nature ecology & evolution

Article

https://doi.org/10.1038/s41559-025-02704-9

# Opportunities and challenges for monitoring terrestrial biodiversity in the robotics age

Received: 11 November 2024

Accepted: 7 April 2025

Published online: 22 May 2025

Check for updates

A list of authors and their affiliations appears at the end of the paper

With biodiversity loss escalating globally, a step change is needed in our capacity to accurately monitor species populations across ecosystems. Robotic and autonomous systems (RAS) offer technological solutions that may substantially advance terrestrial biodiversity monitoring, but this potential is yet to be considered systematically. We used a modified Delphi technique to synthesize knowledge from 98 biodiversity experts and 31 RAS experts, who identified the major methodological barriers that currently hinder monitoring, and explored the opportunities and challenges that RAS offer in overcoming these barriers. Biodiversity experts identified four barrier categories: site access, species and individual identification, data handling and storage, and power and network availability. Robotics experts highlighted technologies that could overcome these barriers and identified the developments needed to facilitate RAS-based autonomous biodiversity monitoring. Some existing RAS could be optimized relatively easily to survey species but would require development to be suitable for monitoring of more 'difficult' taxa and robust enough to work under uncontrolled conditions within ecosystems. Other nascent technologies (for instance, new sensors and biodegradable robots) need accelerated research. Overall, it was felt that RAS could lead to major progress in monitoring of terrestrial biodiversity by supplementing rather than supplanting existing methods. Transdisciplinarity needs to be fostered between biodiversity and RAS experts so that future ideas and technologies can be codeveloped effectively.

To conserve biodiversity effectively, we must be able to accurately and comprehensively monitor species populations to anticipate and ameliorate declines proactively[1]. This is critical, given that recent projections suggest that up to two million species are at risk of extinction, with plants and invertebrates most at threat[2]. Indeed, conservationists need to monitor biodiversity across all ecosystems, from urban areas to inaccessible wilderness, to mitigate the drivers of species loss. These monitoring programmes need to be robust, predicting future species extinctions and ecosystem collapse well in advance of tipping points being reached.

Monitoring terrestrial biodiversity is time-consuming and expensive to replicate spatially and temporally. Many ecological relationships only become apparent following extensive surveys over broad geographic scales, often through time, which can be unfeasible using current methods[3]. Comprehensive monitoring might encompass tens, hundreds or even thousands of sites that need repeated and ideally synchronous surveying. Biodiversity monitoring also requires expertise in field observation and, for some taxa, detailed knowledge of taxonomy or the use of specialist techniques such as collection and analysis of genetic material[4]. In addition, species frequently have restricted habitat associations, meaning that the effectiveness of monitoring can be severely hampered or biased by environmental factors, including whether researchers can access sites and tolerate the conditions within them. Human surveyors can also overlook cryptic, elusive and small species[5]. Overcoming these constraints requires a step change in the methods used to monitor terrestrial species populations across all taxonomic groups.

e-mail: s.pringle@kent.ac.uk

https://doi.org/10.1038/s41559-025-02704-9

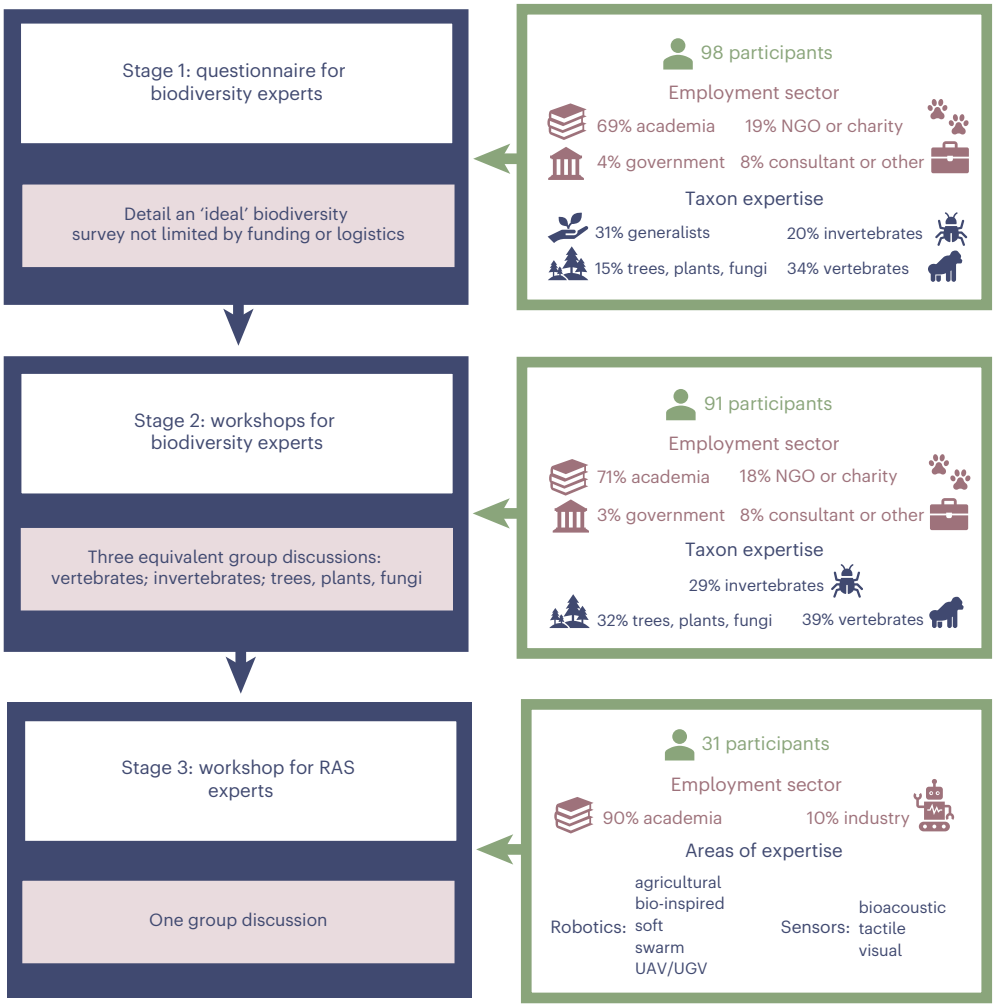



**Fig. 1 | The modified Delphi technique used to identify the methodological barriers that currently hinder terrestrial biodiversity monitoring and the opportunities and challenges that RAS offer in overcoming these barriers.** 'Soft', robots that use compliant materials to mimic natural locomotion; 'swarm', multiple robots, either homogeneous or heterogeneous, that are interconnected; UGV, uncrewed ground vehicle.

RAS are technologies that can sense, analyse, interact with and manipulate their physical environment[6]. RAS have been developed for many applications (for instance, military applications[7], agriculture[8], infrastructure maintenance[9] and surgery[10]) and in recent years have been widely adopted for monitoring of marine ecosystems[11]. The core technology underpinning current applications may also offer the potential to complement and/or extend our terrestrial biodiversity monitoring capabilities[12]. For example, an uncrewed aerial vehicle (UAV)-borne tool (https://outreachrobotics.com) suited to infrastructure inspection has been used to sample plants from inaccessible cliffs. Likewise, technology developed for inspection and maintenance of below-ground pipes and sewers (https://pipebots.ac.uk) could be used to survey species that inhabit burrows.

Mobilizing RAS for biodiversity monitoring could substantially advance conservation efforts[13,14]. However, to date, there has been no systematic attempt to assess this potential. Here, we report the findings from a modified Delphi process[15] that evaluated how we might adapt or develop current RAS to transform species surveys in terrestrial ecosystems (Fig. 1). Through an online questionnaire and workshops, we collated and synthesized knowledge from 98 biodiversity experts and 31 RAS experts, thereby identifying the major methodological barriers that currently hinder monitoring and exploring the opportunities and challenges that RAS offer in overcoming these barriers. The collective field survey experience of biodiversity experts encompassed 109 countries and a diversity of biomes and taxa (Fig. 2 and Extended Data Figs. 1 and 2).

## Results

In stage 1 of our modified Delphi technique, comprising an online questionnaire, we asked biodiversity experts to identify methodological barriers that they expected to encounter in an 'ideal' survey that was not limited by funding or logistics. We did not mention the use of RAS, or how they might be incorporated into surveys. Barriers fell into four broad categories: (1) site access, (2) species and individual detection, (3) data handling and processing, and (4) power and network availability (Table 1). The proportions of experts who highlighted barriers within each category varied by taxon; however, for all taxa, site access and species and/or individual detection were mentioned most frequently (Fig. 3).

In stage 2, which consisted of an online workshop, the same biodiversity experts considered the opportunities and challenges that RAS offered in terms of overcoming the barriers identified in stage 1 (Table 1 and Fig. 4). The opportunities identified most often involved the potential use of RAS to survey large spatial areas, with real-time species identification and handling of high data volumes. The major technological challenges highlighted with respect to RAS were power availability, generation of validated training data, elimination of the need for multiple sensor types and the risk of misidentification by automated classifiers[16].

https://doi.org/10.1038/s41559-025-02704-9

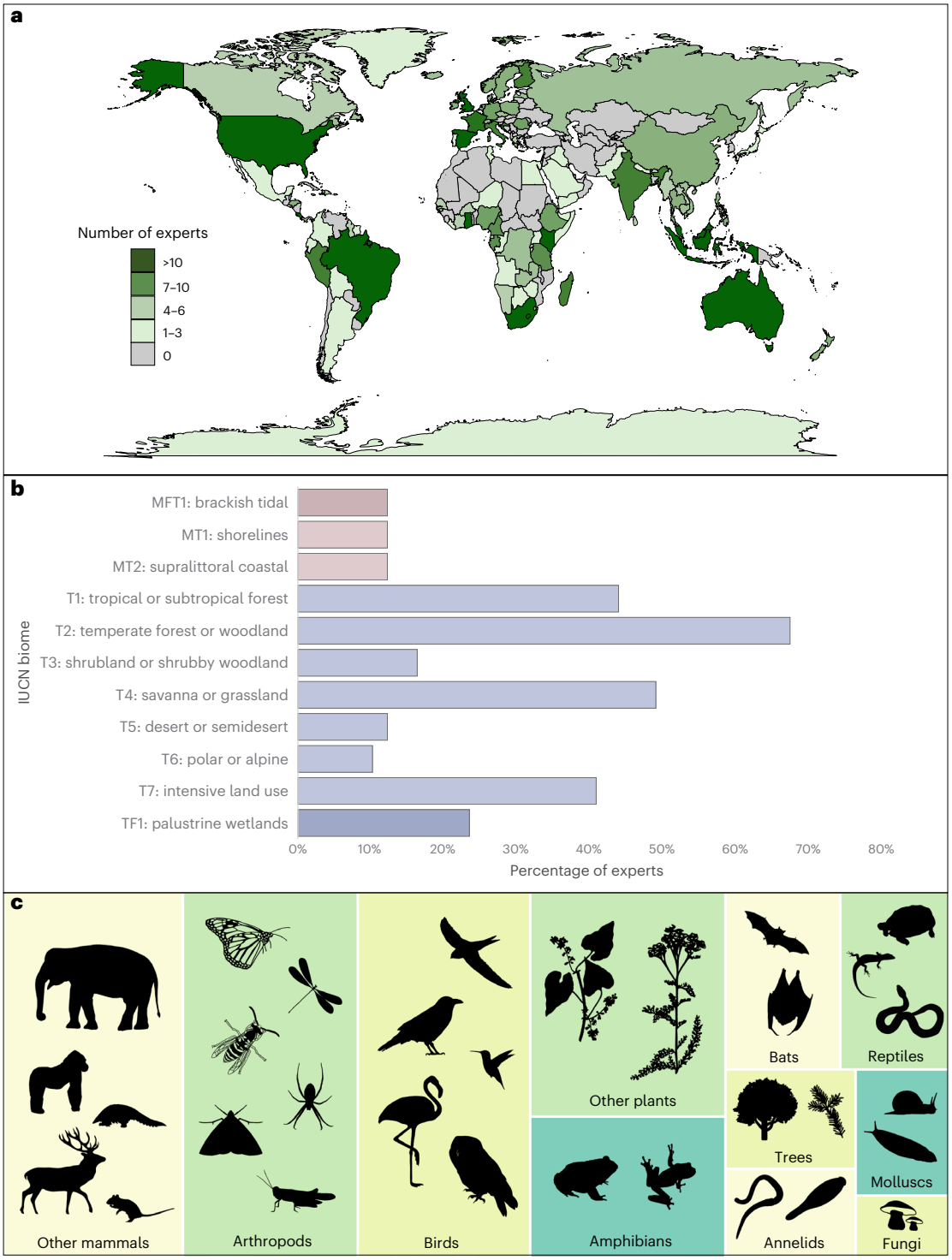



**Fig. 2 | Terrestrial biodiversity monitoring experience of the 98 experts who completed stage 1 of the modified Delphi technique. a**, Countries in which the experts had experience of conducting terrestrial biodiversity monitoring. **b**, Percentage of biodiversity experts with experience of monitoring biodiversity in each biome (86% of experts had conducted surveys in more than one biome), listed according to their IUCN classification[79]. MFT1, terrestrial transitional freshwater/marine; MT1–MT2, terrestrial transitional marine; T1–T7, terrestrial; TF1, terrestrial transitional freshwater. **c**, Relative numbers of experts with experience of monitoring each taxon (indicated by the area of each rectangle; 70% of experts had conducted surveys of more than one taxon). Bats are separated from other mammals, and trees from other plants, because the survey methods are notably different. Credits: World map outline in **a** is ©OpenStreetMap contributors; data are available under an Open Database License (https://openstreetmap.org/copyright). The icons in **c** are from PhyloPic (https://www.phylopic.org) contributed under a Creative Commons licence CC0 1.0.

### Barrier category 1: site access

Biodiversity experts widely acknowledged issues pertaining to site access. All participants identified the potential of RAS to survey over large spatial scales as an opportunity. The ability of RAS to survey over large areas was also seen as facilitating 'true habitat replicates to avoid pseudoreplication'. The opportunity to use RAS to survey repeatedly with high spatial resolution would bring a 'level of confidence that the results are robust and repeatable'. Using multiple RAS to sample

**Table 1 | Methodological barriers that currently hinder terrestrial biodiversity monitoring and the opportunities and challenges that RAS offer in overcoming each of these barriers**

| Barrier category | Barrier | RAS opportunity or challenge | Brief description |
|---|---|---|---|
| 1. Site access | Surveying over large spatial scales | Opportunity | Autonomous monitoring at landscape scales<br>Replicating surveys at multiple sites over large geographical areas |
| | Surveying remote areas far from infrastructure | Opportunity | Accessing locations remote from roads and other infrastructure<br>Monitoring sites that are time-consuming to access |
| | Surveying hazardous or inaccessible sites | Opportunity | Access to sites that need climbing (for instance, cliffs or forest canopies)<br>Sampling sites at night or where personal safety or security is at risk |
| | Surveying taxa at random sites | Opportunity | Enabling representative sampling at suitable scale and stratification<br>Avoiding sample pseudoreplication |
| | Surveying multiple locations simultaneously | Opportunity | Time-synchronous surveys at multiple sites<br>Surveying taxa whose activity may be weather-dependent |
| | Surveying structurally complex habitats | Opportunity | Sampling within dense habitats (for instance, deadwood, grass tussocks or snow)<br>Sampling soils, underground animal burrows, or bat colonies in caves or trees |
| | Surveying at high spatial resolution | Opportunity | Ability of sensor to get to exact locations repeatedly<br>Enabling microscale tracking |
| | Designing environmentally robust sensors | Challenge | Resistance, resilience and durability of the sensors and/or probes in the field<br>Being species-proof and avoiding risk of vandalism or theft |
| | Surveying restricted and off-limits locations | Challenge | Areas affected by legal, conflict and political issues<br>Uncertainty of tenure or ownership status for many locations |
| 2. Species and/or individual detection | Eliminating the need for multiple sensors | Challenge | Integration of multiple sensor types<br>Ablility to deal with wide range of species sizes |
| | Discriminating or identifying individuals at distance | Challenge | Distance limitations of visual sensors (for instance, detection of plant ligules)<br>Difficulties in identifying individuals of a species |
| | Surveying without disturbing taxa or habitats | Challenge | Non-invasive sensors that will not disturb species or habitats<br>Impacts on non-target species |
| | Surveying through objects or in low light levels | Challenge | Detection when visibility is restricted (for instance, through vegetation or cloud)<br>Detection of ectotherms at night |
| | Surveying ecological processes | Challenge | Monitoring interactions (for instance, pollination) or ecological processes<br>Monitoring plant physiology |
| 3. Data handling and processing | Handling high data volumes | Opportunity | Storage, energy costs and edge processing of extreme volumes of data<br>Data transfer in real time to avoid data loss through sensor disturbance |
| | Identification of species in real time | Opportunity | Automated species identification by the RAS equipment<br>Overcoming geographic and taxonomic bias |
| | Surveying over long temporal periods | Opportunity | Surveying sites continuously over extended periods<br>Resurveying sites many times during a year and over many years |
| | Surveying rare, elusive or cryptic species | Challenge | Ensuring species detection (for instance, behaviourally cryptic diurnal taxa)<br>Misidentifying rare or cryptic species and different sexes or life stages |
| | Surveying little-known or 'difficult' taxa | Challenge | Monitoring little-known taxa<br>Monitoring species with poorly defined taxonomy |
| | Risk of misidentification by classifiers | Challenge | Identifying little-known or 'difficult' taxa using AI tools<br>Dealing with undescribed species |
| | Generating validated classifier training data | Challenge | Availability of training data for classifiers and/or expertise for validation<br>Ground-truthing and geographical relevance of classifier data |
| | Designing RAS for non-expert operation | Challenge | Sensor easy to operate (for instance, to facilitate non-expert input)<br>Accessibility of AI methods and training resources for non-experts |
| 4. Power and network availability | Availability of communication network | Opportunity | Areas without access to mobile networks<br>Network connections for real-time or cloud data access and storage |
| | Remote control and maintenance of RAS | Opportunity | Ability to control remotely (for instance, sensors in tree canopies)<br>Self-reporting malfunctions for long-term sensor deployments |
| | Limited power availability | Challenge | Sustainable power, robust to climate, to support monitoring stations<br>Reducing the weight of power systems |
| | Negative environmental impact of e-waste | Challenge | Environmental impact of production and/or decommissioning of RAS<br>Retrieving inaccessible RAS equipment at end of life |

These were identified by biodiversity experts during stage 2 of the modified Delphi technique.

multiple sites simultaneously was viewed as 'important for taxa whose activity may be especially weather-dependent' (for instance, reptiles[17]). RAS surveys of areas distant from infrastructure would be beneficial where 'lone working at remote locations is sometimes dangerous, especially where terrain is rugged'. Furthermore, RAS might transport heavy equipment to inaccessible areas.

In stage 3 of our modified Delphi technique, an online workshop, RAS experts proposed that biodiversity could be monitored using

https://doi.org/10.1038/s41559-025-02704-9

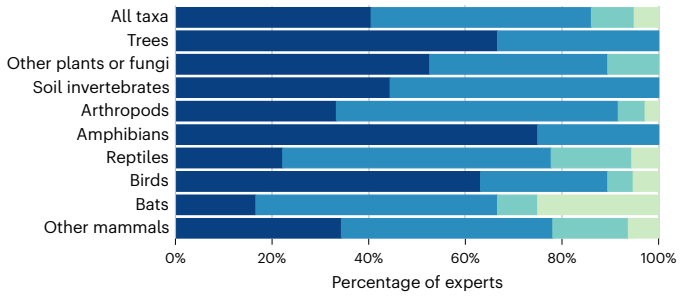



**Fig. 3 | Categories of methodological barriers to monitoring terrestrial biodiversity, as reported by biodiversity experts during stage 1 of the modified Delphi technique.** Percentages of biodiversity experts who identified methodological barriers, in four broad categories, that they expected to encounter in an 'ideal' survey that was not limited by funding or logistics, according to taxa. Dark blue, site access; blue, species and individual detection; teal, data handling and processing; light green, power and network availability.

UAVs, uncrewed ground vehicles or legged (field) robots. Although UAVs are most commonly employed, legged robots with embedded intelligence have been used to monitor vegetation in dunes, screes, grasslands and forests[18]. A legged robotic system has also been shown to generate forest tree inventories faster and more cost-effectively than traditional forestry methods[19]. Recent locomotion developments are likely to extend the operational domain of legged robots to more complex ecosystems[20]. RAS could operate either independently, or collectively as 'swarms' (multiple robots, either homogeneous or heterogeneous, that are interconnected[7]). Multirobot swarms could operate by 'coordinating activity and deciding when to sample, rather than just running on fixed schedules'. For example, multirobot swarms might use artificial intelligence (AI) to divide up a large area, ground-truth the habitat types and then sample from representative locations. UAVs could also place and retrieve sensors across an area using technology developed for environmental monitoring[21]. These methods may be suitable for surveying species such as snow leopards, *Panthera uncia*, which have very low population densities over extremely rugged terrain[22].

RAS experts noted that RAS are extensively used to navigate through structurally complex areas (for instance, nuclear facilities[23] or inside aircraft wings[24]). Tactile feedback manipulators[25] could enable robots to move through dense scrub by 'feeling' their way, whereas UAVs could use visual navigation[26] to avoid collisions within cluttered forests. Tree-climbing robots[27], rather than humans, could survey forest canopies, which would 'circumvent enormous training and h[ealth] & s[afety] issues'. Other technologies, including subcentimetre-sized soft crawling biomedical engineering robots[28], may enable the monitoring of annelids in topsoil. A benefit of soft-bodied robots is their ability to flexibly change shape; they are also considered to be safer in environments where they might interact with humans or species[29]. Harsh weather conditions are challenging for researchers undertaking surveys. They also pose problems for RAS that can fail in extreme temperatures, humidity, rain, electrical storms and strong winds. RAS experts confirmed that 'most lab[oratory]-built robots do not have great corrosion resistance' and that commercial 'electronic components are not built for arctic temperatures'. However, recently engineered 'thermally agnostic' drones, capable of working in very hot and cold environments, offer a potential way forward[30].

Biodiversity experts commented that monitoring sites may be difficult to access for many reasons, including political and security issues or uncertainties surrounding ownership. Certain types of RAS (for instance, UAVs) also have military or surveillance connotations[31]. Illustrating this point, one expert reported that efforts to monitor biodiversity 'had been met with fierce local resistance, with their drones routinely targeted by firearms'. The importance of working within legal constraints, engaging with local communities, and integrating RAS-collected data with local and indigenous knowledge of the environment[32] was stressed.

### Barrier category 2: species and individual detection

To monitor terrestrial biodiversity effectively using visual cues, RAS sensors must be able to detect species over a wide size range (for instance, invertebrates from <<1 mm to 1 m (ref. 33) and vascular plants from ~1 cm to ~100 m (refs. 34,35)). The microscopic size of critical features is problematic for plant surveys, as identifying some species is dependent on almost-invisible ligules and hairs[36]. Similar difficulties are faced with invertebrates as it is 'impossible to ID [identify] some taxa without dissection'. This places substantial demands on sensor design. Many biodiversity experts doubted whether the need for many sensors for multiple taxa[37] could be eliminated. RAS experts agreed that 'realistically, [RAS need to] use multiple sensors for different scales'. Some techniques that are being adopted in biodiversity monitoring might be further developed to extend sensor capabilities. For example, passive acoustic recordings could be enhanced through time-series analysis[38] to address sound attenuation that hampers detection of quiet species. Chemosensors ('electronic noses'), which are used in diverse agricultural and forestry applications[39], might detect unique volatile organic compounds emitted by plants. Collection and removal of physical samples is also possible. Of particular interest are DNA fragments left behind by organisms in their environment (eDNA[40]) that can be used to detect the presence of species. Recent advances in the robotic collection of eDNA samples (for instance, from tree canopies[41]) offer great potential. However, monitoring biodiversity using eDNA requires further development to overcome limitations such as biases[42] and the relationship between DNA biomass and abundance estimators[43].

Using RAS to monitor cryptic species where visibility is restricted (for instance, in dense vegetation or low light) poses additional problems for sensors. The utility of RAS is also affected by the thermoregulation mechanism of target taxa. Passive infrared detectors are widely used for endotherms, but other methods are required for ectotherms, such as bioacoustics[44] and image motion analysis[45]. Although flying UAVs generate sounds that may mask animal vocalizations, UAV-borne recorders have successfully recorded birds[46] and bats[47]. As RAS technology continues to develop quieter platforms, the use of UAVs in bioacoustic monitoring is likely to increase.

The potential for RAS to also monitor ecological processes such as predation and decomposition was perceived as important, with biodiversity experts reflecting that 'ecological function is about processes' and that 'it's not the abundance of a tree species or a seed disperser species that matters, but whether the tree species is regenerating'. RAS experts confirmed that this would be difficult to achieve but pointed to recent successes in the use of RAS to monitor pollination, albeit in a simplified system[48], and remote sensing of plant photosynthesis and primary productivity[49].

Biodiversity experts recognized the challenge of performing RAS surveys while minimizing disturbance of species and habitats[31]. In the case of UAV-based surveys, disturbance of species can be caused by the shape of the UAV and its approach distance, airspeed, and flight pattern, as well as pilot competence and noise[50]. However, it was acknowledged that surveys by humans also cause disturbance[51] and that '[there are] likely to be pros and cons for disturbance from humans versus robots'. RAS experts agreed that 'aerial vehicles are noisy and many wheeled terrestrial vehicles can be destructive in terms of trampling' but noted that the key to developing solutions lies in defining the criteria and thresholds for no or low disturbance to species or habitats[50].

### Barrier category 3: data handling and processing

Ecologists often need to survey biodiversity over many days, months or years, rapidly generating large data volumes[52]. One biodiversity expert stated that 'storage for extreme volumes of data is a top priority

https://doi.org/10.1038/s41559-025-02704-9

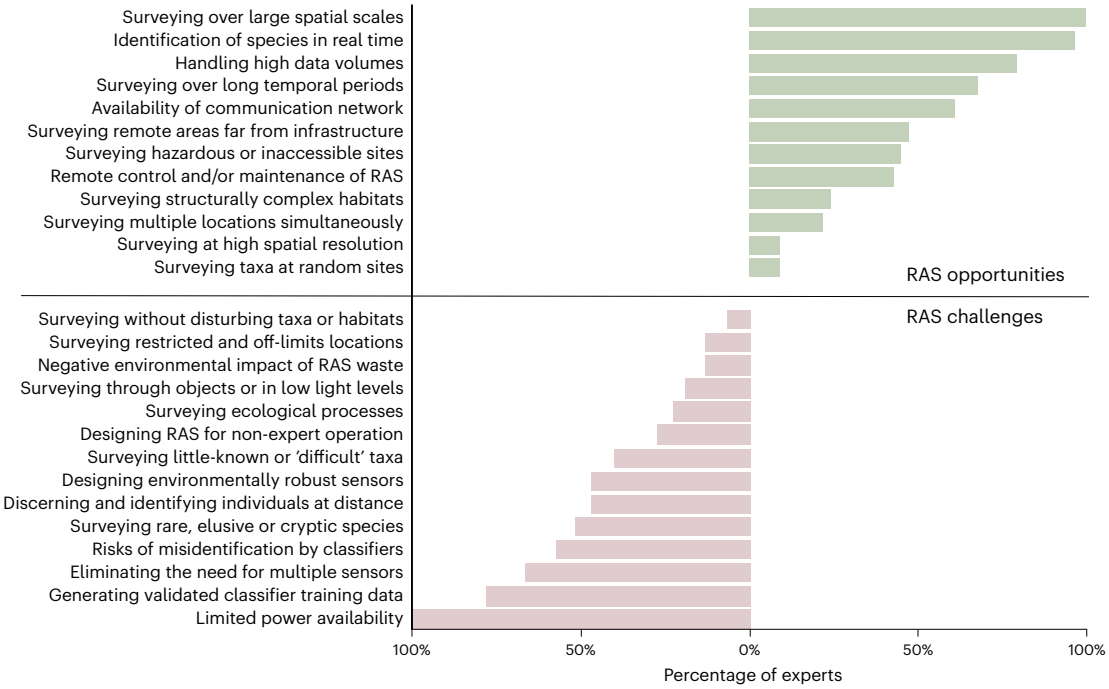



**Fig. 4 | Opportunities and challenges associated with using RAS to monitor terrestrial biodiversity.** These were identified by biodiversity experts during stage 2 of the modified Delphi technique. Each expert was allowed to select up to three opportunities and challenges that they believed would have the most profound impact on an 'ideal' survey.

in the bioacoustic monitoring field.' RAS experts highlighted several technologies that could help. The most commonly used method is edge processing, in which AI computations are used to preprocess data and reduce storage and data transmission requirements[53]. Other suggestions included AI prioritization of data storage based on sampling variation, using lossless data compression techniques[54], and optimizing data storage using a wireless sensor network[55] or data-mule drones[56] to offload data from sensors. Alternatively, data transmission to cloud storage for subsequent offline processing would be possible if RAS could access a communication network. RAS experts emphasized good preparation before sampling so only relevant data are collected.

Recent advances in sound- and image-based biodiversity identification app technology (for instance, https://merlin.allaboutbirds.org, https://plantnet.org) were noted by several biodiversity experts. Virtually all biodiversity experts thought real-time species identification would be an opportunity associated with RAS but also recognized major challenges associated with automated identification. For example, three-quarters (Fig. 4) highlighted the lack of classifier training data and expert validation for most taxa, and more than half foresaw potential risks arising from species misidentification[57]. Additional concerns were raised regarding the accessibility of machine learning methods for non-experts, data ownership, lack of open-access data and data protection.

For some taxa, declining numbers of taxonomists will hamper the verification of species' training data[58]. One biodiversity expert stated 'you can't replace the value of expert interpretation of species, management, habitats, and context'. However, others countered this saying that expert opinions can be fallible[59]. Indeed, a prerequisite of automated analysis is a huge library of expert-certified species images (or sounds), and 'classifiers need to be trained with samples that are geographically relevant'. Some biodiversity experts expressed doubts about compiling suitable datasets, as 'training data tend to be biased towards well sampled areas/groups'. This poses a particular problem for rare, elusive and cryptic species, which was seen as a challenge for RAS to address. Moreover, many biodiversity experts expressed doubts over monitoring little-known and 'difficult' taxa, emphasizing that classifiers 'must be able to recognize when the species is unknown. For instance, not within its training set'. Another apprehension was that classifier identification errors might lead to threatened species being given incorrect IUCN conservation status[60]. It was therefore seen as important that human experts should evaluate error rates of AI species identification, with a consequent need to store raw data for independent validation.

There are few solutions currently available to overcome the difficulties associated with compiling huge annotated datasets for automated species identification. One technique suggested by RAS experts was the use of machine learning approaches that employ techniques with reduced data requirements, such as 'few-shot learning'[61]. For instance, limited real data, supplemented by data augmentation[62] with simulated data, could be used to identify large mammal species in camera trap images[63]. However, few-shot learning techniques applied without adequate validation can lead to serious misrepresentations of biodiversity[64].

### Barrier category 4: power and network availability

Power availability was recognized by all biodiversity experts as a major issue related to monitoring of terrestrial species. They also remarked that some of the RAS challenges were interlinked. For example, whereas edge processing of sensor data and communication network capability could minimize data storage, it may increase RAS power requirements. The ability to control and maintain RAS equipment remotely was identified as an opportunity by biodiversity experts. Maintaining sensors is 'very challenging, even in urban environments', and surveyors 'often need direct access for maintenance [and] signal proximity to control software'. RAS experts noted that this capability could be provided, but that 'networks lack traceability of where the issues arise', and that 'the internet of things is not that mature'.

Although battery technologies have advanced rapidly, battery-powered robots and sensors generally have short operational lives before needing to recharge. Biodiversity experts undertaking monitoring during the winter observed that 'cold can drain power

https://doi.org/10.1038/s41559-025-02704-9

**Table 2 | Status of current RAS technology available for terrestrial biodiversity monitoring**

| Scope of required biodiversity monitoring | Status of current RAS technology | | | | |
|---|---|---|---|---|---|
| | Robot platform | Sensors | AI capability | Power source | Data handling |
| Autonomous robotic swarm for synchronized surveys of multiple sites | | | | | |
| Autonomous robotic system for species identification | | | | | |
| Monitoring of locations where human access is dangerous or impossible | | | | | |
| Monitoring of species at night and when visibility is restricted | | | | | |
| Precision position sensing to monitor exact locations repeatedly | | | | | |
| RAS built with sensors and/or probes resilient in all environmental conditions | | | | | |
| RAS capable of self-reporting malfunctions and remote control | | | | | |
| RAS network communication for real-time and/or cloud data processing | | | | | |
| Survey of sites repeatedly during a year or over many years | | | | | |
| Ability to deal with extreme data volumes from sensors | | | | | |
| Ability to survey complex habitats and communicate through barriers | | | | | |
| Access to geographically-relevant classifier data for species identification | | | | | |
| Detection and identification of rare and/or cryptic species of different sexes or life stages | | | | | |
| Monitoring ecosystem functions and processes (for example, pollination, predation) | | | | | |
| Monitoring of multiple taxa over large areas | | | | | |
| Monitoring of taxa in distant locations with limited support | | | | | |
| RAS built from materials that reduce environmental impact of e-waste | | | | | |
| RAS equipment easy to use (for example, suitable for citizen scientists) | | | | | |
| Resolution and identification of individuals of small species from a distance | | | | | |
| Simultaneous identification of numerous species and individuals | | | | | |
| Ability to optimize survey site selection in relation to taxa | | | | | |
| Performance of monitoring without disturbance or need for destructive sampling | | | | | |
| Monitoring of rare species with poorly defined taxonomy and 'difficult' taxa | | | | | |
| RAS with few sensors to monitor taxa and/or species of all sizes in all habitats | | | | | |

Coloured shading indicates the main areas in which technological developments are needed to enable RAS to perform each monitoring task: grey, field-tested technology exists; sage green, technology exists, but substantial limitations need to be overcome; dark green, working prototypes exist; pink, technology is still in research and development phase; blue, major technological breakthrough required.

sources very, very quickly'. Batteries carried by RAS need to power the robotic movement, the sensor(s) and the controller with storage memory. As a result, the endurance of hovering UAVs is typically 20–40 min (ref. 65). The use of solar power was thought to be helpful by biodiversity experts, but they noted that it 'isn't great for high latitude winters', and 'solar is not viable for [the] understorey'. RAS experts identified several currently available technologies that may help address the challenge of powering RAS. Efficient energy consumption has been demonstrated in multimodal robots that combine aerial and terrestrial locomotion modes within one platform[66]. A similar approach has been adopted in a solar-powered robot that minimizes energy consumption in the manner of tree sloths by traversing wires slowly while performing long-term environmental monitoring[67]. RAS experts also suggested that sensors could use low-powered microchips for onboard computing and energy-efficient cameras to reduce energy needs. Another method would be to employ homing robotic systems that return to recharging hubs to prolong operating times[68]. Other possibilities for providing sustainable power include microbial fuel cells[69], harnessing rain[70], triboelectric nanogenerators for mechanical energy harvesting[71], thermoelectric energy harvesting from soil[72] and chemical energy[73]. Addressing environmental impact is complex, but RAS experts stated that rapid progress is being made in developing biodegradable batteries[74], sensors and soft robotic systems[75].

Overall, the assessment of biodiversity and RAS experts was that widespread adoption of RAS for monitoring biodiversity requires further technological development, and that some areas are likely to be addressed relatively easily, whereas others pose greater challenges (Table 2).

## Discussion

For common species of some taxa, including birds and mammals, RAS are already providing valuable survey data, and this capability is increasing. For these taxa, the main limitation is accurate identification of lesser-known species, for which geographically relevant classifier data may be lacking. In 'difficult' taxa such as fungi, this constraint poses a severe problem. However, the lack of classifier data is only one factor impeding the utility of RAS for biodiversity monitoring. This is because of the complex interrelationships between sensors and sensing techniques used to detect species; the management, communication and processing of sensor data; and the provision of power for these tasks, as well as for the robotic platform.

Many of the enabling technologies and capabilities needed for RAS to monitor terrestrial biodiversity effectively already exist, although they have often been developed for different applications[8–12,23–30]. Several types of robotic platform are already used in biodiversity surveys[11], and rapid development progress (for instance, for subterranean access[28,29]) suggests that this will not be the primary bottleneck. The critical limitations to overcome are sensors and sensing techniques[36–49] with classifier databases[57–60], where major breakthroughs are needed. Progress in these areas could rapidly advance accurate species identification across more taxa but will be dependent on new methods of processing large data volumes[52–54,61–64] in real time. Although not an immediate constraint, power source developments will become increasingly critical to sustain RAS autonomy as the capabilities of other components advance. Without enhanced power availability, RAS can only be deployed to monitor biodiversity for short time periods in some ecosystems and geographical regions. It is not possible to predict when such transformative breakthroughs may occur, but recent advances in power source technology[69–74] are cause for optimism.

Adapting RAS to new environments might be problematic, as considerable time and resources are required to create, service and support robust systems suited to working in uncontrolled conditions. Field-testing of RAS as fully integrated units for terrestrial biodiversity monitoring is a critical step in defining the boundaries of their capabilities. Given these constraints, it may initially be more efficient to deploy multiple stationary sensor systems rather than mobile RAS. This approach could provide the spatial coverage that mobile

robots offer, while avoiding many challenges associated with developing robust navigation and power management systems. Alternatively, readily available RAS could be more widely deployed for repetitive monitoring of well-known taxa and easily accessible ecosystems[12–14]. This could free human surveyor time to focus on specific taxa, habitats and ecosystems for which RAS are currently underdeveloped.

Despite the challenges, the development of RAS able to track changes in species abundance and community composition could deliver profound advances in conservation. In the present study, most biodiversity experts foresaw many opportunities associated with RAS but viewed them as additional tools to supplement rather than supplant existing survey methods. There was some hesitation about the suitability of RAS for certain taxa (for instance, those for which genomic data are needed for accurate identification[4]). One overarching issue was that RAS could quickly generate huge volumes of biodiversity data that could be used to inform policy and practice without critical evaluation. It is unclear whether taxonomic bias, with a focus on some species to the detriment of others[76], may increase or decrease with the use of RAS. Concerns were also raised regarding high costs, e-waste, ethical implications and diversion of resources from other conservation work. Nevertheless, RAS integrated into well-structured, goal-based programmes with standardized protocols could lead to major progress in monitoring of terrestrial biodiversity. As one biodiversity expert observed 'if [RAS] could monitor just 10% of species reliably across all taxonomic groups at appropriate scales and resurvey intervals, it would be a substantial improvement on current approaches'.

Genuinely transdisciplinary approaches to terrestrial biodiversity monitoring need to be fostered between biodiversity and RAS experts, so that ideas and technologies can be codeveloped effectively. Biodiversity experts generally have limited formal training in RAS and big data. Likewise, RAS experts do not routinely consider the complexity of biodiversity, ecosystem functioning and the practicalities associated with field-based monitoring. By promoting and funding cross-disciplinary collaboration aimed at adapting RAS for conservation applications, governments, philanthropists and organizations can drive major progress. One such example is the 'Natural Robotics Contest' (https://www.naturalroboticscontest.com/), an environmental robot design competition. In the longer term, education strategies at all levels should seek to establish and augment interdisciplinary thinking among aspiring engineers, ecologists and computer scientists[77]. This could be achieved by highlighting the major methodological challenges and need for improved technology to support terrestrial biodiversity monitoring in undergraduate engineering and computer science courses, as well as providing explanations of cutting-edge technological applications in ecology and conservation courses. Future generations of researchers may then be able to communicate and work together more readily, bridging the traditional disciplinary boundaries between ecology and engineering.

## Methods

We undertook our modified Delphi technique, a method that is applied widely in conservation and environmental sciences[15], between April and June 2023. The technique involves a structured and iterative survey of a group of participants that aims to capture a broad range and depth of contributions. It has several advantages over standard approaches to gathering opinions from groups of people. For example, participant contributions are anonymous, which minimizes potential biases resulting from social pressures such as groupthink, halo effects and the influence of dominant individuals[15].

Our Delphi approach comprised three stages: an online questionnaire and online workshop for biodiversity experts, followed by an online workshop for RAS experts (Fig. 1). Participants were asked to provide informed consent before participating in any of the activities. We made them aware that their involvement was entirely voluntary, that they could stop at any point and withdraw from the process without explanation, and that the data they provided via the questionnaire and workshop would be anonymous and unidentifiable. Ethical approval was granted by the School of Anthropology and Conservation Research Ethics Committee at the University of Kent (reference 394 2023).

### Stage 1: biodiversity expert online questionnaire

We used a mixed approach to recruiting biodiversity experts for stage 1 to minimize the likelihood of bias associated with relying on a single method. By using global professional networks and identifying authors of recent papers on monitoring of terrestrial taxa, we identified 334 experts from across the world. We also found an additional 154 experts by contacting relevant research institutes, non-governmental organizations and conservation agencies, and by snowball sampling (invitees suggesting other biodiversity experts who might be interested in participating). Our aim was to recruit experts with experience of biodiversity surveys in a diverse range of biomes and covering all terrestrial taxa (Fig. 2 and Extended Data Figs. 1 and 2). Of the 488 biodiversity experts (35% women) in 43 countries who were invited, 98 experts (33% women) in 24 countries took part in stage 1.

The questionnaire was delivered using the online platform Qualtrics (https://qualtrics.com). We asked participants to list their country of residence; their employment sector; their experience of monitoring taxa, habitats, and ecosystems; and the countries in which they had conducted or facilitated terrestrial biodiversity monitoring. We asked participants to detail an 'ideal' biodiversity survey that was not limited by funding or logistics. We did not mention RAS and how it might be incorporated into surveys to ensure that participants were not influenced by their understanding of the capabilities and limitations of RAS. Participants were asked to specify which terrestrial taxa and ecosystems their monitoring would focus on and the methodological barriers that would need to be overcome to make the survey possible. We piloted and pretested the questionnaire content, which helped us to refine the wording of questions and definitions of terminology. We used an inductive approach to analyse the qualitative questionnaire responses. By synthesizing participant statements, we collated the data into four broad barriers (Fig. 3), which were the basis of discussion in stage 2.

### Stage 2: biodiversity expert online workshops

The same group of 98 biodiversity experts were invited to take part in an online workshop, organized on Teams, which aimed to assess the potential for RAS to resolve the barriers articulated in stage 1. Seven participants who had completed the questionnaire did not continue to stage 2. The remaining 91 participants (34% women) were allocated to one of three groups according to whether they self-identified as experts in surveying vertebrates ($n = 36$ participants); invertebrates ($n = 26$); or trees, plants and/or fungi ($n = 29$). Separate workshops were held for each group simultaneously, with each workshop following the same format.

Each workshop opened with a summary of the overall project and its aim, as well as a description of planned workshop activities. We presented the barriers in written format using Padlet (https://padlet.com/), a collaborative web platform where participants can access, upload and organize shared content. We asked participants to consider the opportunities and challenges that RAS offer with respect to overcoming these barriers within each of our four broad barriers (site access, species and individual detection, data handling and processing, and power and network availability; Table 1). We asked participants to clarify, expand, join or add new barriers wherever they felt necessary and to comment on the relevance and appropriateness of the RAS opportunities and/or challenges that emerged. Finally, for each of the four broad categories of barrier, we asked participants to select up to three RAS opportunities and challenges that they believed would have the most profound impact on their 'ideal' survey (Fig. 4).

### Stage 3: RAS expert online workshop

We used a mixed approach to recruit RAS experts to participate in our RAS online workshop. Our objective was to include global experts working at the forefront of RAS applications and development, including those working on closely related technologies such as sensors, AI and machine learning. Relevant experts were identified among authors of recent papers, from professional networks and mailing lists (for instance, the UK-RAS network), and by snowball sampling. Using this information, we emailed 196 experts (21% women) in 24 countries, inviting them to participate in an online workshop, organized on Teams, to discuss the applications of RAS to terrestrial biodiversity monitoring. A pool of 31 RAS experts (26% women) from eight countries took part. The smaller number of experts taking part in this workshop, compared with the biodiversity workshop, reflected the wide range of taxon, biome and global expertise we required from biodiversity specialists.

We began the workshop with an introduction to biodiversity, ecosystems and monitoring methods currently used to survey different taxa. This was followed by discussions of the barriers that had been identified by the biodiversity experts in stage 2. The barriers were grouped into the same four broad categories that had been used previously. RAS experts were asked to identify existing RAS capabilities that were directly transferable to a terrestrial biodiversity monitoring context, as well as nascent technologies or new ideas that might be relevant for the future. Again, we used an inductive approach to analyse the qualitative data derived from the workshop. This enabled us to determine existing RAS capabilities that are closely aligned with biodiversity monitoring requirements, how these capabilities could be extended and potential priorities for future RAS developments.

### Reporting summary

Further information on research design is available in the Nature Portfolio Reporting Summary linked to this article.

## Data availability

The anonymized dataset generated and analysed during this study is available[78] via the University of Kent Data Repository at https://doi.org/10.22024/UniKent/01.01.546.

## References


1. Navarro, L. M. et al. Monitoring biodiversity change through effective global coordination. *Curr. Opin. Environ. Sustainability* **29**, 158–169 (2017).
2. Hochkirch, A. et al. A multitaxon analysis of European red lists reveals major threats to biodiversity. *PLoS ONE* **18**, e0293083 (2023).
3. Pardikes, N. A., Shapiro, A. M., Dyer, L. A. & Forister, M. L. Global weather and local butterflies: variable responses to a large-scale climate pattern along an elevational gradient. *Ecology* **96**, 2891–2901 (2015).
4. Kallimanis, A. S. et al. Efficient biodiversity monitoring: which taxonomic level to study? *Ecol. Indic.* **15**, 100–104 (2012).
5. Thompson, W. L. *Sampling Rare or Elusive Species: Concepts, Designs and Techniques for Estimating Population Parameters* (Island Press, 2004).
6. Marvin, S., While, A. H., Kovacic, M., Lockhart, A. & Macrorie R. *Urban Robotics and Automation* (EPSRC UK-RAS Network, 2018); https://doi.org/10.31256/WP2018.1
7. Schranz, M., Umlauft, M., Sende, M. & Elmenreich, W. Swarm robotic behaviours and current applications. *Front. Rob. AI* **7**, 00036 (2020).
8. Duckett, T., Pearson, S., Blackmore, S., & Grieve, B. *Agricultural Robotics: The Future of Robotic Agriculture* (EPSRC UK-RAS Network, 2018); https://doi.org/10.31256/WP2018.2
9. Richardson, R., et al. *Robotic and Autonomous Systems for Resilient Infrastructure* (EPSRC UK-RAS Network, 2017); https://doi.org/10.31256/WP2017.2
10. Bergeles, C., Cruz Ruiz, A., Rodriguez y Baena, F. & Valdastri, P. *Surgical Robotics* (EPSRC UK-RAS Network, 2021); https://doi.org/10.31256/WP2021.2
11. Verfuss, U. K. et al. A review of unmanned vehicles for the detection and monitoring of marine fauna. *Mar. Pollut. Bull.* **140**, 17–29 (2019).
12. Stephenson, P. J. Integrating remote sensing into wildlife monitoring for conservation. *Environ. Conserv.* **46**, 181–183 (2019).
13. Corcoran, E., Winsen, M., Sudholz, A. & Hamilton, G. Automated detection of wildlife using drones: synthesis, opportunities and constraints. *Methods Ecol. Evol.* **12**, 1103–1114 (2021).
14. Besson, M. et al. Towards the fully automated monitoring of ecological communities. *Ecol. Lett.* **25**, 2753–2775 (2022).
15. Mukherjee, N. et al. The Delphi technique in ecology and biological conservation: applications and guidelines. *Methods Ecol. Evol.* **6**, 1097–1109 (2015).
16. Haidegger, T. et al. Robotics: enabler and inhibitor of the Sustainable Development Goals. *Sustain. Prod. Consum.* **43**, 422–434 (2023).
17. Brown, G. P. & Shine, R. Influence of weather conditions on activity of tropical snakes. *Austral. Ecol.* **27**, 596–605 (2002).
18. Angelini, F. et al. Robotic monitoring of habitats: the natural intelligence approach. *IEEE Access* **11**, 72575–72591 (2023).
19. Malladi, M. V. R. et al. Tree instance segmentation and traits estimation for forestry environments exploiting LiDAR data collected by mobile robots. In *2024 IEEE International Conference on Robotics and Automation (ICRA)* 17933–17940 (IEEE, Yokohama, Japan, 2024). https://doi.org/10.1109/ICRA57147.2024.10611169
20. Lee, J., Hwangbo, J., Wellhausen, L., Koltun, V. & Hutter, M. Learning quadrupedal locomotion over challenging terrain. *Sci. Rob.* **5**, eabc5986 (2020).
21. Hamaza, S., Georgilas, I., Heredia, G., Ollero, A. & Richardson, T. Design, modelling, and control of an aerial manipulator for placement and retrieval of sensors in the environment. *J. Field Rob.* **37**, 1224–1245 (2020).
22. Suryawanshi, K. et al. Estimating snow leopard and prey populations at large spatial scales. *Ecol. Solut. Evid.* **2**, e12115 (2021).
23. Tsitsimpelis, I., Taylor, C. J., Lennox, B. & Joyce, M. J. A review of ground-based robotic systems for the characterization of nuclear environments. *Prog. Nucl. Energy* **111**, 109–124 (2019).
24. Shang, J., Sattar, T., Chen, S., Bridge, B. & Madsen, O. Design of a climbing robot for inspecting aircraft wings and fuselage. *Ind. Robot* **34**, 495–502 (2007).
25. Miriyev, A., Stack, K. & Lipson, H. Soft material for soft actuators. *Nat. Commun.* **8**, 596 (2017).
26. Stokkeland, M., Klausen, K. & Johansen, T. A. Autonomous visual navigation of unmanned aerial vehicle for wind turbine inspection. In *2015 International Conference on Unmanned Aircraft Systems (ICUAS)*, 998–1007 (IEEE, 2015). https://doi.org/10.1109/ICUAS.2015.7152389
27. Lam, T. L. & Xu, Y. A flexible tree climbing robot: Treebot-design and implementation. In *2011 IEEE International Conference on Robotics and Automation* 5849–5854 (IEEE, Shanghai, China, 2011). https://doi.org/10.1109/ICRA.2011.5979833
28. Chen, S. et al. Soft crawling robots: design, actuation, and locomotion. *Adv. Mater. Technol.* **5**, 1900837 (2019).
29. Rus, D. & Tolley, M. Design, fabrication and control of soft robots. *Nature* **521**, 467–475 (2015).
30. Häusermann, D. et al. FireDrone: multi-environment thermally agnostic aerial robot. *Adv. Intell. Syst.* **5**, 2300101 (2023).

31. Millner, N. et al. Exploring the opportunities and risks of aerial monitoring for biodiversity conservation. *Glob. Soc. Chall. J.* **2**, 2–23 (2023).
32. Sandbrook, C. et al. Principles for the socially responsible use of conservation monitoring technology and data. *Conserv. Sci. Pract.* **3**, e374 (2021).
33. Laidre, M. E. Ruler of the atoll: the world's largest land invertebrate. *Front. Ecol. Environ.* **15**, 527–528 (2017).
34. Patel, M. & Reddy, M. N. Discovery of the world's smallest terrestrial pteridophyte. *Sci. Rep.* **8**, 5911 (2018).
35. Shenkin, A. et al. The world's tallest tropical tree in three dimensions. *Front. For. Glob. Change* **2**, 00032 (2019).
36. Wäldchen, J., Rzanny, M., Seeland, M. & Mäder, P. Automated plant species identification—trends and future directions. *PLoS Comput. Biol.* **14**, e1005993 (2018).
37. Wägele, J. W. et al. Towards a multisensory station for automated biodiversity monitoring. *Basic Appl. Ecol.* **59**, 105–138 (2022).
38. Sugai, L. S. M., Silva, T. S. F., Ribeiro, J. W. & Llusia, D. Terrestrial passive acoustic monitoring: review and perspectives. *BioScience* **69**, 15–25 (2019).
39. Wilson, A. D. Diverse applications of electronic-nose technologies in agriculture and forestry. *Sensors* **13**, 2295–2348 (2013).
40. Taberlet, P., Bonin, A., Zinger, L. & Coissac, E. *Environmental DNA: For Biodiversity Research and Monitoring* (Oxford Univ. Press, 2018)
41. Aucone, E. et al. Drone-assisted collection of environmental DNA from tree branches for biodiversity monitoring. *Sci. Rob.* **8**, eadd5762 (2023).
42. Zinger, L. et al. DNA metabarcoding – need for robust experimental designs to draw sound ecological conclusions. *Mol. Ecol.* **28**, 1857–1862 (2019).
43. Jo, T. & Yamanaka, H. Fine-tuning the performance of abundance estimation based on environmental DNA (eDNA) focusing on eDNA particle size and marker length. *Ecol. Evol.* **12**, e9234 (2022).
44. Mutanu, L., Gohil, J., Gupta, K., Wagio, P. & Kotonya, G. A review of automated bioacoustics and general acoustics classification research. *Sensors* **22**, 8361 (2022).
45. Corva, D. M. et al. A smart camera trap for detection of endotherms and ectotherms. *Sensors* **22**, 4094 (2022).
46. Fischer, S., Edwards, A. C., Garnett, S. T., Whiteside, T. G. & Weber, P. Drones and sound recorders increase the number of bird species identified: a combined surveys approach. *Ecol. Inform.* **74**, 101988 (2023).
47. Kloepper, L. N. & Kinniry, M. Recording animal vocalizations from a UAV: bat echolocation during roost re-entry. *Sci. Rep.* **8**, 7779 (2018).
48. Ratnayake, M. N., Dyer, A. G. & Dorin A. Towards computer vision and deep learning facilitated pollination monitoring for agriculture. In *Proc. IEEE/CVF Conference on Computer Vision and Pattern Recognition (CVPR) Workshops* (eds Masson-Forsythe, M. & Mortensen, E.) 2921–2930 (IEEE, 2021). https://doi.org/10.1109/CVPRW53098.2021.00327
49. Porcar-Castell, A. et al. Linking chlorophyll *a* fluorescence to photosynthesis for remote sensing applications: mechanisms and challenges. *J. Exp. Bot.* **65**, 4065–4095 (2014).
50. Mo, M. & Bonatakis, K. Approaching wildlife with drones: using scientific literature to identify factors to consider for minimising disturbance. *Aust. Zool.* **42**, 1–29 (2022).
51. Phillips, O. L., Nuñez, V. P. & Timaná, M. E. Tree mortality and collecting botanical vouchers in tropical forests. *Biotropica* **30**, 298–305 (1998).
52. Kelling, S. et al. Data-intensive science: a new paradigm for biodiversity studies. *BioScience* **59**, 613–620 (2009).
53. Mansouri, Y. & Ali Babar, M. A review of edge computing: features and resource virtualization. *J. Parallel Distrib. Comput.* **150**, 155–183 (2021).
54. Sharma, K. & Gupta, K. Lossless data compression techniques and their performance. In *2017 International Conference on Computing, Communication and Automation* (ICCCA), 256–261 (IEEE, Greater Noida, India, 2017). https://doi.org/10.1109/CCAA.2017.8229810
55. Badescu, A. & Cotofana, L. A wireless sensor network to monitor and protect tigers in the wild. *Ecol. Indic.* **57**, 447–451 (2015).
56. Bhadauria, D., Tekdas, O. & Isler, V. Robotic data mules for collecting data over sparse sensor fields. *J. Field Rob.* **28**, 388–404 (2011).
57. Perez-Granados, C. BirdNET: applications, performance, pitfalls and future opportunities. *Ibis* **165**, 1068–1075 (2023).
58. Löbl, I., Klausnitzer, B., Hartmann, M. & Krell, F.-T. The silent extinction of species and taxonomists – an appeal to science policymakers and legislators. *Diversity* **15**, 1053 (2023).
59. Johansson, Ö. et al. Identification errors in camera-trap studies result in systematic population overestimation. *Sci. Rep.* **10**, 6393 (2020).
60. The IUCN Red List of Threatened Species. Version 2025-1. *IUCN* www.iucnredlist.org (accessed 6 February 2025).
61. Wang, Y., Yao, Q., Kwok, J. & Ni, L. M. Generalizing from a few examples: a survey on few-shot learning. *ACM Comput. Surv.* **53**, 1–34 (2020).
62. Shorten, C. & Khoshgoftaar, T. M. A survey on image data augmentation for deep learning. *J. Big Data* **6**, 60 (2019).
63. Beery, S. et al. Synthetic examples improve generalization for rare classes. In *Proc. IEEE/CVF Winter Conference on Applications of Computer Vision* 863–873 (IEEE, Piscataway, NJ, 2020). https://doi.org/10.1109/WACV45572.2020.9093570
64. Luccioni, A. S. & Rolnick, D. Bugs in the data: how ImageNet misrepresents biodiversity. *Proc. AAAI Conf. Artif. Intell.* **37**, 14382–14390 (2023).
65. Jawad, A. M. et al. Wireless power transfer with magnetic resonator coupling and sleep/active strategy for a drone charging station in smart agriculture. *IEEE Access* **7**, 139839–139851 (2019).
66. Ramirez, J. P. & Hamaza, S. Multimodal locomotion: next generation aerial-terrestrial mobile robotics. *Adv. Intell. Syst.* https://doi.org/10.1002/aisy.202300327 (2023).
67. Notomista, G., Emam, Y. & Egerstedt, M. The SlothBot: a novel design for a wire-traversing robot. *IEEE Rob. Autom. Lett.* **4**, 1993–1998 (2019).
68. Ravankar, A. & Ravankar, A. Distributed docking station system for mobile robots. In *Proc. JSME Annual Conference on Robotics and Mechatronics (Robomec)* 2P2-M06 (The Japan Society of Mechanical Engineers, 2022). https://doi.org/10.1299/jsmermd.2022.2P2-M06
69. Slate, A. J., Whitehead, K. A., Brownson, D. A. C. & Banks, C. E. Microbial fuel cells: an overview of current technology. *Renew. Sustain. Energy Rev.* **101**, 60–81 (2019).
70. Ilyas, M. A. & Swingler, J. Piezoelectric energy harvesting from raindrop impacts. *Energy* **90**, 796–806 (2015).
71. Wu, C., Wang, A. C., Ding, W., Guo, H. & Wang, Z. L. Triboelectric nanogenerator: a foundation of the energy for the new era. *Adv. Energy Mater.* **9**, 1802906 (2019).
72. Huang, Y., Xu, D., Kan, J. & Li, W. Study on field experiments of forest soil thermoelectric power generation devices. *PLoS ONE* **14**, e0221019 (2019).
73. Chu, S., Cui, Y. & Liu, N. The path towards sustainable energy. *Nat. Mater.* **16**, 16–22 (2017).
74. Lee, M. H. et al. A biodegradable secondary battery and its biodegradation mechanism for eco-friendly energy storage systems. *Adv. Mater.* **33**, 2004902 (2021).
75. Sethi, S. S., Kovac, M., Wiesemüller, F., Miriyev, A. & Boutry, C. M. Biodegradable sensors are ready to transform autonomous ecological monitoring. *Nat. Ecol. Evol.* **6**, 1245–1247 (2022).
76. Hughes, A. C. et al. Sampling biases shape our view of the natural world. *Ecography* **44**, 1259–1269 (2021).

77. Carey, C. C. et al. Enhancing collaboration between ecologists and computer scientists: lessons learned and recommendations forward. *Ecosphere* **10**, e02753 (2019).
78. Pringle, S. et al. RAS for biodiversity monitoring modified Delphi. *University of Kent Data Repository* https://doi.org/10.22024/UniKent/01.01.546 (2024).
79. Keith, D. A., Ferrer-Paris, J. R., Nicholson, E. & Kingsford, R. T. (eds). *The IUCN Global Ecosystem Typology 2.0: Descriptive Profiles for Biomes and Ecosystem Functional Groups* (IUCN, 2020).

## Acknowledgements

We thank all questionnaire and workshop participants. In particular, we remember Professor Ibrahim B. Yakubu, who played a full part in the questionnaire and workshop stages but sadly passed away midway through the evolution of this manuscript. This work was funded by the EPSRC UK-RAS Network. D.J.I. is funded by a UK Research and Innovation Future Leaders Fellowship (grant ref: MR/W006316/1), and Z.G.D. and J.C.F. were supported by Research England's 'Expanding Excellence in England' fund. The funders had no role in study design, data collection and analysis, decision to publish or preparation of the manuscript.

## Author contributions

M.A.G., E.H., S.J.L. and Z.G.D. conceived the study. S. Pringle., M. Dallimer, M.A.G., L.K.L.G., E.H., S.J.L. and Z.G.D. developed and tested the questionnaire and workshop materials but did not contribute data. M. Dallimer, J.C.F., M.A.G. and Z.G.D. led the workshops. S. Pringle collated and analysed the questionnaire and workshop data. S. Pringle and J.C.F. arranged data curation. S. Pringle, M. Dallimer, M.A.G. and Z.G.D. wrote the first draft of the paper and contributed to, edited and agreed the submitted version. S.-A.A., M.A., F.A., F.A.C., G.E.A., J.J.B., K.C.R.B., L.F.B., C.B.-L., A.S.B., R.B., A.J.B., J.E.B., J.B, P.J., E.R.B., S.J.B., D.C., C.F.C., A.C., K.F.D., N.J.D., M. Dodd, R.D., D.A.D., G.D., M. Dyrmann, D.P.D., M.S.F., A.F., R. Field, J.C.F., R.J.F., C.W.F., R. Fox, R.M.F., A.M.A.F., A.M.G., C.J.G., I.G., R.A.G., S.H., M. Hanheide, M.W.H., M. Hedblom, T.H., S.P.H., K.A.H., E.R.H., D.J.I., G.J.-M., K.J., T.H.K., L.N.K., S.K.-S., J. Labisko, F.L., J. Lawson, N.L., R.F.d.L., N.A.L., H.H.M., G.L.M., L.C.M., E.M., B.M., A. McConnell, B.A.M., A. Miriyev, E.D.N., A.O., S. Papworth, C.L.P., A.P.-P., G.P., N.P., R.P., S.G.P., M.T.P.-M., L.Q., P.R.-P., S.J.R., M.R., H.R, J.P.S., C.J.S., A. Sanyal, F.S., S.S.S., A. Shabrani, R.S., S.C.S., R.P.H.S., C.D.S., M.C.S., P.A.S., P.J.S., M.J.S., M. Studley, M. Svátek, G.T., N.K.T., K.D.L.U., R.J.W., P.J.C.W., M.J.W., S.W., C.D.W., I.B.Y., N.Y., S.A.R.Z., A.Z. and J.A.Z. contributed questionnaire and/or workshop data and contributed to, edited and agreed the submitted paper. Z.G.D. administered the project. M. Dallimer, M.A.G., L.K.L.G., E.H., S.J.L. and Z.G.D. led funding acquisition.

## Competing interests

The authors declare no competing interests.

## Additional information

**Extended data** is available for this paper at https://doi.org/10.1038/s41559-025-02704-9.

**Supplementary information** The online version contains supplementary material available at https://doi.org/10.1038/s41559-025-02704-9.

**Correspondence and requests for materials** should be addressed to Stephen Pringle.

**Peer review information** *Nature Ecology & Evolution* thanks Holger Klinck and the other, anonymous, reviewer(s) for their contribution to the peer review of this work.

**Reprints and permissions information** is available at www.nature.com/reprints.

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

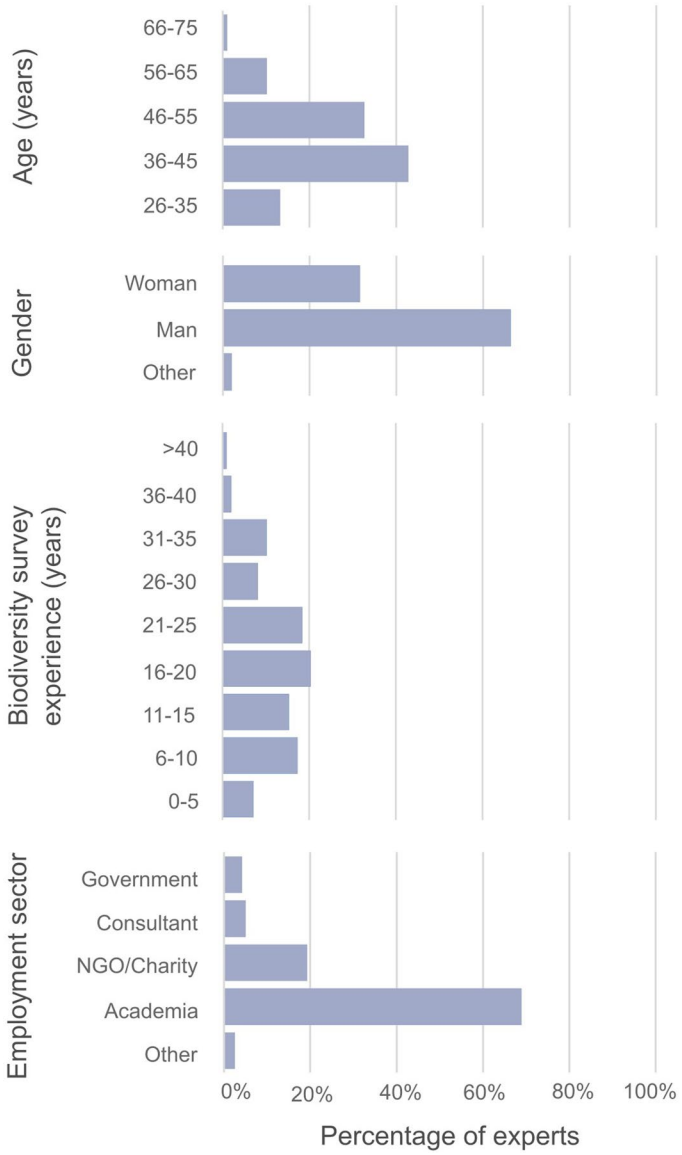
© The Author(s) 2025

**Stephen Pringle[1] ✉, Martin Dallimer[2], Mark A. Goddard[3], Léni K. Le Goff[4], Emma Hart[4], Simon J. Langdale[5], Jessica C. Fisher[1], Sara-Adela Abad[6], Marc Ancrenaz[7], Fabio Angeoletto[8], Fernando Auat Cheein[9], Gail E. Austen[1], Joseph J. Bailey[10], Katherine C. R. Baldock[3], Lindsay F. Banin[11], Cristina Banks-Leite[12], Aliyu S. Barau[13], Reshu Bashyal[14], Adam J. Bates[15], Jake E. Bicknell[1], Jon Bielby[16], Petra Bosilj[17], Emma R. Bush[18], Simon J. Butler[19], Dan Carpenter[20], Christopher F. Clements[21], Antoine Cully[22], Kendi F. Davies[23], Nicolas J. Deere[1], Michael Dodd[24], Rosie Drinkwater[25], Don A. Driscoll[26], Guillaume Dutilleux[27], Mads Dyrmann[28], David P. Edwards[29], Mohammad S. Farhadinia[1], Aisyah Faruk[30], Richard Field[31], Robert J. Fletcher[32], Chris W. Foster[33], Richard Fox[34], Richard M. Francksen[35], Aldina M. A. Franco[36], Alison M. Gainsbury[37], Charlie J. Gardner[1], Ioanna Giorgi[38], Richard A. Griffiths[1], Salua Hamaza[39], Marc Hanheide[17], Matt W. Hayward[40], Marcus Hedblom[41], Thorunn Helgason[42], Sui P. Heon[43], Kevin A. Hughes[44], Edmund R. Hunt[45], Daniel J. Ingram[1], George Jackson-Mills[46], Kelly Jowett[47], Timothy H. Keitt[48], Laura N. Kloepper[49], Stephanie Kramer-Schadt[50], Jim Labisko[51], Frédéric Labrosse[52], Jenna Lawson[53], Nicolas Lecomte[54], Ricardo F. de Lima[55], Nick A. Littlewood[56], Harry H. Marshall[57], Giovanni L. Masala[38], Lindsay C. Maskell[58], Eleni Matechou[59], Barbara Mazzolai[60], Alistair McConnell[61], Brett A. Melbourne[23], Aslan Miriyev[62], Eric Djomo Nana[63], Alessandro Ossola[64], Sarah Papworth[65], Catherine L. Parr[66], Ana Payo-Payo[67], Gad Perry[68], Nathalie Pettorelli[69], Rajeev Pillay[70], Simon G. Potts[71], Miranda T. Prendergast-Miller[3], Lan Qie[72], Persie Rolley-Parnell[73], Stephen J. Rossiter[74], Marcus Rowcliffe[69], Heather Rumble[75], Jon P. Sadler[76], Christopher J. Sandom[77], Asiem Sanyal[78], Franziska Schrodt[31],**

**Sarab S. Sethi[79], Adi Shabrani[80], Robert Siddall[81], Simón C. Smith[4], Robbert P. H. Snep[82], Carl D. Soulsbury[72], Margaret C. Stanley[83], Philip A. Stephens[84], P. J. Stephenson[85], Matthew J. Struebig[1], Matthew Studley[86], Martin Svátek[87], Gilbert Tang[88], Nicholas K. Taylor[89], Kate D. L. Umbers[90], Robert J. Ward[91], Patrick J. C. White[92], Mark J. Whittingham[93], Serge Wich[16], Christopher D. Williams[16], Ibrahim B. Yakubu[94], Natalie Yoh[1], Syed A. R. Zaidi[95], Anna Zmarz[96], Joeri A. Zwerts[97] & Zoe G. Davies[1]**

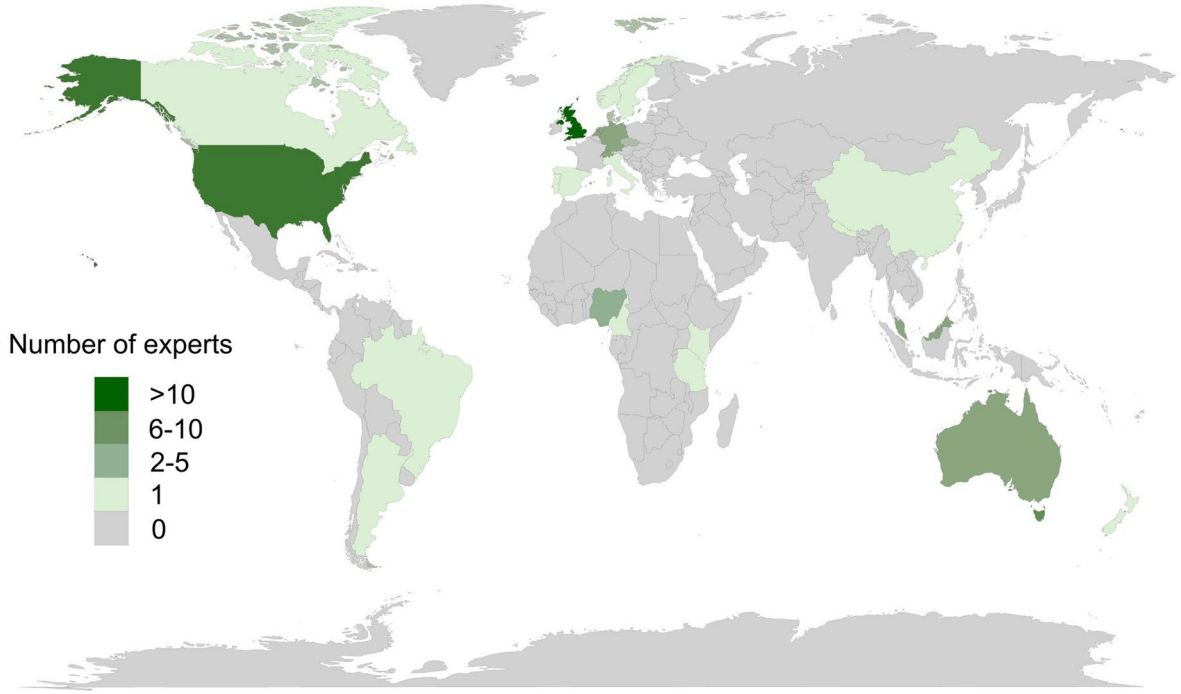
[1]Durrell Institute of Conservation and Ecology (DICE), School of Natural Sciences, University of Kent, Canterbury, UK. [2]Centre for Environmental Policy, Imperial College London, London, UK. [3]Department of Geography and Environmental Sciences, Northumbria University, Newcastle upon Tyne, UK. [4]School of Computing, Engineering and the Built Environment, Edinburgh Napier University, Edinburgh, UK. [5]Synthotech Ltd, Milner Court, Hornbeam Square, Harrogate, UK. [6]Mechanical Engineering Department, University College London, London, UK. [7]HUTAN, SWD, Kota Kinabalu, Malaysia. [8]Programa de Pós-Graduação em Gestão e Tecnologia Ambiental da Universidade Federal de Rondonópolis, Rondonópolis, Brasil. [9]Department of Engineering, Harper Adams University, Newport, UK. [10]Applied Ecology Research Group, School of Life Sciences, Anglia Ruskin University, Cambridge, UK. [11]Centre for Ecology and Hydrology, Penicuik, UK. [12]Department of Life Sciences, Imperial College London, Silwood Park Campus, Ascot, UK. [13]Department of Urban and Regional Planning, Faculty of Earth and Environmental Sciences, Bayero University, Kano, Nigeria. [14]Greenhood Nepal, Kathmandu, Nepal. [15]Animal, Rural & Environmental Sciences, Nottingham Trent University, Nottinghamshire, UK. [16]School of Biological and Environmental Sciences, Liverpool John Moores University, Liverpool, UK. [17]Lincoln Centre for Autonomous Systems, University of Lincoln, Lincoln, UK. [18]Royal Botanic Garden Edinburgh, Edinburgh, UK. [19]School of Biological Sciences, University of East Anglia, Norwich Research Park, Norwich, UK. [20]Digital Ecology Limited, Bristol, UK. [21]School of Biological Sciences, University of Bristol, Bristol, UK. [22]Department of Computing, Imperial College London, London, UK. [23]Department of Ecology and Evolutionary Biology, University of Colorado, Boulder, CO, USA. [24]Faculty of Science Technology, Engineering and Mathematics (STEM), The Open University, Walton Hall, Milton Keynes, UK. [25]Palaeogenomics Group, Faculty of Veterinary Medicine, Ludwig Maximilian University Munich, Munich, Germany. [26]School of Life and Environmental Sciences, Deakin University, Melbourne Burwood Campus, Australia. [27]Acoustics Group, Department of Electronic Systems, Norwegian University of Science and Technology, Trondheim, Norway. [28]AI Lab ApS, Maden 3, Aarhus V, Denmark. [29]Department of Plant Sciences and Centre for Global Wood Security, University of Cambridge, Cambridge, UK. [30]Royal Botanic Gardens Kew, Millennium Seed Bank, Wakehurst Place, Ardingly, UK. [31]School of Geography, University of Nottingham, Nottingham, UK. [32]Department of Zoology, University of Cambridge, Cambridge, UK. [33]School of Biological Sciences, University of Reading, Reading, UK. [34]Butterfly Conservation, Manor Yard, East Lulworth, UK. [35]Institute of Science and Environment, University of Cumbria, Carlisle, UK. [36]School of Environmental Sciences, University of East Anglia, Norwich, UK. [37]Department of Integrative Biology, University of South Florida, St Petersburg, FL, USA. [38]School of Computing, University of Kent, Canterbury, UK. [39]Biomorphic Intelligence Lab, Department of Control & Operations, Faculty of Aerospace Engineering, TU Delft, Delft, The Netherlands. [40]Centre for Conservation Science, University of Newcastle, Callaghan, Newcastle, New South Wales, Australia. [41]Department of Urban and Rural Development, Swedish University of Agricultural Sciences, Uppsala, Sweden. [42]School of Biological Sciences, University of Edinburgh, Edinburgh, UK. [43]Southeast Asia Rainforest Research Partnerships, Kota Kinabalu, Malaysia. [44]British Antarctic Survey, Natural Environment Research Council, High Cross, Cambridge, UK. [45]School of Engineering Mathematics and Technology, University of Bristol, Bristol, UK. [46]Institute of Design, Robotics, and Optimisation (iDRO), School of Mechanical Engineering, University of Leeds, Leeds, UK. [47]Rothamsted Research, Harpenden, UK. [48]Department of Integrative Biology, University of Texas at Austin, Austin, TX, USA. [49]Department of Biological Sciences, University of New Hampshire, Durham, NH, USA. [50]Department of Ecological Dynamics, Leibniz Institute for Zoo and Wildlife Research IZW, Berlin, Germany. [51]Research Department of Genetics, Evolution and Environment, University College London, London, UK. [52]Intelligent Robotics Group, Department of Computer Science, Aberystwyth University, Aberystwyth, UK. [53]UK Centre for Ecology and Hydrology (CEH), Wallingford, UK. [54]Department of Biology and Centre d'Études Nordiques, University of Moncton, Moncton, Canada. [55]Center for Ecology, Evolution and Environmental Changes, Departamento de Biologia Animal, Faculdade de Ciências da Universidade de Lisboa, Lisboa, Portugal. [56]Scotland's Rural College (SRUC), Aberdeen, UK. [57]RSPB Centre for Conservation Science, Cambridge, UK. [58]UKCEH, Lancaster Environment Centre, Bailrigg, UK. [59]School of Engineering, Mathematics and Physics, University of Kent, Canterbury, UK. [60]Istituto Italiano di Tecnologia (IIT), Bioinspired Soft Robotics Laboratory, Genoa, Italy. [61]School of Mathematical and Computer Sciences, Heriot-Watt University, Edinburgh, UK. [62]Physical AI (PAI) Laboratory, Department of Mechanical Engineering, Ben-Gurion University of the Negev, Beer-Sheva, Israel. [63]Interdisciplinary Centre for Conservation Biology, Department of Biology, University of Oxford, Oxford, UK. [64]Department of Plant Sciences, University of California Davis, Davis, CA, USA. [65]Department of Biological Sciences, Royal Holloway University of London, Egham, UK. [66]School of Environmental Sciences, University of Liverpool, Liverpool, UK. [67]Departamento de Biodiversidad, Ecología y Evolución, Facultad de Ciencias Biológicas, Universidad Complutense, Madrid, Spain. [68]Department of Environmental Science and Policy, College of Science, George Mason University, Fairfax, VA, USA. [69]Institute of Zoology, Zoological Society of London, Regent's Park, London, UK. [70]Natural Resources and Environmental Studies Institute, University of Northern British Columbia, Prince George, British Columbia, Canada. [71]Centre for Agri-Environmental Research (CAER), School of Agriculture, Policy and Development, Reading University, Reading, UK. [72]School of Natural Sciences, University of Lincoln, Lincoln, UK. [73]Insect Robotics Group, School of Informatics, University of Edinburgh, Edinburgh, UK. [74]School of Biological and Behavioural Sciences, Queen Mary University of London, London, UK. [75]Centre for Sustainable Planning and Environments, University of the West of England Bristol, Bristol, UK. [76]School of Geography, Earth and Environmental Sciences, University of Birmingham, Birmingham, UK. [77]School of Life Sciences, University of Sussex, Brighton, UK. [78]Fauna & Flora (International), The David Attenborough Building, Cambridge, UK. [79]Department of Life Sciences, Imperial College London, London, UK. [80]Sabah Landscape Programme, WWF-Malaysia, Kota Kinabalu, Malaysia. [81]Institute for Sustainability, School of Mechanical Engineering Sciences, University of Surrey, Guildford, UK. [82]Wageningen Environmental Research, Wageningen University & Research (WUR), Wageningen, The Netherlands. [83]School of Biological Sciences, Waipapa Taumata Rau – University of Auckland, Auckland, New Zealand. [84]Conservation Ecology Group, Department of Biosciences, Durham University, Durham, UK. [85]IUCN SSC Species Monitoring Specialist Group, Laboratory for Conservation Biology, Department of Ecology & Evolution, University of Lausanne, Lausanne, Switzerland. [86]Bristol Robotics Laboratory, UWE Bristol, Bristol, UK. [87]Department of Forest Botany, Dendrology and Geobiocoenology, Mendel University in Brno, Brno, Czech Republic. [88]Centre for Robotics and Assembly, School of Aerospace, Transport and Manufacturing (SATM), Cranfield University, Bedford, UK. [89]Edinburgh Centre for Robotics (ECR), School of Mathematical and Computer Sciences, Heriot-Watt University, Edinburgh, UK. [90]School of Science, Western Sydney University, Penrith, New South Wales, Australia. [91]Amphibian and Reptile Conservation, Boscombe, UK. [92]Centre for Conservation & Restoration Science, Edinburgh Napier University, Edinburgh, UK. [93]School of Natural and Environmental Sciences, Newcastle University, Newcastle, UK. [94]Department of Environmental Management, Faculty of Earth and Environmental Sciences, Bayero University, Kano, Nigeria. [95]School of Electronic and Electrical Engineering, University of Leeds, Leeds, UK. [96]Department of Geoinformatics, Cartography and Remote Sensing, Faculty of Geography and Regional Studies, University of Warsaw, Warsaw, Poland. [97]Ecology and Biodiversity, Utrecht University, Utrecht, The Netherlands. ✉e-mail: s.pringle@kent.ac.uk

https://doi.org/10.1038/s41559-025-02704-9




**Extended Data Fig. 1 | Socioeconomic/demographic background of the 98 biodiversity experts who completed Stage One of the modified Delphi technique.** Percentage of experts according to their age, gender, biodiversity survey experience and employment sector. Eighteen experts were employed in more than one sector.

https://doi.org/10.1038/s41559-025-02704-9




**Extended Data Fig. 2 | Countries in which the 98 biodiversity experts who completed stage 1 of the modified Delphi technique were resident.** World map outline ©OpenStreetMap contributors, data available under the Open Database License openstreetmap.org/copyright.

nature portfolio

Corresponding author(s): Dr Stephen Pringle

Last updated by author(s): Apr 3, 2025

# Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our Editorial Policies and the Editorial Policy Checklist.

## Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

| n/a | Confirmed | |
|---|---|---|
| ☐ | ☒ | The exact sample size (*n*) for each experimental group/condition, given as a discrete number and unit of measurement |
| ☐ | ☒ | A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly |
| ☒ | ☐ | The statistical test(s) used AND whether they are one- or two-sided<br>*Only common tests should be described solely by name; describe more complex techniques in the Methods section.* |
| ☒ | ☐ | A description of all covariates tested |
| ☒ | ☐ | A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons |
| ☒ | ☐ | A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals) |
| ☒ | ☐ | For null hypothesis testing, the test statistic (e.g. *F*, *t*, *r*) with confidence intervals, effect sizes, degrees of freedom and *P* value noted<br>*Give P values as exact values whenever suitable.* |
| ☒ | ☐ | For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings |
| ☒ | ☐ | For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes |
| ☒ | ☐ | Estimates of effect sizes (e.g. Cohen's *d*, Pearson's *r*), indicating how they were calculated |

*Our web collection on statistics for biologists contains articles on many of the points above.*

## Software and code

Policy information about availability of computer code

| | |
|---|---|
| Data collection | Questionnaire data collection was conducted using Qualtrics. |
| Data analysis | Data manipulation and coding were carried out in Excel and data visualisations produced in R. |

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

## Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Anonymised data are available without restrictions from the University of Kent data repository at https://doi.org/10.22024/UniKent/01.01.546.

## Human research participants

Policy information about studies involving human research participants and Sex and Gender in Research.

| | |
|---|---|
| Reporting on sex and gender | Data were collected for gender as determined by participants' self-reporting. Testing for differences between genders was not within the scope of this study. Participants provided written informed consent prior to data collection. |
| Population characteristics | See above |
| Recruitment | Participants were recruited by direct contact with experts working in biodiversity and RAS in the research, public, private, NGO and conservation sectors globally. We aimed to ensure that a broad range of experience and expertise was represented in the study, and no other selection criteria were used. Of the 684 experts invited to take part in the Delphi technique: i) 488 were biodiversity experts residing in 43 countries who had expertise in different taxa, habitats and biomes, and ii) 196 were RAS experts residing in 28 countries who were involved in the development and application of a range of robotic system types, sensors and associated technology. |
| Ethics oversight | Ethical approval was granted by the School of Anthropology and Conservation Research Ethics Committee at University of Kent (reference #394 2023). |

Note that full information on the approval of the study protocol must also be provided in the manuscript.

# Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

☐ Life sciences ☒ Behavioural & social sciences ☐ Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

# Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

| | |
|---|---|
| Sample size | *Describe how sample size was determined, detailing any statistical methods used to predetermine sample size OR if no sample-size calculation was performed, describe how sample sizes were chosen and provide a rationale for why these sample sizes are sufficient.* |
| Data exclusions | *Describe any data exclusions. If no data were excluded from the analyses, state so OR if data were excluded, describe the exclusions and the rationale behind them, indicating whether exclusion criteria were pre-established.* |
| Replication | *Describe the measures taken to verify the reproducibility of the experimental findings. If all attempts at replication were successful, confirm this OR if there are any findings that were not replicated or cannot be reproduced, note this and describe why.* |
| Randomization | *Describe how samples/organisms/participants were allocated into experimental groups. If allocation was not random, describe how covariates were controlled OR if this is not relevant to your study, explain why.* |
| Blinding | *Describe whether the investigators were blinded to group allocation during data collection and/or analysis. If blinding was not possible, describe why OR explain why blinding was not relevant to your study.* |

# Behavioural & social sciences study design

All studies must disclose on these points even when the disclosure is negative.

| | |
|---|---|
| Study description | Mixed methods study incorporating responses to closed and open ended questions, and outputs from online workshops. |
| Research sample | In Stage One, we invited 488 biodiversity experts working in the research, public, private, NGO and conservation sectors globally to take part in the Delphi technique. These experts were identified through professional networks, authorship of recent relevant publications and snowball sampling. All 120 experts who agreed to take part were sent a questionnaire to complete.<br>In Stage Two, all 98 biodiversity experts who had completed questionnaires were invited to take part in an online workshop.<br>In Stage Three, we invited 196 experts working on RAS and closely related technology in the research, public and private sectors globally to take part in the Delphi technique. These experts were identified through professional networks, authorship of recent relevant publications and snowball sampling. Forty-nine RAS experts agreed to take part in an online workshop. |
| Sampling strategy | The sampling strategy was a mix of direct contacts and snowball sampling. All of those contacted were sent an invitation to participate in the project, followed by an acknowledgement to those who agreed to take part. Where relevant, this was followed by |

| | |
|---|---|
| | the questionnaire and a link to join the online workshop. Sample sizes were not chosen, but were a result of how many invitees were willing to take part in each stage. |
| Data collection | Questionnaire data were recorded by participants on their own computers. Facilitators present during the online workshops recorded Padlet data inputs. The study was not experimental, so details on experimental conditions are not applicable. |
| Timing | The Delphi technique stages were completed during April - June 2023. |
| Data exclusions | No participants were excluded from the analyses. |
| Non-participation | In Stage One, 22 of the 120 biodiversity experts who agreed to complete a questionnaire did not return the document.<br>In Stage Two, seven of the 98 biodiversity experts who completed the questionnaire and were invited to participate in the online workshop did not attend.<br>In Stage Three, 18 of the 49 RAS experts who accepted our invitation to take part in the online workshop did not attend.<br>We did not collect motivations for non-participation in any of the stages. |
| Randomization | Participants in Stage Two were allocated to one of three simultaneous workshops ('invertebrates'; 'fungi, plants and trees'; 'vertebrates') based on their indicated taxon speciality. Participants with a broad 'generalist' range of taxon expertise were allocated to the 'fungi, plants and trees' workshop. |

# Ecological, evolutionary & environmental sciences study design

All studies must disclose on these points even when the disclosure is negative.

| | |
|---|---|
| Study description | *Briefly describe the study. For quantitative data include treatment factors and interactions, design structure (e.g. factorial, nested, hierarchical), nature and number of experimental units and replicates.* |
| Research sample | *Describe the research sample (e.g. a group of tagged Passer domesticus, all Stenocereus thurberi within Organ Pipe Cactus National Monument), and provide a rationale for the sample choice. When relevant, describe the organism taxa, source, sex, age range and any manipulations. State what population the sample is meant to represent when applicable. For studies involving existing datasets, describe the data and its source.* |
| Sampling strategy | *Note the sampling procedure. Describe the statistical methods that were used to predetermine sample size OR if no sample-size calculation was performed, describe how sample sizes were chosen and provide a rationale for why these sample sizes are sufficient.* |
| Data collection | *Describe the data collection procedure, including who recorded the data and how.* |
| Timing and spatial scale | *Indicate the start and stop dates of data collection, noting the frequency and periodicity of sampling and providing a rationale for these choices. If there is a gap between collection periods, state the dates for each sample cohort. Specify the spatial scale from which the data are taken* |
| Data exclusions | *If no data were excluded from the analyses, state so OR if data were excluded, describe the exclusions and the rationale behind them, indicating whether exclusion criteria were pre-established.* |
| Reproducibility | *Describe the measures taken to verify the reproducibility of experimental findings. For each experiment, note whether any attempts to repeat the experiment failed OR state that all attempts to repeat the experiment were successful.* |
| Randomization | *Describe how samples/organisms/participants were allocated into groups. If allocation was not random, describe how covariates were controlled. If this is not relevant to your study, explain why.* |
| Blinding | *Describe the extent of blinding used during data acquisition and analysis. If blinding was not possible, describe why OR explain why blinding was not relevant to your study.* |

Did the study involve field work? ☐ Yes ☐ No

## Field work, collection and transport

| | |
|---|---|
| Field conditions | *Describe the study conditions for field work, providing relevant parameters (e.g. temperature, rainfall).* |
| Location | *State the location of the sampling or experiment, providing relevant parameters (e.g. latitude and longitude, elevation, water depth).* |
| Access & import/export | *Describe the efforts you have made to access habitats and to collect and import/export your samples in a responsible manner and in compliance with local, national and international laws, noting any permits that were obtained (give the name of the issuing authority, the date of issue, and any identifying information).* |
| Disturbance | *Describe any disturbance caused by the study and how it was minimized.* |

# Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

## Materials & experimental systems

| n/a | Involved in the study |
|---|---|
| ☒ | ☐ Antibodies |
| ☒ | ☐ Eukaryotic cell lines |
| ☒ | ☐ Palaeontology and archaeology |
| ☒ | ☐ Animals and other organisms |
| ☒ | ☐ Clinical data |
| ☒ | ☐ Dual use research of concern |

## Methods

| n/a | Involved in the study |
|---|---|
| ☒ | ☐ ChIP-seq |
| ☒ | ☐ Flow cytometry |
| ☒ | ☐ MRI-based neuroimaging |

## Antibodies

Antibodies used: *Describe all antibodies used in the study; as applicable, provide supplier name, catalog number, clone name, and lot number.*

Validation: *Describe the validation of each primary antibody for the species and application, noting any validation statements on the manufacturer's website, relevant citations, antibody profiles in online databases, or data provided in the manuscript.*

## Eukaryotic cell lines

Policy information about cell lines and Sex and Gender in Research

Cell line source(s): *State the source of each cell line used and the sex of all primary cell lines and cells derived from human participants or vertebrate models.*

Authentication: *Describe the authentication procedures for each cell line used OR declare that none of the cell lines used were authenticated.*

Mycoplasma contamination: *Confirm that all cell lines tested negative for mycoplasma contamination OR describe the results of the testing for mycoplasma contamination OR declare that the cell lines were not tested for mycoplasma contamination.*

Commonly misidentified lines (See ICLAC register): *Name any commonly misidentified cell lines used in the study and provide a rationale for their use.*

## Palaeontology and Archaeology

Specimen provenance: *Provide provenance information for specimens and describe permits that were obtained for the work (including the name of the issuing authority, the date of issue, and any identifying information). Permits should encompass collection and, where applicable, export.*

Specimen deposition: *Indicate where the specimens have been deposited to permit free access by other researchers.*

Dating methods: *If new dates are provided, describe how they were obtained (e.g. collection, storage, sample pretreatment and measurement), where they were obtained (i.e. lab name), the calibration program and the protocol for quality assurance OR state that no new dates are provided.*

☐ Tick this box to confirm that the raw and calibrated dates are available in the paper or in Supplementary Information.

Ethics oversight: *Identify the organization(s) that approved or provided guidance on the study protocol, OR state that no ethical approval or guidance was required and explain why not.*

Note that full information on the approval of the study protocol must also be provided in the manuscript.

## Animals and other research organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research, and Sex and Gender in Research

Laboratory animals: *For laboratory animals, report species, strain and age OR state that the study did not involve laboratory animals.*

| | |
|---|---|
| Wild animals | *Provide details on animals observed in or captured in the field; report species and age where possible. Describe how animals were caught and transported and what happened to captive animals after the study (if killed, explain why and describe method; if released, say where and when) OR state that the study did not involve wild animals.* |
| Reporting on sex | *Indicate if findings apply to only one sex; describe whether sex was considered in study design, methods used for assigning sex. Provide data disaggregated for sex where this information has been collected in the source data as appropriate; provide overall numbers in this Reporting Summary. Please state if this information has not been collected. Report sex-based analyses where performed, justify reasons for lack of sex-based analysis.* |
| Field-collected samples | *For laboratory work with field-collected samples, describe all relevant parameters such as housing, maintenance, temperature, photoperiod and end-of-experiment protocol OR state that the study did not involve samples collected from the field.* |
| Ethics oversight | *Identify the organization(s) that approved or provided guidance on the study protocol, OR state that no ethical approval or guidance was required and explain why not.* |

Note that full information on the approval of the study protocol must also be provided in the manuscript.

## Clinical data

Policy information about clinical studies

All manuscripts should comply with the ICMJE guidelines for publication of clinical research and a completed CONSORT checklist must be included with all submissions.

| | |
|---|---|
| Clinical trial registration | *Provide the trial registration number from ClinicalTrials.gov or an equivalent agency.* |
| Study protocol | *Note where the full trial protocol can be accessed OR if not available, explain why.* |
| Data collection | *Describe the settings and locales of data collection, noting the time periods of recruitment and data collection.* |
| Outcomes | *Describe how you pre-defined primary and secondary outcome measures and how you assessed these measures.* |

## Dual use research of concern

Policy information about dual use research of concern

### Hazards

Could the accidental, deliberate or reckless misuse of agents or technologies generated in the work, or the application of information presented in the manuscript, pose a threat to:

| No | Yes | |
|---|---|---|
| ☒ | ☐ | Public health |
| ☒ | ☐ | National security |
| ☒ | ☐ | Crops and/or livestock |
| ☒ | ☐ | Ecosystems |
| ☒ | ☐ | Any other significant area |

### Experiments of concern

Does the work involve any of these experiments of concern:

| No | Yes | |
|---|---|---|
| ☒ | ☐ | Demonstrate how to render a vaccine ineffective |
| ☒ | ☐ | Confer resistance to therapeutically useful antibiotics or antiviral agents |
| ☒ | ☐ | Enhance the virulence of a pathogen or render a nonpathogen virulent |
| ☒ | ☐ | Increase transmissibility of a pathogen |
| ☒ | ☐ | Alter the host range of a pathogen |
| ☒ | ☐ | Enable evasion of diagnostic/detection modalities |
| ☒ | ☐ | Enable the weaponization of a biological agent or toxin |
| ☒ | ☐ | Any other potentially harmful combination of experiments and agents |

## ChIP-seq

### Data deposition

☐ Confirm that both raw and final processed data have been deposited in a public database such as GEO.

☐ Confirm that you have deposited or provided access to graph files (e.g. BED files) for the called peaks.

| | |
|---|---|
| Data access links<br>*May remain private before publication.* | *For "Initial submission" or "Revised version" documents, provide reviewer access links. For your "Final submission" document, provide a link to the deposited data.* |
| Files in database submission | *Provide a list of all files available in the database submission.* |
| Genome browser session<br>(e.g. UCSC) | *Provide a link to an anonymized genome browser session for "Initial submission" and "Revised version" documents only, to enable peer review. Write "no longer applicable" for "Final submission" documents.* |

### Methodology

| | |
|---|---|
| Replicates | *Describe the experimental replicates, specifying number, type and replicate agreement.* |
| Sequencing depth | *Describe the sequencing depth for each experiment, providing the total number of reads, uniquely mapped reads, length of reads and whether they were paired- or single-end.* |
| Antibodies | *Describe the antibodies used for the ChIP-seq experiments; as applicable, provide supplier name, catalog number, clone name, and lot number.* |
| Peak calling parameters | *Specify the command line program and parameters used for read mapping and peak calling, including the ChIP, control and index files used.* |
| Data quality | *Describe the methods used to ensure data quality in full detail, including how many peaks are at FDR 5% and above 5-fold enrichment.* |
| Software | *Describe the software used to collect and analyze the ChIP-seq data. For custom code that has been deposited into a community repository, provide accession details.* |

## Flow Cytometry

### Plots

Confirm that:

☐ The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).

☐ The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).

☐ All plots are contour plots with outliers or pseudocolor plots.

☐ A numerical value for number of cells or percentage (with statistics) is provided.

### Methodology

| | |
|---|---|
| Sample preparation | *Describe the sample preparation, detailing the biological source of the cells and any tissue processing steps used.* |
| Instrument | *Identify the instrument used for data collection, specifying make and model number.* |
| Software | *Describe the software used to collect and analyze the flow cytometry data. For custom code that has been deposited into a community repository, provide accession details.* |
| Cell population abundance | *Describe the abundance of the relevant cell populations within post-sort fractions, providing details on the purity of the samples and how it was determined.* |
| Gating strategy | *Describe the gating strategy used for all relevant experiments, specifying the preliminary FSC/SSC gates of the starting cell population, indicating where boundaries between "positive" and "negative" staining cell populations are defined.* |

☐ Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.

## Magnetic resonance imaging

### Experimental design

| | |
|---|---|
| Design type | *Indicate task or resting state; event-related or block design.* |

Design specifications

*Specify the number of blocks, trials or experimental units per session and/or subject, and specify the length of each trial or block (if trials are blocked) and interval between trials.*

Behavioral performance measures

*State number and/or type of variables recorded (e.g. correct button press, response time) and what statistics were used to establish that the subjects were performing the task as expected (e.g. mean, range, and/or standard deviation across subjects).*

## Acquisition

Imaging type(s)

*Specify: functional, structural, diffusion, perfusion.*

Field strength

*Specify in Tesla*

Sequence & imaging parameters

*Specify the pulse sequence type (gradient echo, spin echo, etc.), imaging type (EPI, spiral, etc.), field of view, matrix size, slice thickness, orientation and TE/TR/flip angle.*

Area of acquisition

*State whether a whole brain scan was used OR define the area of acquisition, describing how the region was determined.*

Diffusion MRI ☐ Used ☐ Not used

## Preprocessing

Preprocessing software

*Provide detail on software version and revision number and on specific parameters (model/functions, brain extraction, segmentation, smoothing kernel size, etc.).*

Normalization

*If data were normalized/standardized, describe the approach(es): specify linear or non-linear and define image types used for transformation OR indicate that data were not normalized and explain rationale for lack of normalization.*

Normalization template

*Describe the template used for normalization/transformation, specifying subject space or group standardized space (e.g. original Talairach, MNI305, ICBM152) OR indicate that the data were not normalized.*

Noise and artifact removal

*Describe your procedure(s) for artifact and structured noise removal, specifying motion parameters, tissue signals and physiological signals (heart rate, respiration).*

Volume censoring

*Define your software and/or method and criteria for volume censoring, and state the extent of such censoring.*

## Statistical modeling & inference

Model type and settings

*Specify type (mass univariate, multivariate, RSA, predictive, etc.) and describe essential details of the model at the first and second levels (e.g. fixed, random or mixed effects; drift or auto-correlation).*

Effect(s) tested

*Define precise effect in terms of the task or stimulus conditions instead of psychological concepts and indicate whether ANOVA or factorial designs were used.*

Specify type of analysis: ☐ Whole brain ☐ ROI-based ☐ Both

Statistic type for inference
(See Eklund et al. 2016)

*Specify voxel-wise or cluster-wise and report all relevant parameters for cluster-wise methods.*

Correction

*Describe the type of correction and how it is obtained for multiple comparisons (e.g. FWE, FDR, permutation or Monte Carlo).*

## Models & analysis

| n/a | Involved in the study |
|---|---|
| ☐ | ☐ Functional and/or effective connectivity |
| ☐ | ☐ Graph analysis |
| ☐ | ☐ Multivariate modeling or predictive analysis |

Functional and/or effective connectivity

*Report the measures of dependence used and the model details (e.g. Pearson correlation, partial correlation, mutual information).*

Graph analysis

*Report the dependent variable and connectivity measure, specifying weighted graph or binarized graph, subject- or group-level, and the global and/or node summaries used (e.g. clustering coefficient, efficiency, etc.).*

Multivariate modeling and predictive analysis

*Specify independent variables, features extraction and dimension reduction, model, training and evaluation metrics.*



https://dspace.org/

# Opportunities and challenges for monitoring terrestrial biodiversity in the robotics age

**Pringle, Stephen; Dallimer, Martin; Goddard, Mark A.; Le Goff, Léni K.; Hart, Emma ...**

**2025-06**
Springer Nature
https://doi.org/10.1038/s41559-025-02704-9
Pringle S, Dallimer M, Goddard MA, et al., (2025) Opportunities and challenges for monitoring terrestrial biodiversity in the robotics age. Nature Ecology & Evolution, Volume 9, Issue 6, June 2025, pp. 1031-1042

Downloaded from a DSpace repository. https://dspace.org/