration, microcosms were held under the treatment conditions for 1 week, 2 weeks, or 4 weeks. For each treatment, paired microcosms were analysed for aggregate stability by rainfall simulation; microbiological community composition by phospholipid fatty acid (PLFA) analysis; microbial respiration rate by rapid automated bacterial impedance technique (RABIT); and microbial biomass carbon by chloroform fumigation extraction. Microcosms were paired for the different destructive laboratory analyses, and there were three sets of replicates per treatment.

2.2 Aggregate stability and rainfall simulation

There are numerous methods used to measure aggregate stability, with many methodologies based on the wet-sieving approach (Kemper and Koch 1966; Le Bissonnais 1996; Amézketa 1999). Whilst variants of this method enable the distinction of aggregate breakdown mechanisms, the process is highly mechanical and far removed from natural erosion processes. Comparatively, rainfall simulation has been reported to be the most realistic method for measuring aggregate stability (Almajmaie et al. 2017), and so, this method was selected here to determine aggregate stability. Aggregates held in the microcosms were subjected to a rainstorm generated by a 9-m gravity-fed hypodermic needle rainfall simulator (Allton et al. 2007; Jeffery et al. 2010). Before each rainfall event, all microcosms were air-dried for 24 h (to standardise moisture conditions at the time of rainfall application to less than 7%) and weighed. The microcosms were subjected to a simulated rainfall event of 33 mm h⁻¹ for 5 min, which represents a

gentle rainfall event with a return period of less than 6 months (NERC 1975). After the rainfall simulation, samples were air-dried for 48 h and weighed.

The percentage of stable aggregates was calculated using the following equation (Almajmaie et al. 2017):

Aggregate stability (%)

$$= \frac{(\text{dry weight of aggregates post rainfall} - \text{microcosm structure})}{(\text{dry weight of aggregates pre rainfall} - \text{microcosm structure})} \times 100$$

Percentage data for aggregate stability was normalised using an arcsine transformation for statistical analyses.

2.3 Phospholipid fatty acid analysis

Phospholipid fatty acid (PLFA) analysis was used to characterise microbiological community composition, as current understanding of microbiological stabilisation potential is limited to broad taxonomic classes. PLFAs were extracted based on the procedures by Frostegård et al. (1993), Frostegård and Bååth (1994), and Bardgett et al. (1996), based on the method developed by Bligh and Dyer (1959). Aggregates for PLFA analysis were frozen at -20 °C and freeze-dried. PLFAs were extracted from 10 g of aggregates. Fatty acid methyl esters (FAMES) were identified by gas chromatography (GC) retention time. Results were expressed as a percentage of the total area of the identified peaks on the chromatogram, which was then used to assess relative abundance of fungi, gram-negative bacteria, and gram-positive bacteria. PLFAs 16:1 ω 5, 18:2 ω 6, and cis18:1 ω 9 were used as indicators for fungal biomass (Frostegård et al. 1993, 2011; Frostegård and Bååth 1994). Trans16:1 ω 11, cis16:1 ω 7, cyc17:0iso, cis17:1 ω 8, cis17:0, trans17:1 ω 8, and cis19:0 represented gram-negative bacteria. Finally, i15:0, ai15:0, i16:0, 17:0 i17:0, and ai17:0 represented gram-positive bacteria (Bardgett et al. 1996; Zelles 1999; Russ and Chamberlain 2010). Other PLFAs determined were 14:0, 15:0, i16:1, 16:0, Me17:0iso, Me17:0iso2, cyc17:0iso, 17:0br, 17:1 ω 7, 17:0(12Me), 18:1 ω 7t, 18:1 ω 13, 18:0, 19:1 ω 6, 18:0(Me), 20:4(5,8, 4, 11,14), 20 ω 5(3), and 20:0.

2.4 Microbial respiration rate

Environmental conditions could affect aggregate stability through changes in the microbial metabolic rate, in which greater activity and, potentially, more exudates would be produced when the respiration rate is higher (Costa et al. 2018). Microbial respiration rate was calculated based on the indirect impedance technique and rapid automated bacterial impedance technique (RABIT; Don Whitley Neil, UK) according to Ritz et al. (2006). This method measures the decrease in conductance of alkaline agar over time as the agar absorbs microbially produced CO₂ (Ritz et al. 2006; Butler et al. 2011, 2012). First,

RABIT cells were created by immersing electrodes in 1 ml of potassium hydroxide agar. Next, 1g of soil was weighed into a glass boat and sealed inside the RABIT cell. Samples were stored at 4 °C, then allowed 2 h at 25 °C within the RABIT equipment to allow for equilibration prior to measurement. The basal microbial respiration rate was determined over between 2- and 4-h incubation at 25 °C. The RABIT technique requires a constant temperature, as respiration rates are measured via electrical conductivity; thus, temperature was different from the incubation, but consistent across the temperature treatments. Respiration rates were corrected for dry mass of soil to account for soil moisture conditions consistent with the incubation treatment at the time of measurement. For each sample, three methodological replicates were included. Falsely terminated tests were removed from the dataset, as were outliers (identified as 1.5 times the interquartile range above or below the upper and lower quartiles). Microbial respiration rate was measured after the treatment incubation rather than during incubation due to the destructive nature of sampling and measurement. As such, the measurement of microbial respiration reflects the response of the microbial community to the consistent temperature during RABIT measurements post-incubation, mediated by previous thermal stress conditions during the incubation and substrate availability.

2.5 Microbial biomass carbon

Similar to respiration rates, an increase in microbial biomass carbon might reflect a greater absolute abundance of microbes and/or exudates, which previous studies have found could affect aggregate stability (Degens et al. 1996; Schlöter et al. 2003). It was estimated for three replicates for each treatment, based on the chloroform extraction procedure developed by Jenkinson (1976) and Vance et al. (1987). A subsample equivalent to 12.5 g air-dry weight was placed in a 50-ml Duran bottle and fumigated for 24 h, whilst another subsample was not fumigated as a control. Dissolved carbon was extracted from both samples by shaking the subsamples in 50 ml of 0.5 M K₂SO₄ for 30 min and filtering through Whatman filter papers. Concentrations of dissolved organic carbon were measured using a segmented flow analyser. Carbon flush was assessed as the concentration of DOC from fumigated samples minus the concentration of DOC from the non-fumigated samples. Biomass carbon was estimated using a conversion factor of 0.45 (British Standards Institution 1997).

2.6 Statistical analysis

Analysis of variance (ANOVA) and Tukey's HSD post hoc tests were used to assess differences in aggregate stability, microbial biomass C, and microbial respiration rate between

treatments. Pearson correlation analysis was used to test correlative associations between variables including aggregate stability, temperature, soil moisture, and microbial properties. Multiple regression analysis for both soil textures was completed using the stepwise method. R3.2.5 was used for statistical analysis (R Core Team 2018) using stepAIC from the MASS package (Venables and Ripley 2002) and the ggplot2 package (Wickham 2016), and results were considered significant at $p \leq 0.05$.

3 Results

In both sandy loam and clay samples, the duration of incubation (1 week, 2 weeks, or 4 weeks) had no significant effect on aggregate stability ($p > 0.05$, data not shown). As the focus of this study was on aggregate stability, statistical analyses were completed on pooled results as a single dataset. Furthermore, as there were no significant observed interaction effects between temperature and moisture content on aggregate stability (ANOVA, $p > 0.05$), the main effects of the climatic conditions are considered individually, i.e. the main effects of temperature considered across all moisture contents and conversely.

3.1 Aggregate stability

Aggregate stability responded differently to the temperature and moisture content treatments for the two soils tested (Fig. 1). For the sandy loam soil, aggregate stability was significantly different between moisture treatments ($p < 0.001$), with the highest aggregate stability in the dry treatment (Fig. 1B). There was no significant difference in aggregate stability between temperature treatments ($p > 0.05$, Fig. 1A). Meanwhile, for the clay soil, aggregate stability significantly increased with increasing temperature ($p < 0.005$; Fig. 1C), but there was no significant effect on aggregate stability for the moisture content treatments ($p > 0.05$; Fig. 1D).

3.2 Microbiological variables

The microbiological data provide evidence that the temperature and moisture content treatments produced changes to variables associated with metabolic activity and community composition. It was not possible to monitor respiration during the incubation, so it was measured post-incubation. The results show that the microbial community had higher respiration rates after incubation under the 5 °C treatment than the 15 °C and 30 °C treatments. It also increased with increasing moisture content in the sandy loam soil aggregates (Fig. 2A, B) and clay aggregates (Fig. 3A, B). For both temperature and moisture content, these results suggest that the high respiration rates measured post-incubation may

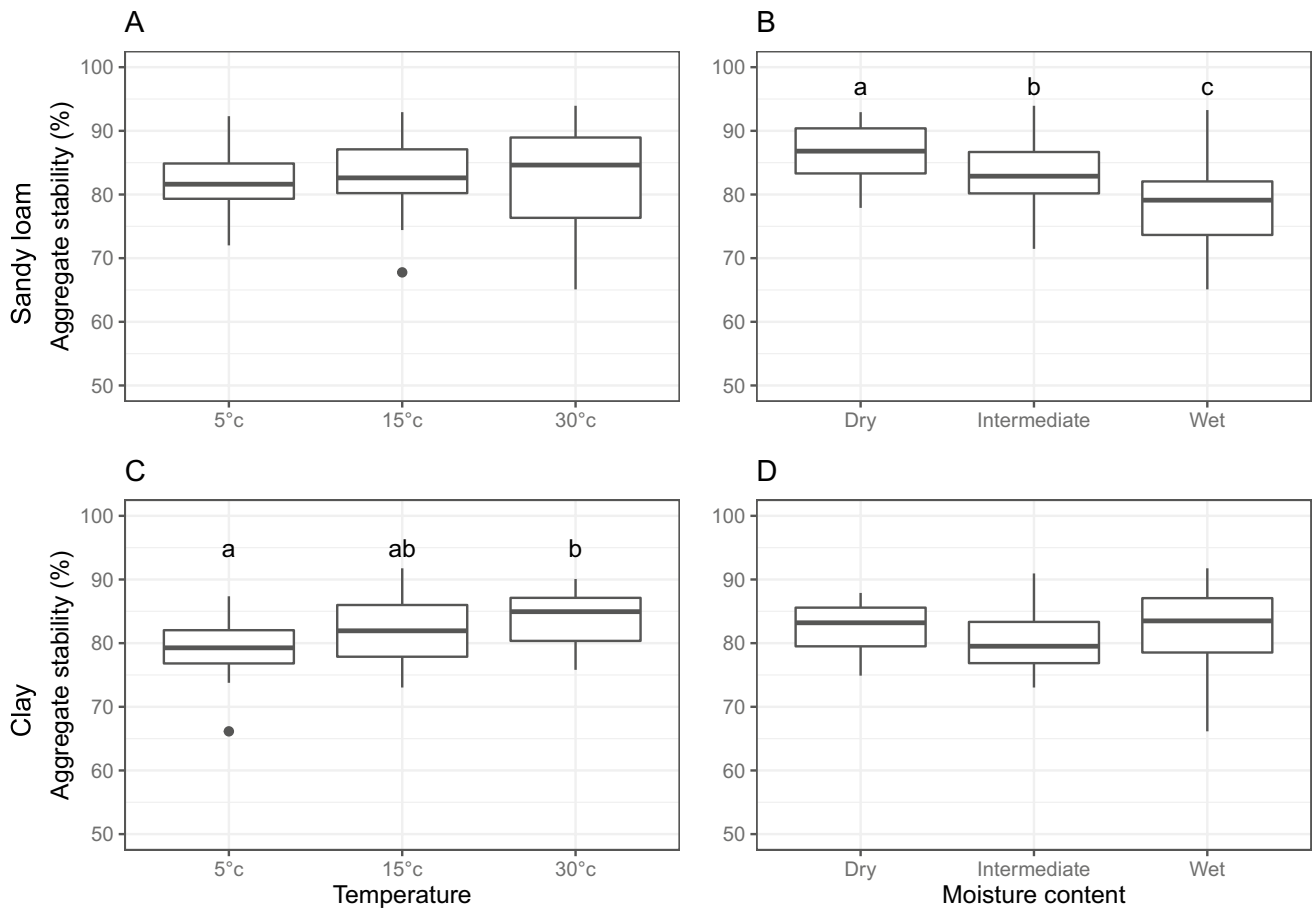


Fig. 1 Aggregate stability by **A, C** temperature and **B, D** moisture content for **A, B** sandy loam and **C, D** clay. Different letters signify statistically significant difference for arcsine transformed percentage aggregate stability

have been caused by lower metabolic activity during incubation, a ramping up of metabolic rates at more favourable thermal conditions, or a more bioavailable carbon sources at the time of measurement. Microbial biomass carbon was not significantly affected by temperature or moisture content in the sandy loam (Fig. 2C, D). However, in the clay soil aggregates, microbial biomass carbon significantly increased with increasing moisture content (Fig. 3D).

The relative abundances of gram-positive and gram-negative bacteria were not significantly affected by temperature or moisture content in the sandy loam soil aggregates (Fig. 4A–D). In the clay soil aggregates, increasing temperature significantly increased gram-positive bacteria abundance, with the relative abundance of gram-positive bacteria significantly higher at 30 °C (Fig. 5A). In contrast, gram-negative bacterial abundance decreased with increasing temperature, as the relative abundance of gram-negative bacteria was significantly higher at 5 °C and decreased with increasing temperature (Fig. 5C). In both soils tested, moisture content did not significantly influence the relative abundance of gram-positive or gram-negative bacteria (Figs. 4 and 5).

In the sandy loam soil aggregates, the relative abundance of fungi was significantly higher in the driest treatment, but fungal abundance was not significantly affected by temperature (Fig. 4E, F). In the clay soil aggregates, temperature and moisture content conditions did not significantly influence fungal abundance (Fig. 5E, F).

3.3 Statistical analyses

For the sandy loam soil, aggregate stability was significantly negatively correlated with moisture content, but had no significant correlation with temperature (Table 1). Temperature was significantly negatively correlated with the relative abundance of fungi. Moisture content was significantly positively correlated with microbial respiration and significantly negatively correlated with the relative abundance of fungi. Aggregate stability showed significant positive correlations with microbial biomass carbon and relative abundance of gram-negative bacteria.

Pearson correlation analysis on the clay aggregates showed a significant positive correlation between aggregate stability

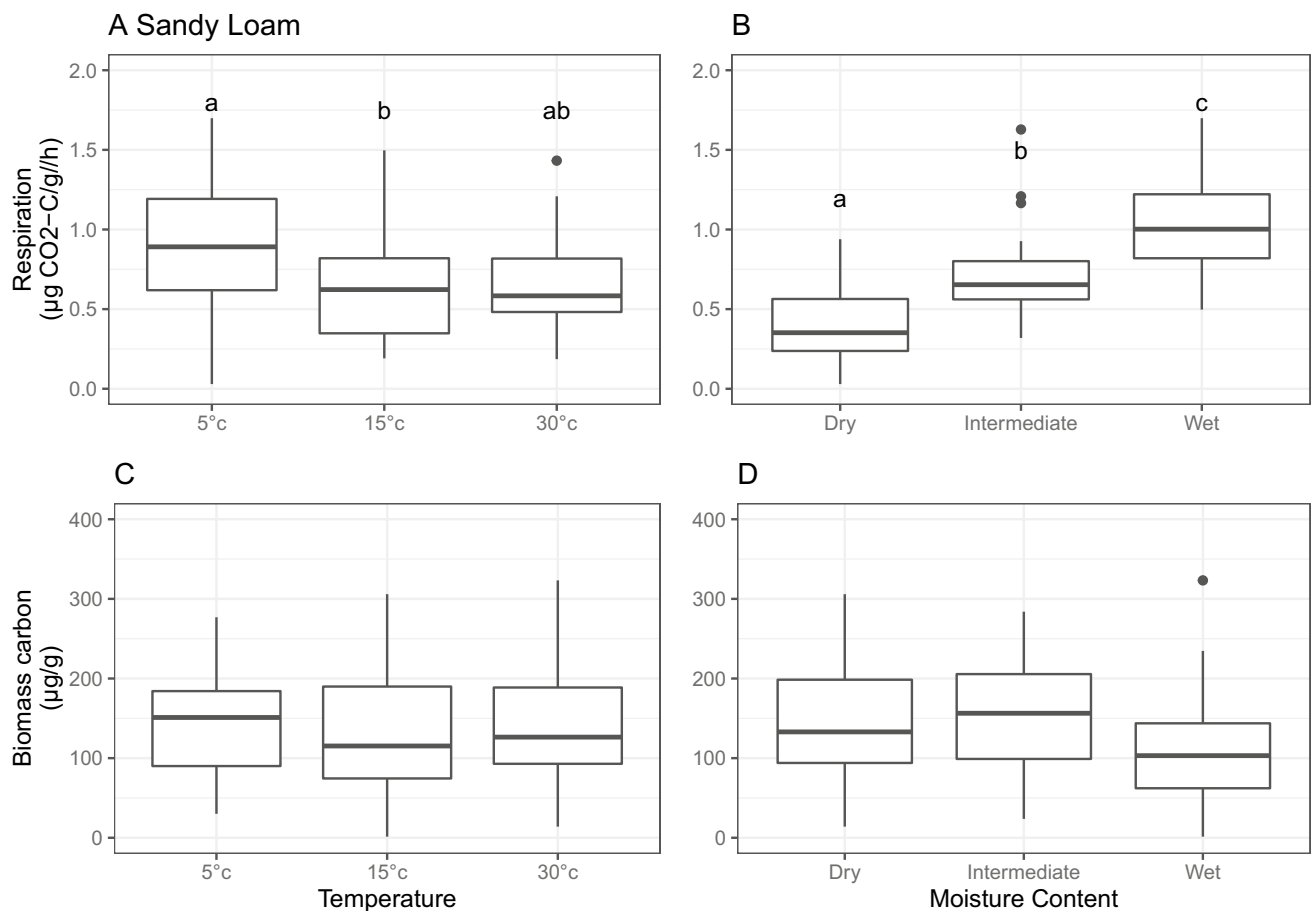


Fig. 2 Microbial respiration (A, B) and microbial biomass carbon (C, D) by temperature and moisture content for sandy loam aggregates. Different letters signify statistically significant differences

and temperature (Table 2). Contrary to the sandy loam soil results, aggregate stability was not correlated with moisture content. Temperature had a significantly negative correlation with microbial respiration and the relative abundance of gram-negative bacteria, but temperature was significantly positively correlated with the relative abundance of gram-positive bacteria. Moisture content had a significant positive correlation with microbial respiration and microbial biomass carbon. Aggregate stability was also shown to be significantly negatively correlated with the relative abundance of gram-negative bacteria.

Multiple regression analysis showed that several microbiological properties were significant predictors of aggregate stability (Table 3). However, there were substantial differences between the sandy loam and clay minimally adequate models. For the sandy loam soil aggregates, stability was best predicted by microbial respiration, biomass carbon, and gram-negative abundance (adjusted $R^2 = 0.209$, $p < 0.001$). Meanwhile, for the clay soil aggregates, it was best predicted by the relative abundance of gram-negative bacteria (adjusted $R^2 = 0.064$, $p = 0.013$).

4 Discussion

The aim of the experimental laboratory study was to determine how shifts in temperature and moisture conditions, which affect the microbiological community, affect soil aggregate stability. The main findings were (i) the effects of temperature and moisture on aggregate stability differed between the two soil types tested but not the duration of incubation (hypothesis 4); (ii) aggregate stability differed significantly by pre-rainfall incubation temperature for the clay soil tested, with the hottest temperature having the greatest stability and relative abundance of gram-positive bacteria (hypothesis 1); (iii) aggregate stability differed significantly by pre-rainfall incubation soil moisture for the sandy loam soil tested, with the driest sandy loam aggregates having the greatest stability and relative abundance of fungi (hypothesis 2); and (iv) no significant interaction effects were observed for moisture and temperature treatments in this factorial experiments for the two soils tested (hypothesis 3).

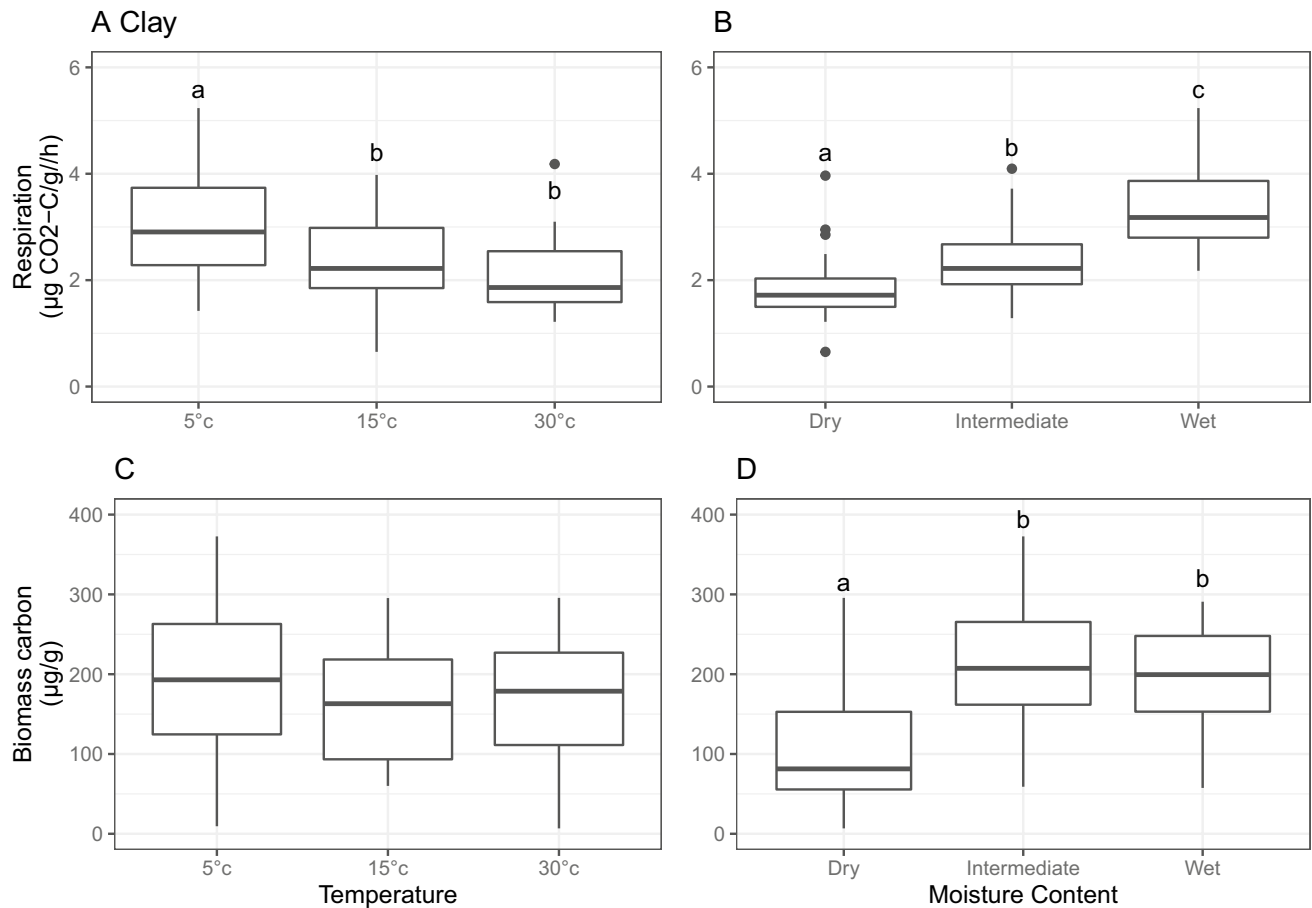


Fig. 3 Microbial respiration (A, B) and microbial biomass carbon (C, D) by temperature and moisture content for clay aggregates. Different letters signify statistically significant differences

In this study, aggregate stability increased significantly with increasing temperature for the clay soil tested (Fig. 1A, C). This positive relationship between aggregate stability and temperature for the clay aggregates is not supported by theory on the physico-chemical effects of temperature, which hypothesises that aggregate stability would decrease with increasing temperature as interparticle bonds are weakened through several physical and chemical mechanisms (Plum and Esrig 1969; Dexter et al. 2010; Kelishadi et al. 2018). Increased temperature is hypothesised to cause an increase in the internal energy of particles resulting in the expansion of entrapped air and double layers in clay, thereby disrupting interparticle bonds and reducing structural stability. However, fieldwork has provided some empirical support for our findings. Lavee et al. (1996) investigated aggregate stability along a climatological transect and found that aggregate stability was highest under temperatures of 15 °C and lower for temperatures between 0 and 10 °C and temperatures exceeding 35 °C. This unimodal peak in aggregate stability was suggested to be a combination of the physical effects of temperature and biotic activity, though

soil microbiological properties were not measured in that study. A similar unimodal distribution in aggregate stability was found across an 800-m elevation gradient in an alpine meadow, which was related to the combined effects of temperature and moisture on microbiological activity and soil carbon (Li et al. 2017).

Aggregate stability decreased significantly with increasing moisture content for the sandy loam soil (Fig. 1A, C). This soil moisture effect on sandy loam aggregates is independent and unrelated to the more widely reported impact of moisture content at the time of rainfall (Cousen and Farres 1984; Ben-Hur and Lado 2008; Almajmaie et al. 2017). Moisture content was equilibrated for all treatments by air-drying prior to the rainfall event. Thus, these results indicate that moisture content during incubation affects processes internal to the aggregate that affect its stability.

The microbiological data confirms that the temperature and moisture treatments did affect community compositional and functional indicator metrics. However, similar to aggregate stability, the results differed between the two soils tested. For both soils, the only microbiological data

A Sandy Loam

Gram-positive bacteria (%)

5°C 15°C 30°C

B

Dry Intermediate Wet

C

Gram-negative bacteria (%)

5°C 15°C 30°C

D

Dry Intermediate Wet

E

Fungi (%)

5°C 15°C 30°C

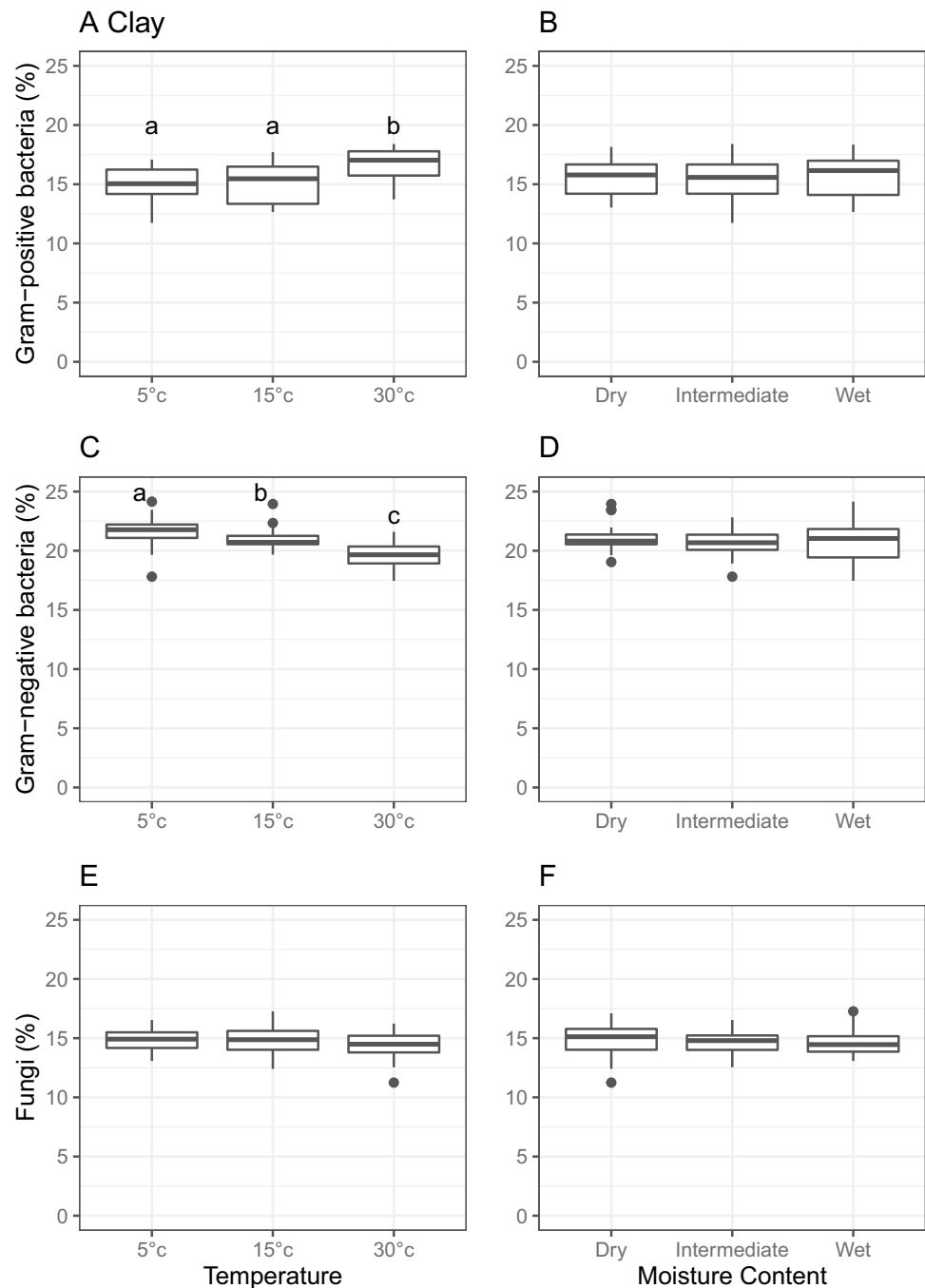
F

Dry Intermediate Wet

Moisture Content

For the moisture effect observed in the sandy loam soil, previous studies have found the fungal hyphae are more tolerant of low soil moisture (Jennings 1987; De Boer et al. 2005) and that fungal biomass correlates with higher aggregate stability due to hyphal growth and particle enmeshment (Degens et al. 1996; Lehmann and Rillig 2015; Lehmann et al. 2020). Reduced aggregate stability is observed consistently in studies that inhibit the growth of the fungal component of the soil microbiological community using fungicides (Beare et al. 1997; Bossuyt et al. 2001; Tang et al. 2011). However,

Fig. 5 The relative abundance (%) of gram-positive bacteria (A, B) gram-negative bacteria (C, D), and fungi (E, F) by temperature and moisture content for clay aggregates. Different letters signify statistically significant differences



soil moisture could affect aggregate stability through both physico-chemical and biological mechanisms. For example, the higher proportion of micropores to macropores in the clay soil aggregates may have made the aggregates less susceptible to fissure than the sandy loam aggregates (Peng and Horn 2005; Borken and Matzner 2009). These physical differences may also underlie biological-mediated responses to soil moisture that affect aggregate stability. Moisture content conditions influence numerous soil properties critical for microbiological life, including soil hydration status, oxygen

availability, pore connectivity, bacterial motility, and thus, resource accessibility (Skopp et al. 1990; Or et al. 2007a; Moyano et al. 2013). These mechanisms could explain why respiration and microbial carbon biomass were identified as significant predictors in the regression analysis for the sandy loam soil (Table 3). Thus, there is evidence that changes in soil moisture affects aggregate stability, at least in the sandy loam soil tested; further research is needed to examine in detail the physico-chemical and biological mechanisms that might be responsible.

Table 1 Pearson correlation analysis for sandy loam aggregates

	Aggregate stability (arcsine)	Temperature	Moisture content	Incubation duration	Microbial respiration ($\mu\text{g CO}_2\text{-C g}^{-1} \text{h}^{-1}$)	Microbial biomass carbon ($\mu\text{g g}^{-1}$)	Gram-positive bacteria (mol%)	Gram-negative bacteria (mol%)
Temperature	0.065							
Moisture	− 0.529***	-						
Incubation duration	0.018	-	-					
Microbial respiration ($\mu\text{g CO}_2\text{-C g}^{-1} \text{h}^{-1}$)	− 0.304**	− 0.213	0.655***	0.192				
Microbial biomass carbon ($\mu\text{g g}^{-1}$)	0.253*	− 0.001	− 0.171	0.065	− 0.049			
Gram-positive bacteria (mol%)	− 0.070	− 0.036	− 0.050	0.115	− 0.157	− 0.013		
Gram-negative bacteria (mol%)	0.273*	− 0.112	− 0.115	− 0.296**	0.068	− 0.022	− 0.151	
Fungi (mol%)	− 0.070	− 0.250*	− 0.314**	0.057	− 0.220*	− 0.102	− 0.229*	− 0.155

* $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$

For the temperature effect observed for the clay soil, the shift in the relative abundances of gram-positive and gram-negative bacteria is supported by previous research, which suggests that temperature-induced substrate constraints limit gram-negative bacteria, whilst gram-positive bacteria are better adapted to acquire resources at higher temperatures (Biasi et al. 2005; Feng and Simpson 2009). Whether

these shifts in microbiological abundance are responsible for the differences in aggregate stability is unknown. Similarly, respiration post-incubation differed by temperature for the clay soil, with lower rates in higher temperature treatments (Fig. 3A). These results suggest that respiration rates were potentially higher during the hot temperature incubation, leading to substrate depletion prior to or during the

Table 2 Pearson correlation analysis for clay aggregates

	Aggregate stability (arcsine)	Temperature	Moisture content	Incubation duration	Microbial respiration ($\mu\text{g CO}_2\text{-C g}^{-1} \text{h}^{-1}$)	Microbial biomass carbon ($\mu\text{g g}^{-1}$)	Gram-positive bacteria (mol%)	Gram-negative bacteria (mol%)
Temperature	0.373***							
Moisture	0.031	-						
Incubation duration	− 0.112	-	-					
Respiration ($\mu\text{g CO}_2\text{-C g}^{-1} \text{h}^{-1}$)	− 0.138	− 0.430***	0.657***	− 0.001				
Microbial biomass carbon ($\mu\text{g g}^{-1}$)	− 0.133	− 0.104	0.396***	0.056	0.380***			
Gram-positive bacteria (mol%)	0.099	0.412***	0.036	− 0.008	− 0.130	− 0.154		
Gram-negative bacteria (mol%)	− 0.275*	− 0.603***	− 0.165	− 0.059	0.163	0.007	− 0.518***	
Fungi (mol%)	− 0.047	− 0.213	− 0.027	0.180	0.098	0.240*	− 0.522***	− 0.115

* $p < 0.05$; *** $p < 0.001$

Table 3 Multiple regression analysis for aggregate stability and microbial predictor variables for the sandy loam and clay aggregates, using step-wise method

Predictors	Aggregate stability arcsine (sandy loam)			Aggregate stability arcsine (clay)		
	Coefficient estimates	Std. error	<i>p</i> value	Coefficient estimates	Std. error	<i>p</i> value
Intercept	0.36	0.22	0.1	1.3	0.14	< 0.001
Respiration ($\mu\text{g CO}_2\text{-C g}^{-1} \text{ h}^{-1}$)	− 0.1	0.03	0.0025			
Microbial biomass carbon ($\mu\text{g g}^{-1}$)	0.0004	0.00016	0.016			
Gram-negative bacteria (mol%)	0.029	0.0097	0.0036	− 0.17	0.0065	0.013
Observations	81			81		
R^2 /adjusted R^2	0.239/0.209			0.076/0.064		
<i>p</i> value	< 0.001			0.013		

post-incubation testing (Moyano et al. 2013). A higher respiration rate during incubation could generate higher microbial abundances and exudates (Degens et al. 1996), but this mechanism is not supported by the data from this study (Fig. 3C). Furthermore, there are other significant differences in respiration rates for other treatments and the sandy loam soil that do not correspond to changes in aggregate stability (e.g. Figs. 2A and 3B). Thus, whilst microbial biomass carbon and respiration reflect microbial turnover of carbon and biomass synthesis and growth (Degens et al. 1996) and have often been used as indicators of microbially mediated processes (Schloter et al. 2003; Ritz et al. 2009; Truu et al. 2009), further research is necessary to determine how environmental conditions affect aggregate stability through changes in microbiological community composition, spatial arrangement, and biomechanical properties of the biologically generated organic compounds and structures.

This study was inspired by Dr. Ian G. Droppo's long and rich research career. His aquatic sediment research advanced our scientific understanding of the biological mechanisms for suspended sediment aggregation and the influence of chemical and physical factors on floc strength and sediment erodibility (Liss et al. 1996; Droppo 2001, 2004; Droppo et al. 2007, 2008, 2015, 2016; Grabowski et al. 2011, 2012). This whole system perspective on sediment composition and consideration of particle interactions has been influential in refocusing fundamental erosion and transport research at the floc and aggregate scale and broadening the factors investigated (e.g. Chen et al. 2022). Dr. Droppo has also helped to unify research soils and sediments, through his pioneering research on floc/aggregate characterisation (Droppo et al. 2005) and the transformation of eroded soil into riverine flocs (Grangeon et al. 2014). Through this research, we hope to encourage more cross-disciplinary and integrative research, informed by Dr. Droppo's work, to support renewed investigation into the dynamic and responsive nature of soil erodibility. To do this, we recommend further research on the mechanistic connections between environmental conditions, soil properties, microbiological

community composition, and their stabilising functions. We recommend a renewed focus at the aggregate scale to determine how changes in environmental conditions (trends and increased variability) impact on the production of biogenic stabilising structures and functions to affect particle interactions at the aggregate scale. This work should attempt to separate out abiotic and biotic controls, for example, by identifying causal relationships through the comparison of sterile and non-sterile soil microcosms, and use advanced methods to monitor changes in communities (e.g. quantitative PCR, amplicon sequencing, and/or metagenomics) and their metabolic activity (e.g. CO_2 respiration during incubation, metabolomics). This fundamental research should link to larger scale work to consider how these factors in turn influence the hydrological and geomorphological processes that determine overland flow, erosion mechanisms (e.g. sheet and rill erosion), and sediment transport.

5 Conclusions

In this study, controlled laboratory experiments were used to determine how shifts in soil temperature and soil moisture, which alter the soil microbiology, affect the stability of soil aggregates. The key results were that these two climate-related conditions affected the two tested soils differently. Soil moisture had a significant effect on aggregate stability for the sandy loam soil, but temperature had a significant effect on the clay soil. The sandy loam soil aggregates were most stable following incubation under the driest condition and least stable under the wettest condition. Importantly, these results are different from the oft-reported effect of soil moisture on aggregate stability at the time of rainfall, as moisture content was equilibrated before stability testing. The clay soil aggregates were most stable following incubation under the hottest temperature (30 °C) and least stable at the coolest temperature (5 °C). The two soils differed in texture but were otherwise as similar as possible for natural soil samples (e.g. same organic content, managed by the

same farmer, and found in close proximity (< 500 m apart)). The changes in aggregate stability aligned with shifts in the microbiological community, a higher relative abundance of fungus in the sandy loam aggregates under dry conditions and more gram-positive bacteria in the clay soil aggregates under hot conditions. These results highlight the need to investigate further the interactions and feedbacks between climatic conditions, soil properties, microbiological communities, and aggregate stability. The study adds to a growing body of literature that documents that aggregate stability is a dynamic soil property responsive to shifts in climate, which must be investigated in greater detail and at larger spatial scales to better understand their implications for soil erodibility and the prediction of soil erosion with climate change.

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Data availability Data from this study are openly available from Cranfield University's Online research data repository (CORD) at DOI: <https://doi.org/10.17862/cranfield.rd.23668803>.

Declarations

Competing interests The authors declare no competing interests.

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Do temperature and moisture conditions impact soil microbiology and aggregate stability?

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