

Applied and Environmental Microbiology, Volume 83, Issue 10, May 2017, Article number e03462-16
DOI:10.1128/AEM.03462-16

1 **COREXIT 9500 enhances oil biodegradation and changes microbial community**
2 **structure of oil-enriched microcosms**

3
4 Stephen M. Techtman¹, Mobing Zhuang², Pablo Campo³, Edith Holder⁴, Michael Elk⁴, Terry C.
5 Hazen^{5,6}, Robyn Conmy⁷, Jorge W. Santo Domingo^{7*}

6
7 ¹Department of Biological Sciences, Michigan Technological University, Houghton MI, USA

8 ²Department of Biomedical, Chemical and Environmental Engineering, University of
9 Cincinnati, Cincinnati, OH45221, USA

10 ³Cranfield Water Science Institute, Cranfield University, Cranfield MK43 0AL, UK

11 ⁴Pegasus, Inc., Cincinnati, OH, 45268, USA

12 ⁵Department of Civil & Environmental Engineering, University of Tennessee, Knoxville,
13 TN, USA

14 ⁶Center for Environmental Biotechnology, University of Tennessee, Knoxville, TN USA

15 ⁷U. S. Environmental Protection Agency, Cincinnati, OH 45268, USA

16
17 Running Title: Effect of COREXIT 9500 on biodegradation

18
19 Keywords: Oil Biodegradation, microbial community, Gulf of Mexico, COREXIT 9500,
20 dispersant

21
22 * Corresponding author: Jorge W. Santo Domingo; santodomingo.jorge@epa.gov

23
24
25
26
27
28
29 **ABSTRACT**

30 To better understand the impact of COREXIT 9500 on the structure and activity levels of
31 hydrocarbon degrading microbial communities, we analyzed next-generation 16S rRNA
32 gene sequencing libraries of hydrocarbon enrichments grown at 5 and 25 °C by using
33 both DNA and RNA extracts as sequencing templates. Oil biodegradation patterns in
34 both 5 and 25°C enrichments were consistent with those reported in the literature (i.e.,
35 aliphatics were degraded faster than aromatics). A slight increase in biodegradation was
36 observed in the presence of COREXIT at both temperatures. Differences in community
37 structure were observed between treatment conditions in the DNA-based libraries. The
38 25 °C consortia was dominated by *Vibrio*, *Pseudoidiomarina*, *Marinobacter*,

39 *Alcanivorax*, and *Thalassospira* species, while the 5 °C consortia were dominated by
40 several genera of *Flavobacteria*, *Alcanivorax* and *Oleispira*. Most these genera have been
41 linked to hydrocarbon degradation and have been observed after oil spills. *Colwellia* and
42 *Cycloclasticus*, known aromatic degraders, were also found in these enrichments. The
43 addition of COREXIT did not have an effect on the active bacteria community structure
44 of the 5 °C consortia, while at 25 °C a decrease in the relative abundance of
45 *Marinobacter* was observed. At 25 °C, *Thalassospira*, *Marinobacter*, and
46 *Pseudodidiomarina* were present at higher relative abundance in the RNA than DNA
47 libraries suggesting that they were active in degradation. Similarly, *Oleispira* was greatly
48 stimulated by the addition of oil at 5 °C.

50 **IMPORTANCE**

51
52 While COREXIT 9500 is widely applied after oil spills for its reported dispersing
53 activity, there is still debate on the effectiveness on enhancing oil biodegradation and its
54 potential toxic effect on oil-degrading microbial communities. The results of this study
55 provide some insights on the microbial dynamics of hydrocarbon degrading bacterial
56 populations in the presence of COREXIT 9500. Operational Taxonomic Unit (OTU)
57 analyses indicated that several OTUs were inhibited by the addition of COREXIT.
58 Conversely, a number of OTUs were stimulated by the addition of the dispersant, many
59 of which were identified as known hydrocarbon degrading bacteria. The results highlight
60 the value of using RNA-based methods to further understand the impact of dispersant on
61 the overall activity of different hydrocarbon degrading bacterial groups.

62

63 **Introduction**

64 Diverse marine microbes have the ability to degrade crude oil under both aerobic and
65 anaerobic conditions (1, 2). Crude oil is a complex mixture of hydrocarbons, with
66 constituents including both branched and straight chain alkanes as well as polycyclic
67 aromatic hydrocarbons (PAHs). In addition to natural inputs of oil into the marine
68 environment(3), oil has been released via accidental oil spills. These oil spills have the
69 potential to drastically impact ecosystems and result in long-term environmental changes
70 (4, 5). Various strategies have been employed to aid in the cleanup of oil spills, including
71 dispersant application (6). Chemical dispersants contain a mixture of chemicals designed
72 to break up an oil slick by reducing the interfacial surface tension of the oil, causing it to
73 form micron-sized droplets. These droplets can then be diluted in the water column
74 preventing the formation of a surface slick. Indigenous oil-degrading microbes are
75 believed to degrade these dispersed oil droplets, with smaller droplets being more rapidly
76 degraded (7).

77 During the *Deepwater Horizon* oil spill 2.1 million gallons of the chemical
78 dispersant COREXIT 9500 were applied at both the surface and at the seafloor (8).
79 There is debate about the effect of COREXIT 9500 on both the rates of oil
80 biodegradation as well as its effects on microbial community composition. Several
81 reports from diverse marine locations have demonstrated that the addition of COREXIT
82 9500 enhances oil biodegradation (9,10,7,11,12). Other studies have shown a neutral or
83 inhibitory effect of COREXIT 9500 addition on microbial activity (13-15). A number of
84 factors could explain these discrepancies including differences in experimental set up,

length of experiment, water source (including handling and storage) and concentrations of oil and dispersants. These contrary findings underscore the need for more in-depth analysis of the effect of COREXIT 9500 on oil biodegradation.

There are also differing reports of the impact of COREXIT on the microbial community composition. One recent study used high-throughput 16S rRNA gene sequencing to investigate the effect of oil and dispersant on microbial community structure (9). They demonstrated that many of the same taxa dominated in the oil-only and oil plus dispersant treatments. The minimal effect of COREXIT 9500 on community structure in Baelum *et al* (2012) is underscored by the very high COREXIT 9500 concentration used in their study. While a 1:25 dispersant to oil ratio was the target ratio for subsea dispersant application during the *Deepwater Horizon* oil spill (10, 16), Baelum *et al* (2012) used a dispersant to oil ratio of 1:1.6. Another recent study investigated the impact of COREXIT 9500 on both biodegradation of the water accessible fraction (WAF) of oil as well the microbial community structure (15). The authors conclude that COREXIT 9500 addition resulted in a distinct community of oil degraders.

The majority of previous studies examining the effect of COREXIT 9500 addition on oil biodegradation have focused on South Louisiana Crude oils. Alaska North Slope (ANS) crude oil is another important oil type that has been focus of oil biodegradation studies. COREXIT 9500 has been shown to effectively disperse ANS in wave tank experiments (18) as well as enhance biodegradation of ANS crude oil by Chukchi Sea microbes in microcosms incubated at -1 °C (10). However, the impact of COREXIT 9500 on the composition of microbial communities degrading ANS oil has not been reported. We investigated the effect of COREXIT 9500 on biodegradation of ANS crude

oil and the impact of COREXIT addition on microbial community structure of two hydrocarbon degrading consortia. Next-generation sequencing of the 16S rRNA gene using both DNA and RNA extracts as sequencing targets was used to study the microbial community composition. By comparing the results on both the DNA and RNA level we determined the effect of COREXIT 9500 on the total bacteria community structure (DNA) as well as the active bacterial fraction (RNA). The goal of this present study was better understand the effects of COREXIT 9500 on oil biodegradation, and to seek to reconcile the contrary findings regarding the impacts of COREXIT 9500 addition on oil biodegrading and community structure changes.

117

118 **Experimental Methods**

119 Microcosm set-up

Two different enrichment cultures generate from samples collected nearby the site of *Deepwater Horizon* oil spill (19). One culture was derived from a surface sample and the other was derived from a deep-water sample. These enrichments were grown at temperatures that reflect those from where the samples were originally collected. The methods by which these enrichments were derived can be found elsewhere (17). Briefly, the enrichments were originally grown at 5 °C (referred to as the cryo enrichment) or 25 °C (referred to as the meso enrichment) on GP2 medium (pH 7.5) (20) amended with South Louisiana crude oil, harvested via centrifugation, followed by two saline washes to remove oil. The washed cells were resuspended in saline as 10 fold concentration with glycerol (10% final concentration), and stored at -20 °C until used in the microcosms studies. The cryo enrichment was derived from a sample collected from

131 deep water from the Gulf of Mexico at a depth of 1,240 m and is designed to be
132 representative of a deep water oil degrading consortium. The meso enrichment was
133 derived from a surface water sample from the Gulf of Mexico at a depth of 5 m and is
134 designed to be representative of a surface water oil degrading consortia.

135 In the present study, several frozen stocks of the appropriate enrichment were
136 thawed, combined, and washed once with twice the volume of saline (to remove
137 glycerol). Aliquots of the final resuspended stock (0.5 mL) were then used to inoculate
138 each of the flasks containing GP2 medium amended with ANS oil, dispersant (COREXIT
139 9500), or ANS oil and dispersant at a volumetric dispersant-to-oil ratio of 1:25 (final
140 COREXIT concentration of 1.12 µg/L). Treatments were incubated aerobically by
141 shaking at 200 rpm and incubated at the appropriate temperature (25 °C for the meso and
142 5 °C for the cryo enrichments). Sodium azide killed controls were also prepared to
143 account for abiotic hydrocarbon losses, similarly to the set up described in Campo-
144 Moreno *et al.* (2013) (17). Experiments were conducted in triplicate per treatment with a
145 total of 100 mL of medium in 250 mL flasks. Flasks were sacrificed at 0, 2, 4, 8, 12, 16,
146 24, 32, 40, 48 days. The cryo enrichment had an additional time point at 56 days. At
147 each time point, flasks were sacrificed to measure hydrocarbon residual concentrations
148 and to conduct molecular analyses.

149 Hydrocarbon measurements

150 Subsamples were collected from each flask to determine the concentration of dioctyl
151 sodium sulfosuccinate (DOSS) by liquid chromatography tandem mass spectrometry
152 (LC-MS/MS) and of hydrocarbons by gas chromatography tandem mass spectrometry
153 (GC-MS/MS). The procedures used for DOSS and oil extractions and internal standards

154 used to normalize for DOSS, total alkanes and total aromatics are described elsewhere
155 (17, 19). Hydrocarbons are reported as a ratio to the conservative biomarker hopane,
156 which has been routinely used in determination of biodegradation of oil (21)

157

158 DNA/RNA Extraction and sequencing

159 For each time point, samples (1 mL) were transferred from the microcosms to
160 sterile microcentrifuge tubes. Samples were then centrifuged at $10,000 \times g$ for 10 min.
161 Supernatants were decanted without disturbing cell pellets and tubes were stored at -80
162 °C until nucleic acid extractions. Both RNA and DNA were extracted from the frozen
163 cell pellets with AllPrep DNA/RNA Mini Kit (Qiagen GmbH) (22). RNA extracts were
164 further purified with TURBO DNA-freeTM (Ambion) kits according to the routine
165 DNase treatment step. Purified RNA was eluted to a final volume of 30 µL and stored at
166 -80°C. cDNA was synthesized from the purified total RNA extracts using the Superscript
167 III (Life Technologies, San Francisco, CA). RNA and DNA concentrations were
168 measured by Qubit RNA High Sensitivity (Life Technologies) and dsDNA High
169 Sensitivity (Life Technologies) assay kits and the Qubit 2.0 Fluorometer (Life
170 Technologies). Aliquots (2 µL) of nucleic acid extracts were used in all quantification
171 reaction mixtures. Concentration of DNA extracts ranged from 0.05 to 1 ng/µL while
172 RNA extracts ranged from 2 to 8 ng/uL, with most within 2 to 5 ng/µL. cDNA synthesis
173 was performed using SuperScript III First-Strand Synthesis kits (Invitrogen) using
174 random hexamer primers according to the manufacturer's instructions. cDNA synthesis
175 was performed the same day of the RNA extraction to maximize RNA and minimize
176 RNA degradation. Aliquots of the RNA extracts were used as PCR templates to verify

177 the absence of DNA contaminants. An 8 μ L volume of RNA was used for the synthesis
178 generating a final product volume of 21 μ L. cDNA was stored at -20 °C and excess RNA
179 was stored at -80 °C.

180 The bacterial composition of each sample was determined via sequencing analysis
181 of the 16S rRNA gene V4 region using the primers 515f/806r as described elsewhere
182 (23). PCR assays used to generate the sequencing libraries were performed in 25 μ L
183 volumes using the Ex Taq kit (Takara) with 200 nM each of the forward and reverse
184 primer and 2 μ L of template nucleic acid. Cycling conditions involved an initial 5 min
185 denaturing step at 95 °C, followed by 35 cycles at 95 °C for 45 s, 50 °C for 60 s, 72 °C
186 for 90 s, and a final elongation step at 72 °C for 10 min. Each barcode corresponded to
187 an eight-base sequence unique to each sample. Amplicons were visualized on an agarose
188 gel to confirm product sizes. Aliquots of each amplicon were pooled based on band
189 intensity and sequenced on an Illumina Miseq benchtop sequencer using 250 bp paired-
190 end kits at the Cincinnati Children's Hospital DNA Core facility.

191

192 16S rRNA library construction and analysis

193 The resulting DNA sequences were analyzed by using the QIIME version 1.8.0-
194 dev pipeline (24). Paired-end raw reads were assembled with fastq-join (25). The
195 assembled sequences were demultiplexed and quality filtered in QIIME to remove reads
196 with phred scores below 20 (-q 19). Chimera detection was then performed on assembled
197 reads with UCHIME (26, 27). Sequences were then clustered into operation taxonomic
198 units (OTUs, 97% similarity) with UCLUST (26) by using the open reference clustering
199 protocol. The resulting representative sequences were aligned using PyNAST (28) and

200 given a taxonomic assignment using RDP (29) retrained with the May 2013 Greengenes
201 release. The resulting OTU table was filtered to keep OTUs that were present at greater
202 than 0.005%, and then rarified to the minimum number of remaining sequences in the
203 samples (1670 sequences). Bray-Curtis dissimilarity (30) weighted and unweighted
204 unifrac distances (31) were calculated from the rarefied OTU table with the
205 beta_diversity.py script in QIIME. Several OTUs were originally misclassified as
206 *Methylococcales* spp. Classification of these OTUs via RDP Classifier indicated that the
207 appropriate classification for these OTUs was *Cycloclasticus*. Therefore, we included
208 these OTUs in the *Cycloclasticus* group.

209

210 Statistical Analysis of Sequencing Data

211 In order to test the hypothesis that microbial communities in different treatments
212 were significantly similar to each other and statistically different from other treatments,
213 non-metric multidimensional scaling (NMDS) was used. Weighted Unifrac distances
214 were used to construct two-dimensional NMDS plots. The lowest stress configuration
215 was chosen from 50 iterations of plot construction. Stress values were calculated by the
216 default stress calculation in the NMDS command in the ecodist package. To test if
217 samples from the same treatment were significantly different from each other
218 PERMANOVA (32) analysis was performed on both the Bray Curtis and weighted
219 Unifrac matrices with the Adonis function in the vegan package (33) in R (34). Samples
220 were grouped according to the treatment. PERMANOVA analysis was performed by
221 using 999 permutations. PERMANOVA analysis of all three treatments will indicate if
222 there is a significant difference between all three groups. To distinguish which

223 treatments were different from each other, PERMANOVA analysis was done on subsets
224 of the weighted Unifrac distance matrix that only include samples from two of the three
225 treatments. These pair-wise PERMANOVAs were used to distinguish which treatments
226 were different from each other.

227 The MetagenomeSeq package (35), as implemented in QIIME, was used to
228 identify which OTUs were differentially abundant between treatments for each
229 temperature regime. The MetagenomeSeq package provides a more robust normalization
230 method than total-sum scaling or relative abundance. It uses cumulative sum-scaling, in
231 which counts are divided by the cumulative sum of counts up to a percentile determined
232 using a data-driven approach. Furthermore, MetagenomeSeq employs the zero-inflated
233 Gaussian to account for under sampling of the microbial community in low coverage
234 samples. This enables a more robust analysis of differential abundance between samples
235 or conditions. Each treatment was compared to the other treatments incubated at the
236 same temperature, and OTUs that were significantly different (corrected p value < 0.05)
237 in one treatment compared to the other two were considered to be differentially abundant.

238 **Results and Discussion**

239 Hydrocarbons are rapidly degraded at both 25 and 5 °C

240 Hydrocarbons were measured at various points throughout the course of the experiment.
241 These hydrocarbons were grouped according to total alkanes and PAHs. Many of the
242 hydrocarbon classes were readily degraded with and without COREXIT 9500 (Figure 1),
243 which indicates that the microbial community present in our microcosms contained a
244 robust community of oil-degraders. As Figure 1 depicts, the trend in oil biodegradation
245 in these experiments followed those observed in other studies (10, 36). Total alkane

246 degradation occurred rapidly at 25 °C, with the majority of alkanes being degraded by
247 day 4 in the presence of COREXIT and by day 8 in the absence of COREXIT (Figure
248 1B). In fact, the respective first-order degradation rates were 0.19 ± 0.03 and $0.15 \pm$
249 0.02 d^{-1} in the presence and absence of COREXIT. At 5 °C, minimal alkane
250 degradation was observed the first few days. Substantial alkane degradation was
251 observed after day four and by day 12 the majority of alkanes were degraded both with
252 and without COREXIT. PAHs showed longer lag phases than alkanes at both
253 temperatures (Figure 1C). At 25°C, substantial overall PAH degradation occurred after
254 four days as the extent of removal was 28% in the presence of COREXIT, while in its
255 absence the initial PAHs load remained at near 100%. By day 8, the extent of removal
256 was approximately 77% in both conditions. At 5 °C, PAH degradation was observed to
257 begin after day 12 in the presence of COREXIT 9500 and after day 16 in the absence of
258 COREXIT 9500. Similar trends are observed for particular classes within the broad
259 categories of alkanes and PAHs (Supplemental Figure 1).

260 The effect of COREXIT 9500 was clearly seen when comparing the *n*-alkanes with
261 chain lengths between 30 and 35 (Figure 1D). At 25°C, this group of alkanes was
262 degraded to below detection ($< 2 \text{ mg/mg}$ hopane) after six days in the presence of
263 COREXIT 9500, while in its absence this group of alkanes was measurable in samples
264 collected at day 16. A similar trend is observed at the lower temperature. In the presence
265 of COREXIT 9500, there is a dramatic decrease in the concentration of this group of
266 alkanes after eight days. However, in the absence of COREXIT 9500 the concentration
267 gradually decreased and reaches the lowest level after 24 days. These results suggest that
268 COREXIT 9500 enhances the rates of oil biodegradation, which is in line with a number

269 of studies that report increased rates of oil biodegradation in the presence of dispersants
270 (10, 17, 37). This finding is in contrast to the recent paper suggesting that COREXIT
271 9500 inhibits oil biodegradation (20).

272 Degradation of DOSS was measured in order to understand the fate of dispersant
273 in these conditions (Figure 1A). At 25 °C, DOSS was quickly degraded in the COREXIT
274 9500 alone condition, but remained above the detection limit after 48 days. However, in
275 the presence of oil, DOSS was completely degraded after 40 days. This seems to indicate
276 that DOSS degradation occurred more rapidly in the presence of oil at 25 °C. DOSS
277 degradation was much slower at 5 °C with no DOSS degraded over the 56 day
278 experiment. These results are comparable with previous studies that have shown a
279 substantial difference in dispersant degradation at 25 °C and 5°C (17, 38) and persistence
280 of DOSS in the deep ocean (39).

281 Different treatments select for distinct microbial community composition

282 Sequencing of the 16S rRNA gene and 16S rRNA gene cDNA transcripts was
283 performed to better understand the effect of COREXIT 9500 on microbial community
284 composition and relative activity of identified members, respectively. In these
285 experiments, there were stark differences between the total community (DNA) and the
286 active community (RNA) composition (Figure 2 and 3). The total microbial community
287 is a measure of all cells present in these samples. Many of these cells may be present at
288 high abundances, but not active during the time of sampling. Therefore, measurement of
289 the microbial community on the RNA level provides insights into which taxa are active
290 during the time of sampling. This approach provides key insights into which of the oil
291 degrading taxa are actively metabolizing the oil components as well as clarifies the

292 impact of COREXIT addition on the active microbes. Analysis of the total community
293 can often be misleading, as it does not identify the taxa that are actively metabolizing at
294 the time of sampling. While many of the same taxa are present in both the total and the
295 active communities, the dominant taxa are different between the two communities
296 indicating that the key organisms involved in metabolizing hydrocarbons may be
297 incorrectly identified if using only DNA-based total community data.

298 Multivariate statistics indicate that samples from the same treatment tend to have
299 similar microbial community composition and group together on non-metric
300 multidimensional scaling plots (Figure 3). PERMANOVA analysis indicated that there
301 were statistically significant differences between treatments at both the active and the
302 total community levels (Table 1). These differences indicate that the treatments select for
303 a distinct set of taxa. These differences appear to be the strongest when comparing
304 treatments with and without oil, suggesting that the addition of oil most strongly impacts
305 the community structure. While there are statistically significant differences between the
306 oil-only and oil plus COREXIT treatment, these differences are only weakly significant
307 in some cases (Table 1).

308 Similarly to what was observed in the Gulf of Mexico during the *Deepwater*
309 *Horizon* oil spill (DWH), the oil-degrading microbial community at 25°C was
310 dramatically different from the community at 5°C (40) (Figure 2). The 25°C consortium
311 was dominated by unclassified members of the *Vibrio*, *Pseudoidiomarina*, *Marinobacter*,
312 *Alcanivorax*, and *Thalassospira* species. This was similar to the community found in the
313 surface water during the DWH oil spill, which was dominated by members of the
314 *Alteromonadales* and the *Vibrionales* (41). The 5°C consortia were dominated by several

genera of *Flavobacteria*, as well as *Oceanospirillales* related to *Alcanivorax* and *Oleispira*. Additionally, *Colwellia* spp. were found to be present in the majority of samples during the incubation. Microbes classified as *Cycloclasticus* were present in the later time points of many conditions at both 25°C and 5°C. These same taxa were present at high abundance in the deep-water plume during the DWH oil spill (40). Therefore, the major oil-degrading taxa in these enriched consortia are related the dominant members of the microbial community that responded to the spilled oil in the Gulf of Mexico (42).

Treatment effects on the active microbial community at 25 °C

Analysis of the composition of the active bacterial community provides a clearer understanding of the microbial taxa actively responding to the treatment. A number of taxa were present at different relative abundances in the active community compared to the total community. For example, the total community at 25 °C was dominated *Vibrionaceae* (up to 80% of the recovered reads), whereas the *Vibrionaceae* only represented ~25% of recovered reads in the active community. Members of the *Thalassospira*, *Marinobacter*, and *Pseudodidiomarina* were present at much higher relative abundance in the active community compared to the total community. Additionally, *Marinobacter* represented between 2 – 7% of the total community across all treatments. In the active community, *Marinobacter* represented 1.7% of recovered reads in the COREXIT 9500-only treatment. However, in oil-amended conditions *Marinobacter* comprised 27 and 17% of reads in the oil-only and the oil plus COREXIT 9500 treatments, respectively. This finding suggests that *Marinobacter* is stimulated by the addition of oil. *Marinobacter* are a well-known oil-degrading bacteria (43) that were present in the Gulf of Mexico during the DWH oil spill (44).

338 The role of *Vibrionales* in these consortia is somewhat enigmatic. *Vibrio* spp.
339 have been identified as responding to the DWH oil spill in beach sediments (45, 46);
340 however, evidence for degradation of hydrocarbons by *Vibrio* is sparse. There is some
341 previous evidence of their ability to use hydrocarbons (47). One study performed during
342 the DWH oil spill demonstrated an enrichment of *V. vulnificus* in tar balls collected from
343 Gulf of Mexico beaches (48). Conversely, a number of *V. parahaemolyticus* isolates
344 from water and sediments from the Gulf of Mexico were shown to not be able to degrade
345 PAHs (49). Further, little to no oil consumption was observed for *Vibrio* strains isolated
346 from Gulf of Mexico beach sands (46). *Vibrio* are common inhabitants of the ocean and
347 are known to grow on a wide range of organic compounds (47). It is therefore possible
348 that these *Vibrio* species are responding to the spilled oil as well as metabolizing the
349 compounds produced by other oil-degrading taxa.

350 Treatment effects on the active microbial community at 5 °C

351 Differences between the active and total community were also observed at 5 °C.
352 The *Flavobacteriaceae* related to *Winogradskyella* were more dominant members of the
353 active community in the COREXIT 9500 only condition compared to the oil-amended
354 conditions. *Oleispira* were also present at high abundance in the total community in all
355 conditions (on average between 14 – 17% of recovered reads). *Oleispira* represented a
356 much higher proportion of the active community in the oil-amended conditions (on
357 average 15% of the oil-only and 9.4% of oil plus COREXIT 9500) compared to the total
358 community, but were only a minor constituent of the active community in COREXIT
359 9500-only conditions (on average 2.9%). The increase in the proportion of *Oleispira* in
360 the oil-amended conditions suggests that *Oleispira* is stimulated by the addition of oil in

361 these microcosms. *Oleispira* spp. are known as psychrophilic alkane-degraders that were
362 originally described as an isolate from the Antarctic (50). The closest cultured
363 representative to the *Oceanospirillales*, which dominated the deep-water plume at the
364 early stages of the *Deepwater Horizon* oil spill, is *O. antarctica* (51).

365 *Cycloclasticus* spp. have been shown to be numerically dominant members of the
366 *in situ* microbial community that responded to the *Deepwater Horizon* oil spill (40, 44).
367 Many of the described species of *Cycloclasticus* are able to degrade a variety of PAHs.
368 In our microcosms, sequences classified as *Cycloclasticus* were recovered from the active
369 communities at both temperatures. At 5 °C, *Cycloclasticus* spp. were present at low
370 abundance in the early time points (on average 0.8% at Day 8) and remained at low
371 abundance until day 24 in the only oil condition and Day 16 in the oil plus COREXIT
372 9500 condition, when they increased to 5% of the active community. This increase in the
373 relative abundance of *Cycloclasticus* corresponds with the beginning of PAH degradation
374 at 5°C (Figure 1C), further supporting the role of *Cycloclasticus* spp. in degrading PAHs
375 in these microcosms.

376 *Colwellia* spp. were also important members of the deep water plume at later time
377 points in the spill (40, 52). *Colwellia* are ubiquitous psychrophilic heterotrophs. Data
378 from experiments performed during the spill indicated that they degraded gaseous
379 hydrocarbons, namely propane, ethane, and benzene (40). *Colwellia* were on average
380 4.9% of the total community in all conditions, whereas in the active community, they
381 were 13.2% in the COREXIT 9500-only condition, and only 2.1 and 3.8% of the active
382 community in oil-only and oil plus COREXIT 9500 treatments, respectively. In the
383 COREXIT 9500-only condition, reads classified as *Colwellia* increased over the course

384 of the experiment; increasing from 0.9% to a maximum of 22.4% on day 24. This
385 increase in *Colwellia* in the COREXIT only conditions may be due to metabolism of
386 COREXIT components by *Colwellia* as suggested previously (15, 41).

387 Analysis of these microcosms further confirms that the oil-degrading microbial
388 communities derived from surface water (meso enrichment) are distinct from the
389 microbial communities derived from deep water (cryo enrichment). The surface
390 microbial community was able to rapidly degrade the majority of the components of oil
391 and COREXIT 9500. The surface microbial community was dominated by well
392 characterized oil-degrading microbes such as *Marinobacter* species, which are common
393 mesophilic oil-degrader. The deep-water community was composed of distinct taxa
394 compared to the surface community and degraded oil at a different rate than under
395 surface water conditions. These differences are in part due to differences in temperature.

396 While it is expected that low temperature would act to slow the rates of oil
397 degradation, we find that the deep-water community is dominated by many of cold-
398 adapted oil-degrading taxa. This suggests that deep water oil biodegradation would be
399 more rapid than would be expected based on the Q10 approach (53). Many of the same
400 cold-adapted oil-degrading taxa found in the 5°C microcosms were found to be important
401 in responding to oil spills in other cold locations (50, 54, 55). The presence of many
402 these same taxa in other oil-impacted locations suggests that there is a ubiquitous
403 complement of cold-adapted oil degraders that may respond to oil spills in cold climates.
404 Interestingly, one of the dominant oil-degraders in the 5 °C incubation was an
405 *Alcanivorax* sp. *Alcanivorax* are ubiquitous oil-degraders commonly found in surface
406 seawater. They were not reported as being dominant in deep water plume during the

407 DWH oil spill. Previous work has demonstrated that *Alcanivorax* sp. were dominant in
408 oil-impacted seawater when sufficient nutrients are present. It is possible that in the deep
409 ocean, nutrient limitation impedes the growth of *Alcanivorax* species. However, in these
410 microcosms, nutrients are supplied in the artificial seawater allowing for the proliferation
411 of these *Alcanivorax* strains. While *Alcanivorax* species have been isolated from the
412 deep sea, they are not typically thought of as cold-adapted microbes. Therefore, the fact
413 that *Alcanivorax* species dominate the active community at 5 °C suggests the potential for
414 cold-adapted members of the *Alcanivorax* to be important responders in cold
415 environments assuming that sufficient nutrients are present.

416 Microbial communities are distinct in different locations and may respond
417 differently due to local conditions. Many of the oil degraders present in this model
418 community are representative of microbes shown to respond to oil spills throughout the
419 world (2). While this model community may not provide information about how a
420 specific location would respond to an oil spill, it does provide a system in which we can
421 reproducibly study the impact of various response strategies on a representative oil
422 degrading consortia.

423 Our results indicate distinct microbial community dynamics in response to oil as
424 well as other treatments. Microbial communities exposed to oil demonstrate a
425 coordinated dynamic with the initial responders typically being alkane degraders
426 followed by microbes specialized in degrading more recalcitrant compounds such as
427 aromatic compounds. Previous studies have indicated that environmental microbial
428 communities exhibit dynamics to degrade the preferred carbon source (42). The oil
429 degrading microbial communities in these enrichments exhibit similar dynamics with the

430 initial bloom of alkane degraders (*Marinobacter*, *Alcanivorax*, etc.) followed by an
431 increase in known aromatic hydrocarbon degraders (*Cycloclasticus*, *Alteromonadales*,
432 *Thalassospira*, etc.). Growth on the decaying bloom of oil degrading bacteria was
433 observed during the *DWH* spill (42), but was not observed in these enrichments..
434

435 Differential effects of COREXIT 9500 on community composition based on temperature

436 The primary goal of this study was to assess the effect of COREXIT 9500 on the
437 oil biodegradation and microbial community structure. Since the active community
438 should be the most diagnostic for the effect of COREXIT 9500, we will only discuss the
439 differences in the active community between treatments. PERMANOVA analysis
440 indicated that there were significant differences in the microbial community structure
441 between treatments at both temperatures. Pairwise PERMANOVAs were used to
442 determine which treatments were significantly different. At 25 °C, there was a
443 significant difference in the microbial community present in all three treatments (Table
444 1). However, at 5 °C, there were significant differences between the COREXIT 9500-
445 only conditions and the oil-containing conditions (oil alone and oil plus COREXIT
446 9500). There was no statistically significant difference between the oil-only and the oil
447 plus COREXIT 9500 treatments at 5 °C (PERMANOVA p-value: 0.367). These findings
448 suggest that at 5°C, the addition of COREXIT 9500 to oil-amended microcosms has a
449 negligible impact on the community structure compared to the structure in the oil-only
450 conditions. This would suggest that at cold temperatures the active community structure
451 is minimally impacted by the addition of COREXIT 9500. The observation that
452 COREXIT addition stimulated oil biodegradation at 5 °C despite the minimal impact on

453 the microbial community, may imply that the COREXIT increases the bioavailability of
454 oil for the natural oil degrading population. Alternate explanations for the impact of
455 COREXIT on community structure could be due to COREXIT serving as a carbon source
456 for microbes in these consortia. At 5°C there is little DOSS degradation. However, there
457 was substantial DOSS degradation at 25°C. This difference in DOSS degradation could
458 indicate that the distinct community structure in response to COREXIT could be due to
459 microbes consuming the COREXIT components.

460 COREXIT 9500 differentially impacts members of the same taxonomic grouping

461 To better understand which taxa were most affected by COREXIT 9500 at 25 °C,
462 differentially abundant taxa were identified between the only oil and the oil plus
463 COREXIT 9500 conditions. Forty operational taxonomic units (OTUs) were identified
464 as being differentially abundant in the oil plus COREXIT 9500 condition compared to the
465 oil-only condition at 25 °C (Table 2). No OTUs were identified as showing significantly
466 different abundances between the oil-only and oil plus COREXIT 9500 treatment at 5 °C,
467 which further supports the limited effect of COREXIT 9500 on these microbial consortia
468 at 5 °C. The differentially abundant OTUs were classified into eight orders representing
469 10 families. Of the 40 differentially abundant OTUs, 17 were more abundant in the oil
470 and COREXIT 9500 condition, whereas 23 were more abundant in the oil-only condition.
471 Five of the differentially abundant OTUs were assigned as an unclassified
472 *Gammaproteobacteria*. Three of the unclassified *Gammaproteobacterial* OTUs were
473 enriched in the oil plus COREXIT 9500 condition, whereas two of them were enriched in
474 the oil-only condition.

475 A previous study demonstrated that COREXIT 9500 addition resulted in
476 decreased viability of a *M. hydrocarbonoclasticus* isolate obtained from a Gulf of Mexico
477 beach (14). In the present study, *Marinobacter* comprised a large proportion of the active
478 community in the oil-amended conditions incubated at 25 °C. *Marinobacter* spp. were
479 present at an average relative abundance of 27% in the oil-only conditions and 17% in the
480 oil plus COREXIT 9500 condition. This result could be interpreted as an inhibitory
481 effect of COREXIT 9500 on *Marinobacter* species, however, changes in relative
482 abundance could be due to a variety of factors. One possible explanation for this change
483 in relative abundance is that there were less *Marinobacter* cells in the oil plus COREXIT
484 9500 condition. Alternatively, since these data were obtained from cDNA libraries, it
485 could be that *Marinobacter* were present, yet less active in the oil plus COREXIT 9500
486 conditions. A third possible explanation is that *Marinobacter* spp. remain at the same
487 level (either cells or activity) in both treatments, but other organisms increased in
488 abundance or activity in the oil plus COREXIT 9500 condition. This third scenario
489 would result in an apparent decrease in the relative abundance of *Marinobacter*. Further
490 complicating matters is the fact that the relative abundance measured was the combined
491 abundance of all of the OTUs classified as *Marinobacter*. Therefore, an examination of
492 difference of the relative abundance of specific OTUs would provide a more accurate
493 estimation of the effect of COREXIT 9500 between treatments.

494 Eighteen *Marinobacter* OTUs were identified as being differentially abundant
495 between treatments. Eight of these *Marinobacter* OTUs showed enrichment in the oil
496 plus COREXIT 9500 condition, and the other ten were enriched in the oil-only condition
497 relative to the oil plus COREXIT 9500 condition. This would indicate that, while there

498 were certain *Marinobacter* OTUs that may be sensitive to the addition of COREXIT
499 9500, many of the *Marinobacter* OTUs were stimulated by the addition of COREXIT
500 9500. For example, one OTU was on average present at 0.3% and 0.4% in the COREXIT
501 9500 only and the oil-only samples, respectively. However, in the presence of both oil
502 and COREXIT 9500 this OTU increased to 2.2%, suggesting that this specific
503 *Marinobacter* population was stimulated by the addition of COREXIT 9500 when oil is
504 present. This potential stimulation would be in contrast to a previous report, which found
505 inhibition of an *M. hydrocarbonclasticus* isolate with the addition of COREXIT 9500
506 (14). This discrepancy could be due to the high-levels of COREXIT 9500 used in the *M.*
507 *hydrocarbonoclasticus* study. In this present study, the concentration of COREXIT 9500
508 was 1.12 µg/L and may not be inhibitory to all *Marinobacter* species.

509 There were some examples of *Marinobacter* OTUs that exhibited the opposite
510 trend. For example, one OTU was present at 0.7% of cDNA libraries with COREXIT
511 9500 alone. In the samples with oil, this OTU increased to 21% of the recovered
512 sequences. When oil and COREXIT 9500 are present, the abundance of this OTU was
513 only 10% of the library. Thus, there are fewer sequences recovered in the oil and
514 COREXIT 9500 treatment compared to the oil alone. This two-fold decrease in
515 recovered sequences could be indicative of an inhibitory effect of COREXIT 9500
516 addition on this OTU. Similar phenomena were observed with many of the other
517 differentially abundant OTUs, with some members of the same genus showing higher
518 abundance in the oil-only versus oil and COREXIT 9500 condition and visa versa.
519 However, there were some differentially abundant taxa that were only enriched in one
520 condition. For example, three OTUs, classified as *Alcanivorax* spp., were enriched in the

521 only oil condition compared to the oil plus COREXIT 9500 condition. One of the
522 differentially abundant *Alcanivorax* OTUs was the dominant *Alcanivorax* OTU found in
523 these microcosms. At 25°C, this OTU comprised on average 6.7% of the oil-only cDNA
524 communities and 3.5% of the oil plus COREXIT 9500 communities. This statistically
525 significant difference suggests that in the COREXIT 9500-amended microcosms, this
526 *Alcanivorax* sp. was not as competitive. Interestingly, in the 5°C incubation, this same
527 OTU comprised on average 56.8% and 59.5% of the active community in the oil-only
528 and the oil plus COREXIT 9500 microcosms, respectively. This suggests that at 25°C
529 there is a possibility that this OTU is inhibited by COREXIT 9500; in contrast, at 5°C the
530 addition of COREXIT 9500 has no effect. Therefore, the effect of COREXIT 9500
531 inhibition may be species-specific and dependent upon temperature. This is in agreement
532 with recent studies that demonstrated that COREXIT 9500 did not affect the growth of *A.*
533 *borkumensis* (Liu and Hazen, unpublished results).

534

535 **Conclusions**

536 The goal of this study was to provide additional insights into the effects of COREXIT
537 9500 on biodegradation of ANS crude oil as well its effects on the community structure
538 of oil-degrading microbes. These results combine to show that biodegradation is
539 enhanced by the addition of COREXIT 9500. Our results indicate that microbial
540 community structure is only impacted by COREXIT 9500 addition at 25 °C and not at 5
541 °C. The effect of COREXIT 9500 addition at 25 °C is stronger in the active community.
542 We have demonstrated that while there were some OTUs negatively impacted by the
543 addition of COREXIT 9500, a similar number of differentially abundant OTUs were

544 stimulated by the addition of COREXIT 9500. Our results suggest that the effect of
545 COREXIT 9500 is somewhat species specific and even within a species, there can be
546 differing effects depending on the individual strain/organism. Multiple species have the
547 ability to degrade single classes of hydrocarbons. Therefore, if one species is inhibited
548 by COREXIT 9500 addition, another may grow in its place. This study confirms
549 previous studies that have shown a positive impact of dispersants on rates of oil
550 biodegradation. While these results demonstrate that the microbial community is
551 impacted by COREXIT addition, it appears to be a species-specific phenomenon.

552

553

554

555 **Acknowledgements**

556 We are grateful to Devi Sundaravadivelu and Ruta Deshpande for critical discussions.
557 The U.S. Environmental Protection Agency, through its Office of Research and
558 Development, provided funds and collaborated in the research described herein. This
559 work has been subjected to the agency's administrative review and has been approved for
560 external publication. Any opinions expressed in this paper are those of the authors and do
561 not necessarily reflect the views of the agency; therefore, no official endorsement should
562 be inferred. Any mention of trade names or commercial products does not constitute
563 endorsement or recommendation for use.

564

565

566 **References**

- 567 1. **Head IM, Jones DM, Roling WFM.** 2006. Marine microorganisms make a
568 meal of oil. *Nature Reviews Microbiology* **4**:173-182.
- 569 2. **Hazen TC, Prince RC, Mahmoudi N.** 2016. Marine Oil Biodegradation.
570 *Environmental Science & Technology* **50**:2121-2129.
- 571 3. **Kvenvolden KA, Cooper CK.** 2003. Natural seepage of crude oil into the
572 marine environment. *Geo-Marine Letters* **23**:140-146.
- 573 4. **Li H, Boufadel MC.** 2010. Long-term persistence of oil from the Exxon Valdez
574 spill in two-layer beaches. *Nature Geoscience* **3**:96-99.
- 575 5. **Bowen L, Miles A, Ballachey B, Bodkin J, Esler D.** 2015. Gulf Watch Alaska
576 Long-term Monitoring Program-Evaluating Chronic Exposure of Harlequin
577 Ducks and Sea Otters to Lingering Exxon Valdez Oil in Western Prince
578 William Sound. Exxon Valdez Oil Spill Restoration Project Final Report
579 (Restoration Project 12120114-Q), Pacific Wildlife Foundation and Centre for
580 Wildlife Ecology, Simon Fraser University, Delta, British Columbia, Canada.
581 US Geological Survey, Alaska Science Center, Anchorage, Alaska:7.
- 582 6. **Atlas RM, Hazen TC.** 2011. Oil Biodegradation and Bioremediation: A Tale of
583 the Two Worst Spills in US History. *Environmental Science & Technology*
584 **45**:6709-6715.
- 585 7. **Brakstad OG, Nordtug T, Throne-Holst M.** 2015. Biodegradation of
586 dispersed Macondo oil in seawater at low temperature and different oil
587 droplet sizes. *Marine Pollution Bulletin* **93**:144-152.
- 588 8. **National Commission on the BP Deepwater Horizon Oil Spill and**
589 **Offshore Drilling.** 2011. The Use of Surface and Subsea Dispersants During
590 the BP Deepwater Horizon Oil Spill.
- 591 9. **Baelum J, Borglin S, Chakraborty R, Fortney JL, Lamendella R, Mason**
592 **OU, Auer M, Zemla M, Bill M, Conrad ME, Malfatti SA, Tringe SG, Holman**
593 **HY, Hazen TC, Jansson JK.** 2012. Deep-sea bacteria enriched by oil and
594 dispersant from the Deepwater Horizon spill. *Environmental Microbiology*
595 **14**:2405-2416.
- 596 10. **McFarlin KM, Prince RC, Perkins R, Leigh MB.** 2014. Biodegradation of
597 Dispersed Oil in Arctic Seawater at -1 degrees C. *Plos One* **9**.
- 598 11. **Zahed MA, Aziz HA, Isa MH, Mohajeri L.** 2010. Effect of Initial Oil
599 Concentration and Dispersant on Crude Oil Biodegradation in Contaminated
600 Seawater. *Bulletin of Environmental Contamination and Toxicology* **84**:438-
601 442.
- 602 12. **Prince RC, McFarlin KM, Butler JD, Febbo EJ, Wang FCY, Nedwed TJ.**
603 2013. The primary biodegradation of dispersed crude oil in the sea.
604 *Chemosphere* **90**:521-526.
- 605 13. **Lindstrom JE, Braddock JF.** 2002. Biodegradation of petroleum
606 hydrocarbons at low temperature in the presence of the dispersant Corexit
607 9500. *Marine Pollution Bulletin* **44**:739-747.
- 608 14. **Hamdan LJ, Fulmer PA.** 2011. Effects of COREXIT (R) EC9500A on bacteria
609 from a beach oiled by the Deepwater Horizon spill. *Aquatic Microbial Ecology*
610 **63**:101-109.

- 611 15. **Kleindienst S, Seidel M, Ziervogel K, Grim S, Loftis K, Harrison S, Malkin**
612 **SY, Perkins MJ, Field J, Sogin ML, Dittmar T, Passow U, Medeiros PM,**
613 **Joye SB.** 2015. Chemical dispersants can suppress the activity of natural oil-
614 degrading microorganisms. *Proceedings of the National Academy of Sciences*
615 *of the United States of America* **112**:14900-14905.
- 616 16. **National Research Council.** 2005. Oil spill dispersants: efficacy and effects.
617 The National Academies Press, Washington DC.
- 618 17. **Campo P, Venosa AD, Suidan MT.** 2013. Biodegradability of Corexit 9500
619 and Dispersed South Louisiana Crude Oil at 5 and 25 degrees C.
620 *Environmental Science & Technology* **47**:1960-1967.
- 621 18. **Belore RC, Trudel K, Mullin JV, Guarino A.** 2009. Large-scale cold water
622 dispersant effectiveness experiments with Alaskan crude oils and Corexit
623 9500 and 9527 dispersants. *Marine Pollution Bulletin* **58**:118-128.
- 624 19. **Zhuang M, Abulikemu G, Campo P, Platten WE, Suidan MT, Venosa AD,**
625 **Conmy RN.** 2016. Effect of dispersants on the biodegradation of South
626 Louisiana crude oil at 5 and 25° C. *Chemosphere* **144**:767-774.
- 627 20. **Spotte S, Adams G, Bubucis PM.** 1984. GP2 medium is an artificial seawater
628 for culture or maintenance of marine organisms. *Zoo Biology* **3**:229-240.
- 629 21. **Venosa AD, Suidan MT, King D, Wrenn BA.** 1997. Use of hopane as a
630 conservative biomarker for monitoring the bioremediation effectiveness of
631 crude oil contaminating a sandy beach. *Journal of Industrial Microbiology &*
632 *Biotechnology* **18**:131-139.
- 633 22. **Kapoor V, Li X, Elk M, Chandran K, Impellitteri CA, Domingo JWS.** 2015.
634 Impact of Heavy Metals on Transcriptional and Physiological Activity of
635 Nitrifying Bacteria. *Environmental Science & Technology* **49**:13454-13462.
- 636 23. **Caporaso JG, Lauber CL, Walters WA, Berg-Lyons D, Huntley J, Fierer N,**
637 **Owens SM, Betley J, Fraser L, Bauer M, Gormley N, Gilbert JA, Smith G,**
638 **Knight R.** 2012. Ultra-high-throughput microbial community analysis on the
639 Illumina HiSeq and MiSeq platforms. *Isme Journal* **6**:1621-1624.
- 640 24. **Caporaso JG, Kuczynski J, Stombaugh J, Bittinger K, Bushman FD,**
641 **Costello EK, Fierer N, Pena AG, Goodrich JK, Gordon JI, Huttley GA,**
642 **Kelley ST, Knights D, Koenig JE, Ley RE, Lozupone CA, McDonald D,**
643 **Muegge BD, Pirrung M, Reeder J, Sevinsky JR, Tumbaugh PJ, Walters**
644 **WA, Widmann J, Yatsunenko T, Zaneveld J, Knight R.** 2010. QIIME allows
645 analysis of high-throughput community sequencing data. *Nature Methods*
646 **7**:335-336.
- 647 25. **Aronesty E.** 2011. ea-utils : "Command-line tools for processing biological
648 sequencing data",
- 649 26. **Edgar RC.** 2010. Search and clustering orders of magnitude faster than
650 BLAST. *Bioinformatics* **26**:2460-2461.
- 651 27. **Edgar RC, Haas BJ, Clemente JC, Quince C, Knight R.** 2011. UCHIME
652 improves sensitivity and speed of chimera detection. *Bioinformatics*
653 **27**:2194-2200.
- 654 28. **Caporaso JG, Bittinger K, Bushman FD, DeSantis TZ, Andersen GL, Knight**
655 **R.** 2010. PyNAST: a flexible tool for aligning sequences to a template
656 alignment. *Bioinformatics* **26**:266-267.

- 657 29. **Wang Q, Garrity GM, Tiedje JM, Cole JR.** 2007. Naive Bayesian classifier for
658 rapid assignment of rRNA sequences into the new bacterial taxonomy.
659 *Applied and Environmental Microbiology* **73**:5261-5267.
- 660 30. **Bray JR, Curtis JT.** 1957. An Ordination of the Upland Forest Communities of
661 Southern Wisconsin. *Ecological Monographs* **27**:326-349.
- 662 31. **Lozupone C, Knight R.** 2005. UniFrac: a new phylogenetic method for
663 comparing microbial communities. *Applied and Environmental Microbiology*
664 **71**:8228-8235.
- 665 32. **Anderson MJ.** 2001. A new method for non-parametric multivariate analysis
666 of variance. *Austral Ecology* **26**:32-46.
- 667 33. **Oksanen J, Blanchet FG, Kindt R, Legendre P, Minchin PR, O'Hara RB,**
668 **Simpson GL, Solymos P, Stevens MHH, Wagner H.** 2013. vegan:
669 Community Ecology Package. R package version 2.0-10 [http://CRAN-](http://CRAN.R-project.org/package=vegan)
670 [project.org/package=vegan](http://CRAN.R-project.org/package=vegan).
- 671 34. **R Core Team.** 2013. R: A Language and Environment for Statistical
672 Computing. R Foundation for Statistical Computing Vienna, Austria
673 [http://wwwR-project.org/](http://www.R-project.org/).
- 674 35. **Paulson JN, Stine OC, Bravo HC, Pop M.** 2013. Differential abundance
675 analysis for microbial marker-gene surveys. *Nature Methods* **10**:1200-+.
- 676 36. **Douglas GS, McCarthy KJ, Dahlen DT, Seavey JA, Steinhauer WG, Prince**
677 **RC, Elmendorf DL.** 1992. The use of hydrocarbon analyses for
678 environmental assessment and remediation. *Journal of Soil Contamination*
679 **1**:197-216.
- 680 37. **Venosa AD, Holder EL.** 2007. Biodegradability of dispersed crude oil at two
681 different temperatures. *Marine Pollution Bulletin* **54**:545-553.
- 682 38. **Kujawinski EB, Soule MCK, Valentine DL, Boysen AK, Longnecker K,**
683 **Redmond MC.** 2011. Fate of Dispersants Associated with the Deepwater
684 Horizon Oil Spill. *Environmental Science & Technology* **45**:1298-1306.
- 685 39. **White HK, Lyons SL, Harrison SJ, Findley DM, Liu Y, Kujawinski EB.** 2014.
686 Long-term persistence of dispersants following the Deepwater Horizon oil
687 spill. *Environmental Science & Technology Letters* **1**:295-299.
- 688 40. **Redmond MC, Valentine DL.** 2012. Natural gas and temperature structured
689 a microbial community response to the Deepwater Horizon oil spill.
690 *Proceedings of the National Academy of Sciences of the United States of*
691 *America* **109**:20292-20297.
- 692 41. **Chakraborty R, Borglin SE, Dubinsky EA, Andersen GL, Hazen TC.** 2012.
693 Microbial response to the MC-252 oil and Corexit 9500 in the Gulf of Mexico.
694 *Frontiers in Microbiology* **3**.
- 695 42. **Dubinsky EA, Conrad ME, Chakraborty R, Bill M, Borglin SE, Hollibaugh**
696 **JT, Mason OU, Piceno YM, Reid FC, Stringfellow WT, Tom LM, Hazen TC,**
697 **Andersen GL.** 2013. Succession of Hydrocarbon-Degrading Bacteria in the
698 Aftermath of the Deepwater Horizon Oil Spill in the Gulf of Mexico.
699 *Environmental Science & Technology* **47**:10860-10867.
- 700 43. **Gauthier MJ, Lafay B, Christen R, Fernandez L, Acquaviva M, Bonin P,**
701 **Bertrand JC.** 1992. *Marinobacter-Hydrocarbonoclasticus* Gen-Nov, Sp-Nov, a

- 702 New, Extremely Halotolerant, Hydrocarbon-Degrading Marine Bacterium.
703 International Journal of Systematic Bacteriology **42**:568-576.
- 704 44. **Gutierrez T, Singleton DR, Berry D, Yang TT, Aitken MD, Teske A.** 2013.
705 Hydrocarbon-degrading bacteria enriched by the Deepwater Horizon oil spill
706 identified by cultivation and DNA-SIP. *Isme Journal* **7**:2091-2104.
- 707 45. **Lamendella R, Strutt S, Borglin S, Chakraborty R, Tas N, Mason OU,**
708 **Hultman J, Prestat E, Hazen TC, Jansson JK.** 2014. Assessment of the
709 Deepwater Horizon oil spill impact on Gulf coast microbial communities.
710 *Front Microbiol* **5**:130.
- 711 46. **Kostka JE, Prakash O, Overholt WA, Green SJ, Freyer G, Canion A,**
712 **Delgardio J, Norton N, Hazen TC, Huettel M.** 2011. Hydrocarbon-Degrading
713 Bacteria and the Bacterial Community Response in Gulf of Mexico Beach
714 Sands Impacted by the Deepwater Horizon Oil Spill. *Applied and*
715 *Environmental Microbiology* **77**:7962-7974.
- 716 47. **Grimes DJ, Johnson CN, Dillon KS, Flowers AR, Noriega NF, Berutti T.**
717 2009. What Genomic Sequence Information Has Revealed About *Vibrio*
718 Ecology in the Ocean-A Review. *Microbial Ecology* **58**:447-460.
- 719 48. **Tao Z, Bullard S, Arias C.** 2011. High Numbers of *Vibrio vulnificus* in Tar
720 Balls Collected from Oiled Areas of the North-Central Gulf of Mexico
721 Following the 2010 BP Deepwater Horizon Oil Spill. *Ecohealth* **8**:507-511.
- 722 49. **Smith CB, Johnson CN, King GM.** 2012. Assessment of polyaromatic
723 hydrocarbon degradation by potentially pathogenic environmental *Vibrio*
724 *parahaemolyticus* isolates from coastal Louisiana, USA. *Marine Pollution*
725 *Bulletin* **64**:138-143.
- 726 50. **Yakimov MM, Giuliano L, Gentile G, Crisafi E, Chernikova TN, Abraham**
727 **WR, Lunsdorf H, Timmis KN, Golyshin PN.** 2003. *Oleispira antarctica* gen.
728 nov., sp nov., a novel hydrocarbonoclastic marine bacterium isolated from
729 Antarctic coastal sea water. *International Journal of Systematic and*
730 *Evolutionary Microbiology* **53**:779-785.
- 731 51. **Mason OU, Hazen TC, Borglin S, Chain PSG, Dubinsky EA, Fortney JL, Han**
732 **J, Holman HYN, Hultman J, Lamendella R, Mackelprang R, Malfatti S,**
733 **Tom LM, Tringe SG, Woyke T, Zhou JH, Rubin EM, Jansson JK.** 2012.
734 Metagenome, metatranscriptome and single-cell sequencing reveal microbial
735 response to Deepwater Horizon oil spill. *Isme Journal* **6**:1715-1727.
- 736 52. **Mason O, Han J, Woyke T, Jansson J.** 2014. Single-cell genomics reveals
737 features of a *Colwellia* species that was dominant during the Deepwater
738 Horizon oil spill. *Frontiers in Microbiology* **5**.
- 739 53. **Bagi A, Pampanin DM, Brakstad OG, Kommedal R.** 2013. Estimation of
740 hydrocarbon biodegradation rates in marine environments: A critical review
741 of the Q 10 approach. *Marine environmental research* **89**:83-90.
- 742 54. **Brakstad OG, Nonstad I, Faksness LG, Brandvik PJ.** 2008. Responses of
743 microbial communities in Arctic sea ice after contamination by crude
744 petroleum oil. *Microbial Ecology* **55**:540-552.
- 745 55. **Giudice AL, Bruni V, Domenico DM, Michaud L.** 2010. Psychrophiles -
746 Cold-Adapted Hydrocarbon-Degrading Microorganisms, p 1897-1921. *In*
747 Timmis KN (ed), *Handbook of Hydrocarbon and Lipid Microbiology*

748 doi:10.1007/978-3-540-77587-4_139. Springer Berlin Heidelberg, Berlin,
749 Heidelberg.
750

751 **Figure Legends**

752
753 Figure 1: Oil Biodegradation. (A) Degradation of DOSS in the presence and absence of ANS oil. The left side of panel A shows
754 degradation of DOSS in the presence of ANS. The right side of panel A shows DOSS degradation in the absence of ANS. (B-D)
755 Comparison of biodegradation of ANS crude oil in the presence (left) and absence (right) of COREXIT 9500 at 25°C (open symbols)
756 and 5°C (filled symbols). (B) Biodegradation of hopane-normalized total alkanes. (C) biodegradation of hopane-normalized total
757 PAHs (D) biodegradation of *n*-alkanes (nC30 – nC35).
758

759 **Figure 2 Microbial Community Dynamics.** Analysis of the microbial community structure at the total (DNA – Left) and active
760 (cDNA- right) levels for the 25°C (top) and 5°C (bottom) microcosms. Taxa are shown grouped at the genus level. Sequences from
761 replicate microcosms were pooled and shown as a single bar for each time point and condition.
762

763 **Figure 3: Comparison of microbial community structure.** Non-metric multidimensional scaling of weighted unifrac distances.
764 Samples with only COREXIT 9500 were colored in gray, samples with oil-only were colored in orange, and samples with oil plus
765 COREXIT 9500 were colored in blue. Comparison of the total microbial community composition in the 5°C microcosm (A) and the
766 25°C microcosm (C). The microbial community composition was compared in the active community of the 5°C microcosm (B) and
767 the 25°C microcosm (D) Stress for the configuration of the different plots are 0.0622 (A), 0.0681 (B), 0.10604 (C), and 0.1997 (D).
768

Table 1: Pairwise PERMANOVAs of weighted unifrac distances

Temp.	Nucl. Acid	Comparison	pvalue	F stat	R2
5°C	DNA	COREXIT v. Oil+COREXIT	0.001	18.78	0.2446
		Oil v. Oil+COREXIT	0.012	3.9657	0.06613
		COREXIT v Oil	0.001	44.231	0.43266
	cDNA	COREXIT v. Oil+COREXIT	0.001	78.728	0.56344
		Oil v. Oil+COREXIT	0.367	1.0117	0.01714
		COREXIT v Oil	0.001	89.818	0.59554
25°C	DNA	COREXIT v. Oil+COREXIT	0.004	4.1372	0.06996
		Oil v. Oil+COREXIT	0.047	2.6213	0.04799
		COREXIT v Oil	0.001	9.684	0.14971
	cDNA	COREXIT v. Oil+COREXIT	0.001	6.5336	0.10691
		Oil v. Oil+COREXIT	0.001	11.935	0.18668
		COREXIT v Oil	0.001	20.136	0.26799

Table 2: Differentially abundant taxa between samples.

Family	Genus	Number of OTUs enriched in oil-only	Number of OTUs enriched in oil plus COREXIT 9500
<i>Nocardiaceae</i>	<i>Rhodococcus</i>	0	1
<i>Phycisphaerales</i>	Unclassified	0	1
<i>Rhodobacterales</i>	<i>Hyphomondadaceae</i>	2	0
<i>Rhodobacterales</i>	<i>Rhodobacteraceae</i>	3	0
<i>Sphingomonadlaes</i>	<i>Erythrobacteraceae</i>	0	1
<i>Gammaproteobacteria</i>	Unclassified	2	3
<i>Alteromonadaceae</i>	<i>Marinobacter</i>	10	8
<i>Idiomarinaceae</i>	<i>Pseudidiomarina</i>	1	1
<i>Alcanovoraceae</i>	<i>Alcanovorax</i>	0	3
<i>Vibrionaceae</i>	Unclassified	3	1
	Total	21	19

Figure 1

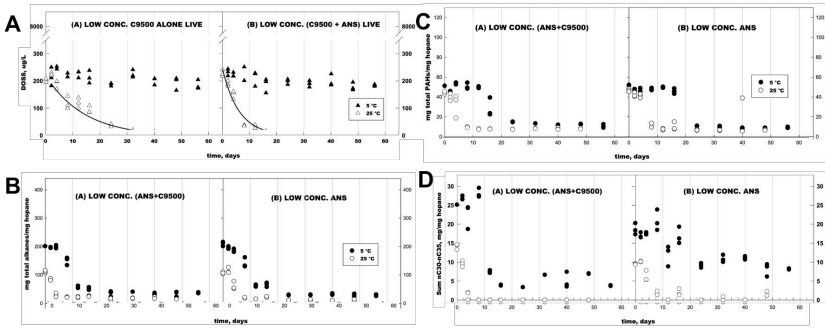


Figure 2

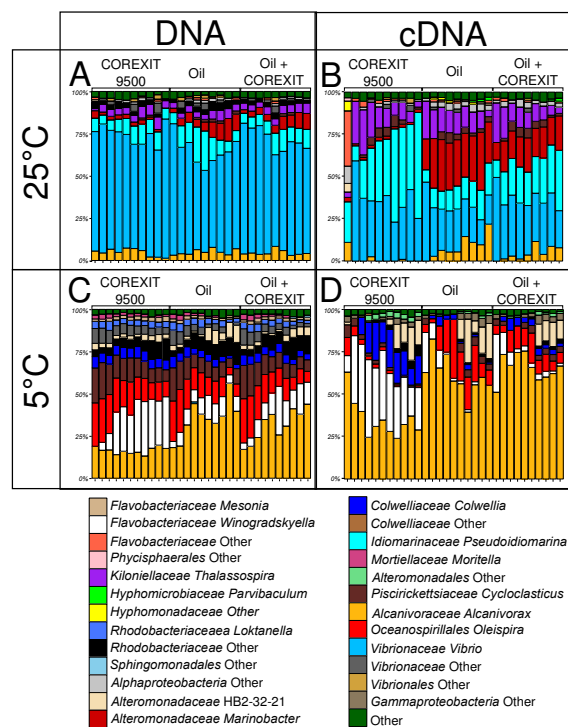
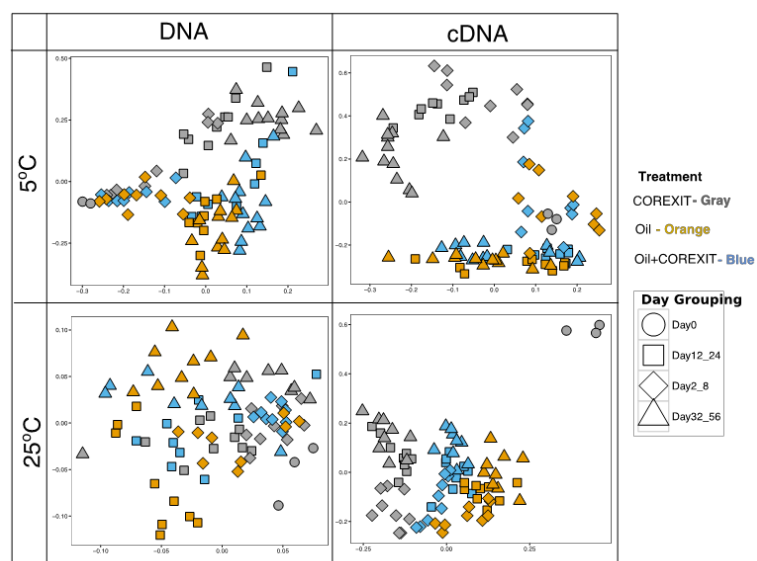


Figure 3





DSPACE

<https://dspace.org/>

COREXIT 9500 enhances oil biodegradation and changes microbial community structure of oil-enriched microcosms

Techtmann, Stephen M.; Zhuang, Mobing; Campo Moreno, Pablo; Holder, Edith; Elk, Michael ...

2017-03-10

American Society for Microbiology

<https://doi.org/10.1128/AEM.03462-16>

Techtmann SM, Zhuang M, Campo P, et al., (2017) COREXIT 9500 enhances oil biodegradation and changes microbial community structure of oil-enriched microcosms, Applied and Environmental Microbiology, Volume 83, Issue 10, May 2017, Article number e03462-16